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4 **Upgrading municipal lagoons in temperate and**
5 **cold climates: Total nitrogen removal and phosphorus**
6 **assimilation at ultra-low temperatures**

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28 **Abstract**

29 In this study, a municipal lagoon with high wintertime effluent total ammonia
30 nitrogen (TAN) concentrations was upgraded with a pilot-scale nitrifying-nitrifying-
31 denitrifying (NIT-NIT-DENIT) moving bed biofilm reactor (MBBR) treatment train to
32 characterize its effluent over wintertime operation, investigate the feasibility of upgrading
33 lagoons to achieve substantial biological total nitrogen removal across ultra-low
34 temperatures (0.6 – 3.0°C) and investigate nitrification inhibition pathways in facultative
35 lagoon systems at ultra-low temperatures. Throughout the study, it was observed that the
36 system substantially reduced total nitrogen (TN) and total phosphorus (TP) effluent
37 concentrations by an average of $69.8 \pm 24.5\%$ and $74.7 \pm 20.1\%$, respectively.
38 Furthermore, it was observed that sulfide toxicity may play an important role in the
39 inhibition of nitrifying organisms in lagoons. Finally, the MBBR treatment technology has
40 emerged as a suitable and sustainable upgrade technology for existing lagoon and waste
41 stabilization pond facilities operating in temperate, northern, and cold climate countries.

42

43 Keywords: wastewater treatment, water quality, pollution, seasonal, environmental
44 management

45 **1. Introduction**

46 The impoverishment of surface waters is a global problem that impact many
47 communities around the world. Anthropogenic release of nutrients to natural waters is
48 widely recognized as a leading cause of eutrophication and water toxicity (Smith et al.,
49 1999; Lewis et al., 2011). Furthermore, it has been recognized that the effluent discharge
50 of water resource recovery facilities (WRRF) is a major source of these anthropogenic
51 nutrients (nitrogen & phosphorus) that are released to the natural environment (Preston
52 et al., 2011). As a result of the impact of wastewater discharge, several countries and
53 states have imposed stringent effluent discharge regulations for total nitrogen (TN) and
54 total phosphorus (TP) (Directive Council of the European Union, 1991, 1996; Ministry of
55 Ecology and Environment of the People’s Republic of China, 1996; USEPA, 2002; MOE
56 Ontario, 2008).

57 In many countries, waste stabilization ponds, also named lagoons, remain one of
58 the most common types of WRRFs. Currently, over 8,000, 3,000, 2,500 and 1,200
59 lagoons are in operation in the U.S., Germany, France and Canada, respectively. (Mara,
60 2009; USEPA, 2011; Statistics Canada, 2016). Lagoons typically consist of single or
61 multi-celled basins that hold wastewater for extended periods (7 to 30+ days) (USEPA,
62 2000; Delatolla and Babarutsi, 2005; Bruce, 2009). They can be operated in non-aerated
63 (facultative) or aerated configurations, and lagoon-type WRRF installations often combine
64 several types of cells to achieve the desired treatment level. Lagoons are designed to
65 reduce the concentration of total suspended solids (TSS), carbonaceous biochemical
66 oxygen demand (CBOD) and, at some installations, total ammonia nitrogen (TAN), prior

67 to their discharge to receiving waters (Asano et al., 2007). Due to their simplicity and low
68 operational intensity, lagoons remain one of the preferred methods of treating municipal
69 wastewaters in smaller communities where general land availability allows for their
70 construction (Mittal, 2006; Muga and Mihelcic, 2008).

71 Despite the extensive installation and use of lagoons in many countries, these
72 systems are subject to several drawbacks and limitations. The nutrient treatment removal
73 capabilities of lagoons situated in northern climates often becomes significantly limited
74 due to temperature-induced decreases in the metabolic activity of the microbial
75 populations (Andreottola et al., 2000; Wessman and Johnson, 2006; Delatolla et al., 2009,
76 2010, 2012; Hoang, 2013; Hoang et al., 2014; Ragush et al., 2015; Young et al., 2016a).
77 During fall and winter in temperate and northern climates, wastewater temperatures in
78 lagoons can often fall below 1.0°C (Heaven et al., 2003; Krkosek et al., 2012; Ahmed et
79 al., 2019). Cold-temperature driven loss of bacterially-mediated nitrification in lagoons
80 located in northern climates has often been shown to cause effluent TAN concentrations
81 to substantially increase (Painter and Loveless, 1983), leading to the harmful release of
82 ammonia to receiving waters (Painter and Loveless, 1983). Additionally, removal of
83 phosphorus in lagoons is typically performed via the chemical addition of coagulants or
84 lime (Narasiah et al., 1994), as few naturally occurring biological pathways exist within
85 these systems that are capable of effectively reducing the release of phosphorus. Limited
86 overall performance and specifically limited nutrient removal during low temperature
87 operation has created a need to upgrade these systems to maintain compliance with
88 increasingly stringent nutrient regulations, and specifically to remove nitrogenous and
89 phosphorous compounds prior to discharge.

90 Treatment technologies designed to upgrade lagoons should be appropriate in
91 their design to require similar, low level operational intensity; as lagoon facilities often do
92 not have full-time operators always present. Several technologies have been employed
93 to upgrade lagoons; such as extended aeration of aerated lagoons (Melcer et al., 1995),
94 trickling filters (Archer and O'Brien, 2005; Avsar et al., 2008), rotating biological
95 contactors (RBC) (Hassard et al., 2015), and constructed wetlands (Cameron et al., 2003;
96 Butterworth et al., 2016). In recent years, studies have outlined the use of rock or
97 aggregate-based attached growth systems (Swanson and Williamson, 1980; Mara and
98 Johnson, 2006; Mattson et al., 2018), stationary in-lagoon fixed film media (Shin and
99 Polprasert, 1988; Srinivas, 2007; Gan and Champagne, 2015) and moving bed biofilm
100 reactor (MBBR) systems (Wessman and Johnson, 2006; Delatolla et al., 2010; Hoang,
101 2013) to upgrade lagoons either by enhancing microbially-mediated nitrification or by
102 increasing the lagoon system's volumetric or loading capacity. Numerous studies have
103 recently demonstrated the effectiveness of nitrifying attached growth technologies such
104 as the MBBR to enhance TAN-removal performance at the end of lagoons to meet
105 stringent ammonia effluent guidelines at ultra-low (0.6 – 3.0°C) temperatures (Delatolla
106 et al., 2010; Hoang, 2013; Almomani et al., 2014; Young et al., 2016b; Ahmed et al., 2019;
107 Patry et al., 2019). Post-carbon nitrifying MBBR systems utilize plastic media with a high
108 specific surface area to encourage the adhesion and attachment of nitrifying bacterial
109 communities. The microbial populations in nitrifying MBBR systems have been shown to
110 be highly diversified (Ottawa et al., 2006), allowing nitrifying MBBR systems to operate in
111 a wide range of temperatures (1°C – 20+°C). Biologically active organisms in nitrifying
112 biofilms are ammonia oxidizing bacteria (AOB) and nitrite oxidizing bacteria (NOB)

113 (Gieseke et al., 2003). In locales where total nitrogen is regulated, nitrifying MBBR
114 systems can be supplemented with a follow-up denitrifying MBBR system to convert
115 produced nitrate to nitrogen gas, effectively removing nitrogen from the aqueous
116 environment. The denitrifying MBBR technology has been utilized for over 20 years
117 (Aspegren et al., 1998) however it is almost exclusively applied in larger, mechanical
118 plants, where mesophilic conditions exist ($10^{\circ}\text{C}+$). In the context of lagoon facility
119 upgrades and retrofits with denitrifying MBBR technology, current available literature does
120 not address operation below 5°C (Dale et al., 2015). As a result, a fundamental lack of
121 knowledge remains regarding the capacity of utilizing the MBBR technology to perform
122 total nitrogen removal (nitrification and denitrification) as an add-on technology at a
123 municipal lagoon in temperate and cold climates, where water temperatures below 1°C
124 may routinely be encountered during wintertime. In addition, there is a gap of knowledge
125 with respect to biologically mediated upgrade systems for lagoons to enhance total
126 phosphorus removal. Ensuring effective total nitrogen and total phosphorus removal in
127 lagoon treatment systems is necessary to ensure the perennality of water resources in
128 various countries (Brettle et al., 2016; Statistics Canada, 2018) and as such feasible and
129 appropriate nutrient removal upgrade technologies are urgently needed as low
130 operational intensity add-on technologies for lagoon WRRFs.

131 The aim of this study is to investigate the performance of a nitrifying and denitrifying
132 multi-reactor MBBR system installed as an upgrade system to facultative lagoons
133 operating at ultra-low temperatures ($0.6 - 3.0^{\circ}\text{C}$), quantify the removal efficiency of TAN
134 and NO_x within the multi-reactor MBBR treatment system at ultra-low temperatures, and
135 to understand the inhibition of nitrification within facultative lagoon systems during ultra-

136 low temperature operation. Furthermore, the study also investigates phosphorus
137 assimilation in a nitrifying-denitrifying upgrade MBBR system as an additional benefit to
138 the operation and as a potentially significant pathway for total phosphorus removal from
139 lagoon effluent.

140 **2. Materials and methods**

141 **2.1. Study site**

142 A pilot-scale wastewater treatment plant was installed at the Casselman, Ontario,
143 Canada ([45°19'26.3"N, 75°04'48.6"W](#)) municipal facultative lagoon WRRF from Dec.
144 2017 until Jun. 2018. The facultative lagoon is characterized by high effluent TAN
145 concentrations in wintertime due to seasonal temperature-driven loss of bacterial
146 nitrification. The lagoon system consists of three cells operated in-series. Cells #1 (7.4
147 ha) and #2 (7.2 ha) are operated as facultative cells, but cell #3 (4.1 ha) is equipped with
148 bottom-mounted aerators and is aerated prior to planned discharge to alleviate sulfides
149 and enhance effluent polishing (Figure 1). Cell #3 was not aerated from day 1-76. Pre-
150 planned aeration in cell #3 began on day 77 of the study, lasting until the end of April (day
151 125) due to required lagoon discharge. Spring discharge of the WRRF commenced on
152 day 116 of the study and lasted approximately 30 days. The lagoon is typically discharged
153 twice a year, in the fall and in the spring. The study was separated in three distinct periods
154 of comparable lengths: i) ice-forming period (days 1-50), ii) full ice-cover period (days 51-
155 115) and iii) spring thaw period (days 116-165).

156 **2.2. Pilot plant configuration**

157 The pilot plant housed an influent storage vessel and three MBBR reactors in
158 series. The pilot-scale treatment plant was fed with the effluent of the last cell of the three-
159 cell lagoon treatment system in Casselman, Ontario. The lagoon effluent was pumped
160 from the lagoon and into an influent storage vessel in the pilot via a 0.75 kW submersible

161 pump suspended by a buoy approximately 40 cm below the water surface (near to the
162 surface) providing flow to the MBBR treatment train. As such, the treatment train
163 consisted of an influent storage vessel (retention time of less than 10 minutes), followed
164 by three cylindrical 223 L MBBR reactors (total dimensions: 61 cm Ø x 122 cm H) in-
165 series; a nitrifying MBBR reactor followed by a second nitrifying MBBR reactor followed
166 by a denitrifying MBBR reactor, referred to in this manuscript as a NIT-NIT-DENIT
167 configuration (Figure 1). This choice of configuration (NIT-NIT-DENIT) was selected due
168 to concerns of potential sulfide toxicity to the nitrifying microbial communities, and due to
169 intrinsic improvements to nitrifying removal efficiency due to the process being rate-
170 limited at cold temperatures. Sizing of the reactors in the pilot unit was based on the
171 kinetics calculated at projected minimum temperatures, the total flowrate and the TAN
172 loading. Carrier movement in the first and second nitrifying MBBR reactors (R1 & R2)
173 were maintained by supplied aeration. The carriers in the denitrifying reactor (R3) were
174 maintained in motion with a mechanical mixer. The airflow in the reactor was adjusted
175 periodically to maintain adequate carrier motion throughout the experiment. The
176 mechanical mixer speed was also adjusted periodically to maintain adequate carrier
177 motion. The effluent of the pilot-scale treatment system was then recirculated to the first
178 cell of the lagoon treatment system. The carbon source dosed in the third, denitrifying
179 reactors was Hydrex™ 6860 supplied by Veolia Water Technologies (Montreal, Canada).
180 All MBBR carrier media utilized in this study were Anox™ K5 carriers (surface area to
181 volume ratio of 800m²/m³). All reactors in this study had a carrier fill percentage of 50%
182 and had an operational HRT of 4.0 hours, based on the design criteria of low temperature

183 nitrifying MBBR systems treating lagoon effluent (Young et al., 2017a; Ahmed et al.,
184 2019).

185 2.3. Pilot plant start-up

186 Prior to the beginning of the study, all three MBBR reactors in the pilot plant were
187 seeded with carriers harvested from an integrated fixed-film activated sludge (IFAS)
188 facility in Hawkesbury, ON, Canada, to promote rapid inoculation and growth of the
189 nitrifying community (Young et al., 2017b). The carriers were collected with 20 L HDPE
190 strainers and transported in drained 200 L barrels to the Casselman lagoon facility,
191 approximately 60 kilometres away, and loaded into the MBBR reactors. The three MBBR
192 reactors in the pilot treatment unit were operated in a continuous flow configuration for a

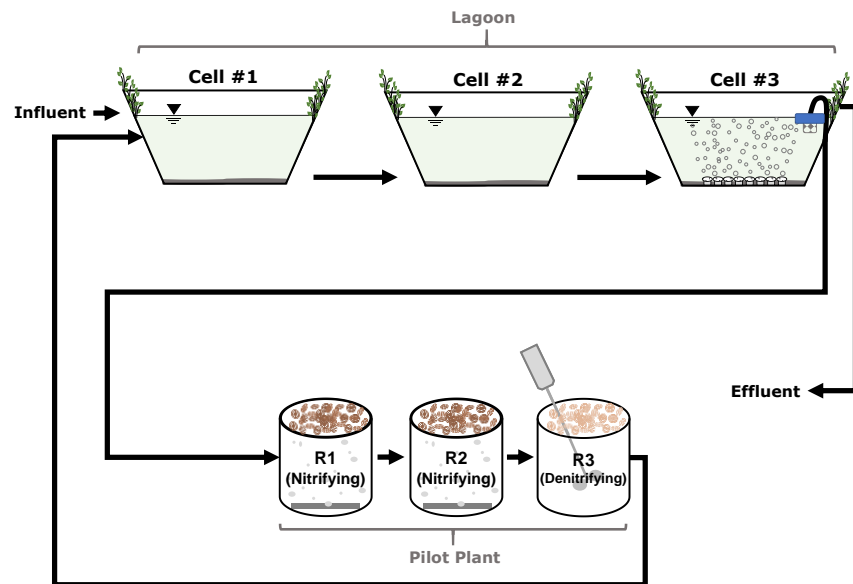


Figure 1: Schematic of the Casselman municipal facultative lagoon WRRF and MBBR nitrogen removal MBBR pilot systems.

193 period of one month prior to the start of the study to acclimatize the seeded carriers and
194 were fed with TAN-augmented lagoon effluent with the help of a peristaltic pump injecting

195 ammonium chloride solution into the first reactor in-series to stimulate the growth of
196 ammonia-oxidizing bacteria. The water quality parameters of the first reactor in the
197 treatment train during the acclimatization period were as follows: TAN \geq 25.0 mg-N/L,
198 BOD \leq 30.0 mg/L, DO \geq 5.0 mg/L. All three reactors demonstrated steady state operation
199 after one month of operation; with steady state defined as TAN and NO₃⁻ removal rate
200 fluctuations within a \pm 10% variation of the geometric mean.

201 **2.4. Water quality analyses**

202 Water samples were collected from the influent holding tank and each of the
203 reactors, and analyzed at the pilot facility every 2-5 days over a period of 165 days.
204 Samples which could not immediately be analysed were stored at 4°C for a maximum of
205 24 hours prior to analysis in the laboratory. The following constituents were analyzed:
206 biological oxygen demand (BOD) (5210 B) (APHA, WEF, 2012), chemical oxygen
207 demand (COD) (SM 5220 D) (APHA, WEF, 2012), total suspended solids (TSS) (SM
208 2540 D) (APHA, WEF, 2012), total ammonia nitrogen (HACH Method 8155) (Reardon et
209 al., 1966; Hach Company, 2015), nitrate (SM 4500-NO₃⁻ B) (APHA, WEF, 2012), nitrite
210 (SM 4500-NO₂⁻ B) (APHA, WEF, 2012), alkalinity (SM 2320 B) (APHA, WEF, 2012), total
211 sulfide (SM 4500-S²⁻D) (APHA, WEF, 2012), sulfate (US EPA 375.4) (U.S. Environmental
212 Protection Agency, 1978) and total phosphorus (SM 4500-P E) (APHA, WEF, 2012).
213 Dissolved oxygen (DO) and pH were measured using HACH LDO101 and PHC201
214 probes paired with a HACH HQ40d meter (Loveland, CO).

215 **2.5. Statistical analyses**

216 Statistical analyses were performed in order to establish statistical significance (p -
217 value and Pearson's R) between water quality parameters throughout the study, linear
218 regressions with a 95% confidence interval were performed between parameters of
219 interest. The assumptions associated with a linear regression model were validated.

220 **3. Results and discussion**

221 **3.1. Lagoon effluent characteristics**

222 Domestic wastewater entering the Casselman municipal lagoon flows through
223 cells #1 and #2, which are facultative, followed by cell #3, which is intermittently aerated
224 prior to seasonal discharge. The effluent of cell #3 fed the pilot treatment plant and was
225 monitored over a period of 165 days, which encompassed the seasonal spring discharge
226 period from the end of December to early June. The following water quality parameters
227 (COD, TSS, nitrite, alkalinity, sulfate, and pH) were separated in three different lagoon
228 operation periods: the ice-forming period (days 1-50), and the full ice-cover period (days
229 51-115) and the spring thaw period (days 116-165) (Table 1). BOD (data not shown) and
230 TSS (Table 1) concentrations met or exceeded regulatory standards (<25 mg/L cBOD₅
231 and <25 mg/L TSS) (Canadian Council of Ministers of the Environment, 2009) throughout
232 almost the entire study. During the snowmelt period, COD and alkalinity decreased
233 slightly (Table 1). It is possible that COD and alkalinity concentrations in the third cell of
234 the lagoon may have decreased slightly during the spring thaw period either due to a
235 restart of aerobic carbon removal and nitrifying activity, or potentially due to dilution. Due
236 to the location of the pump used to collect water from the lagoon (near the surface), some

237 biological processes and concentration gradients of different water quality constituents
 238 may not have been fully observed, particularly during the full ice-cover period.

239 Table 1. Average lagoon effluent concentrations of water quality parameters throughout the
 240 study period.

Parameter	Ice-forming period, Average \pm SD, (n)	Full ice-cover period, Average \pm SD, (n)	Spring thaw period, Average \pm SD, (n)
COD (mg DO/L)	59.2 \pm 5.6, (13)	51.1 \pm 9.1, (16)	32.6 \pm 5.0, (11)
TSS (mg/L)	13.6 \pm 3.2, (5)	11.6 \pm 10.7, (18)	12.4 \pm 11.7, (10)
NO₂⁻ (mg-N/L)	0.01 \pm 0.01, (13)	0.00 \pm 0.01, (17)	0.46 \pm 0.73, (11)
Alk. (mg CaCO₃/L)	315.3 \pm 16.3, (7)	299.0 \pm 41.6, (16)	203.1 \pm 60.0, (11)
SO₄²⁻ (mg-S/L)	40.7 \pm 5.6, (5)	42.6 \pm 6.2, (18)	36.9 \pm 2.9, (8)
pH	7.9 \pm 0.6, (12)	7.6 \pm 0.2, (19)	7.6 \pm 0.4, (10)

241
 242 Temperature, TAN, nitrate, total sulfides (TS), TP and DO values in the lagoon
 243 system demonstrated distinct concentrations during three distinct periods of operation
 244 (Table 2): i) ice-forming period (days 1-50), ii) full ice-cover period (days 51-115) and iii)
 245 spring thaw period (days 116-165). During the ice-forming period (Days 1-50), water
 246 temperatures in cell #3 were sufficiently low ($1.8 \pm 0.5^{\circ}\text{C}$) to limit benthic nitrification,
 247 leading to lagoon-wide increases in TAN concentrations (peaking at 18.41 mg-N/L) and
 248 low nitrate concentrations (max. of 2.09 mg-N/L). TAN concentrations in cell #3 during
 249 this first period increased from an initial concentration of 14.59 mg-N/L at day 1 of the
 250 study to 18.41 mg-N/L around day 50 of the study. Furthermore, the lack of observable
 251 nitrite build-up throughout the period is indicative that nitrification was likely not suppressed
 252 due to toxic effects.

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254
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Table 2. Average \pm SD of TAN, nitrate, TS and TP concentrations in cell #3 throughout the study.

<i>In-lagoon (Cell #3) water quality parameters</i>	<i>Ice-forming period, Average \pm SD, (n)</i>	<i>Full ice-cover period, Average \pm SD, (n)</i>	<i>Spring thaw period, Average \pm SD, (n)</i>
<i>Temperature ($^{\circ}$C)</i>	1.8 \pm 0.5, (12)	2.1 \pm 0.8, (20)	13.4 \pm 6.2, (11)
<i>TAN (mg-N/L)</i>	17.01 \pm 1.62, (14)	19.23 \pm 3.52, (18)	12.58 \pm 5.60, (11)
<i>NO₃⁻ (mg-N/L)</i>	1.32 \pm 0.62, (13)	0.33 \pm 0.25, (18)	0.42 \pm 0.42, (11)
<i>TS (mg-S/L)</i>	0.14 \pm 0.14, (4)	6.26 \pm 6.17 ^{α} , (10) / 0.07 \pm 0.08 ^{β} , (9)	0.04 \pm 0.04, (11)
<i>TP (mg-P/L)</i>	0.38 \pm 0.05, (4)	0.38 \pm 0.05, (4)	0.08 \pm 0.03, (11)
<i>Dissolved Oxygen (mg O₂/L)</i>	0.8 \pm 0.6, (13)	0.3 \pm 0.1 ^{α} , (7) / 9.0 \pm 5.5 ^{β} , (13)	8.9 \pm 4.4, (11)

α : concentration prior to start of aeration in cell #3; β : concentration after start of aeration in cell #3

256

257 As a result the observed data suggests limited nitrifying and denitrifying microbial
258 activity in the lagoon system in wintertime may occur due to a temperature-induced
259 (<4.0 $^{\circ}$ C) reduction in microbial activity and low-level sulfide toxicity in the benthos, which
260 is known to be inhibitory to the nitrifying and denitrifying microbial communities (Joye and
261 Hollibaugh, 1995; Senga et al., 2006; Bejarano Ortiz et al., 2013). Hypoxic conditions
262 prevailed throughout cell #3 across days 1 to 50. The low DO conditions (0.8 \pm 0.6 mg
263 O₂/L) were also potentially exacerbated by the production of hydrogen sulfide in the
264 benthic zone of the lagoon, causing a high sediment oxygen demand (Wang, 1980; Chen
265 et al., 2017; D'Aoust et al., 2018, Alqaralleh et al., 2019). However, TS concentrations in
266 the water column remained low during this period (0.14 \pm 0.14 mg-S/L). Meanwhile, TP
267 concentrations were stable and saw little fluctuations, with concentrations of 0.38 \pm 0.05
268 mg-P/L being observed during this period.

269 During the ice-cover period (days 51-115), ice and snow covered the entirety of
270 the lagoon and water temperatures averaged 2.1 \pm 0.8 $^{\circ}$ C. TAN concentrations further

271 increased during this period, reaching an average concentration of 19.23 ± 3.52 mg-N/L
272 and ultimately plateauing at 20.27 mg-N/L between days 94 to 102 (Supplemental Figure
273 1). Seasonal increases in TAN concentrations show strong correlation to decreases in
274 water temperature ($R=0.651$, $p<0.02$), highlighting the sensitivity of lagoon nitrifying
275 communities to the wastewater temperature (Van Dyke et al., 2003). Ice cover completely
276 inhibited aerobic conditions in the top portion of the lagoon water column (Macdonald et
277 al., 1991; Chen et al., 2017; D'Aoust et al., 2017),

278 The low DO concentrations of the lagoon during ice-cover and the lack of
279 reaeration processes in cell #3 during wintertime caused widespread anaerobic
280 processes to take hold. Specifically, sulfate-reduction in the lagoon led to an average TS
281 concentration of 6.94 ± 6.15 mg-S/L across the ice-cover period, peaking at 17.11 ± 5.66
282 mg-S/L (Supplemental Figure 1). As ice-cover forms and becomes covered by snow, light
283 and reaeration processes ceased, causing the death and degradation of most lagoon
284 macrophytes, which could potentially provide carbon to sustain sulfate-reduction. With
285 this onset of TS production, a rapid decrease in residual nitrate concentration (1.98 to
286 0.09 mg-N/L) was observed to occur simultaneously with increases in TS concentrations
287 (Supplemental Figure 1). This correlation between TS and nitrate concentration is
288 believed to be caused by sulfide toxicity to nitrifying microorganisms (Joye and
289 Hollibaugh, 1995; Senga et al., 2006; Bejarano Ortiz et al., 2013), and may be indicative
290 that sulfide toxicity in the benthic zone further limits nitrification in lagoons at low
291 temperatures, beyond effects caused by low temperatures alone. The identification of
292 sulfide benthic toxicity of nitrifiers in lagoons during low temperatures as a contributing
293 factor to loss of nitrification modifies conventional belief that nitrification in lagoons located

294 in northern or temperate climates ceases solely due to temperature inhibition of
295 nitrifying organisms at temperatures lower than 4°C (Sharma and Ahlert, 1977; Painter
296 and Loveless, 1983). As both nitrification and sulfate-reduction processes occur in the
297 benthos of lagoons, it is plausible that in-sediment sulfide exposure may be a secondary,
298 significant inhibitor of nitrification (Caffrey et al., 2019) in wastewater lagoons. Another
299 possible cause for decreases in nitrate could be due to potential autotrophic denitrification
300 performed by sulfate-reducing bacteria (Shao et al., 2010), but the authors believe this is
301 less likely due to relatively low nitrate concentrations. During this period, TP
302 concentrations did not fluctuate significantly, averaging a concentration of 0.32 ± 0.10
303 mg-P/L.

304 Hypoxic conditions in the lagoon continued from day 51 until day 77 (0.3 ± 0.0 mg
305 O₂/L), at which point DO concentrations increased significantly due to the planned lagoon
306 cell mechanical aeration. On day 77 of the study, aeration in cell #3 was activated to
307 polish the lagoon effluent in preparation for seasonal discharge to receiving waters during
308 the thaw period. The start of the aeration was governed by the partner municipality and
309 was required due to the limited allowed discharge period outlined in their Environmental
310 Compliance Approval (ECA). The mechanical aeration had the effect of stripping sulfides
311 out of the water column of cell #3. The average DO concentrations in the cell after aeration
312 commenced increasing slowly to 1.6 mg O₂/L by day 92. Average DO concentrations from
313 day 92 to day 115 were of 12.6 ± 0.6 mg O₂/L, outlining a lag period of approximately 15
314 days between the beginning of aeration and the cessation of widespread pond hypoxia,
315 possibly due to sulfidic oxygen demand in the lagoon masking immediate effects of

316 aeration on DO concentrations. Meanwhile, after stripping sulfides, TS concentrations
317 decreased significantly to 0.09 ± 0.09 mg/L.

318 Finally, during the spring thaw period (days 116-165), wastewater temperatures
319 increased rapidly from 1.3°C on day 116 to 10.3°C on day 123 and 15.1°C on day 130.
320 This rapid increase in wastewater temperature led to rapid dilution effects, causing TAN
321 concentrations to decrease to 9.00 mg-N/L. Nitrate concentrations during this period were
322 low ($<2\%$ of TN as NO_3^-) until temperatures reached 15°C and above, and subsequently
323 increased to 2.51 mg-N/L (31% of TN as NO_3^-). Meanwhile, TS concentrations ($0.04 \pm$
324 0.04 mg-S/L) remained suppressed due to sustained aeration in cell #3 of the lagoon. At
325 the same time, TP concentrations significantly decreased, with an average of 0.08 ± 0.03
326 mg-P/L in the water column of cell #3. The decrease in TP concentration is believed to
327 have occurred due to spring thaw period induced dilution and biochemical effects.

328 Spring thaw caused a reaeration of the lagoon water column due to a significant
329 increase in influent flow rates while simultaneously reallowing wind reaeration pathways
330 (surface oxygen transfer) to increase water column DO. This subsequently caused the
331 cessation of widespread hypoxic conditions in cell #3 of the lagoon. In the presence of
332 oxygen, benthic sulfate-reduction was demonstrated to cease (Atkinson et al., 1995),
333 potentially preventing sulfide-induced dissolution of naturally occurring benthic Fe(III)-
334 (oxyhydr)oxides and the reduction of Fe(III) to Fe(II) (Jansson, 1987; Ding et al., 2012;
335 Kumar et al., 2018), lowering total phosphorus (and specifically orthophosphate)
336 concentrations in the water column. The presence/absence of sulfides has been
337 recognized in many studies to be a major cause of release/adsorption of phosphorus to

338 the water column, due to the interactions between sulfide and other phosphate-containing
339 minerals and compounds (Nürnberg, 1984; Morse et al., 1987; Boström et al., 1988). It is
340 therefore believed that upon reaeration of the lagoon, surface adsorption of phosphate to
341 newly available ferric reaction sites in the benthic zone could potentially be a second,
342 minor pathway for phosphorus removal in the lagoon cells during spring-melt.

343 **3.2. Upgrade MBBR treatment train performance**

344 **3.2.1. Nitrifying MBBR reactors**

345 The pilot treatment plant installed at the outlet of a multi-cell facultative wastewater
346 lagoon was operated across a 165-day period spanning winter and spring. During the
347 study, the temperature from the pilot plant's influent wastewater remained below 5°C for
348 a period of 119 days. The performance of the upgrade MBBR pilot system is separated
349 and presented as three distinct operational periods: i) ice-forming period (days 1-50), ii)
350 full ice cover period (days 51-115) and iii) spring thaw period (days 116-165). These
351 periods were selected due to an interest in delineating treatment performance across
352 influent wastewater characteristics entering the pilot system and associating performance
353 of the upgrade pilot treatment train to the lagoon operation. The water quality
354 characteristic results are shown below in Figure 2 across these three periods.

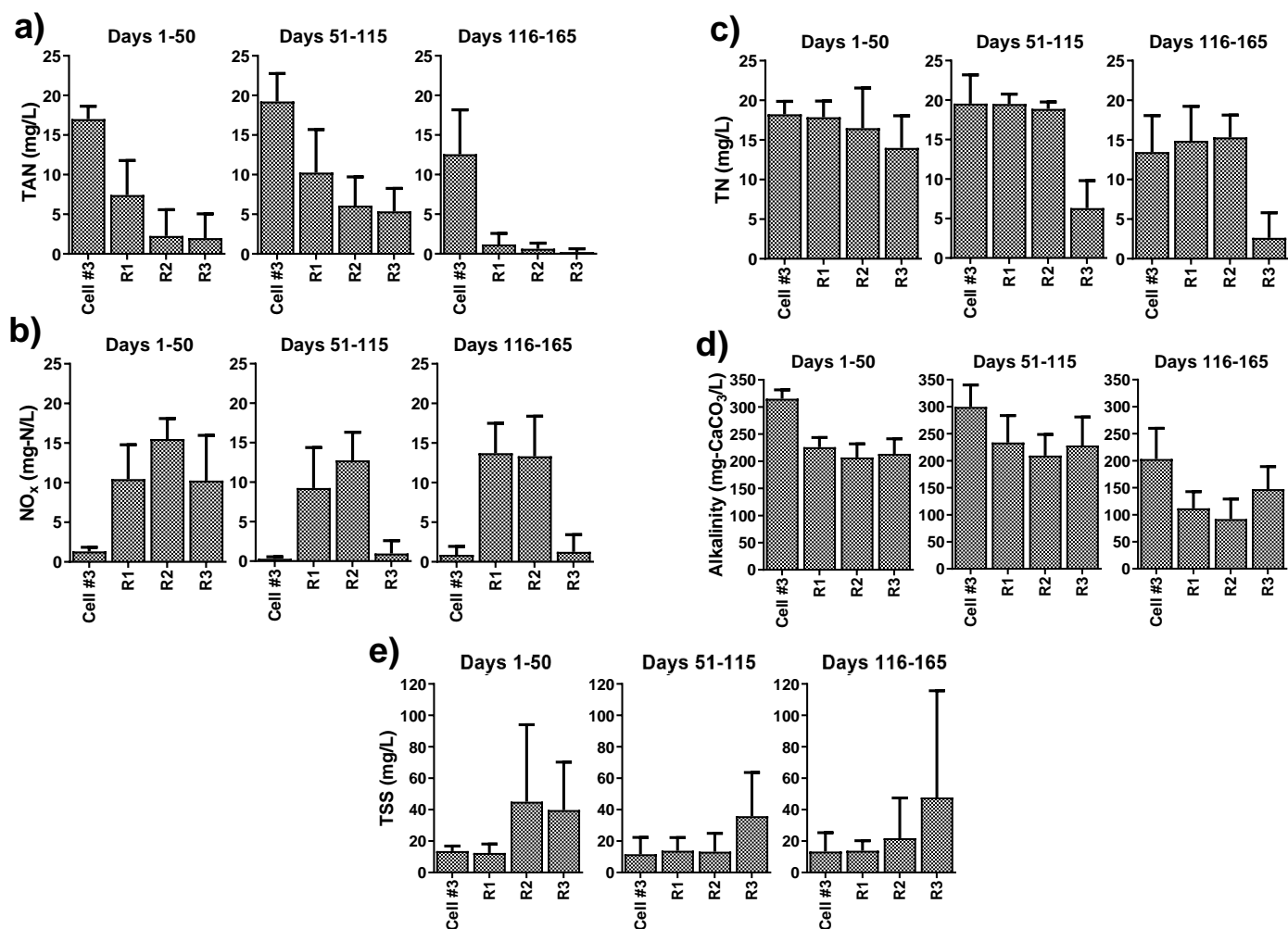


Figure 2. Influent and effluent concentrations across three distinct operational periods: ice-forming period (days 1-50); full ice covered period (days 51-115) and spring thaw period (days 116-165). a) TAN; b) NO_x; c) TN, d) alkalinity and e) TSS.

355 ***Ice-forming period (days 1-50)***

356 The temperatures in the two nitrifying reactors in series, R1 and R2, decreased to
 357 an average value of $3.9 \pm 1.9^\circ\text{C}$ and $3.4 \pm 2.2^\circ\text{C}$ during the ice-forming period of lagoon
 358 operation, respectively. The representative period (days 1-50) removal performance in
 359 terms of removal percentage and removal rate in R1 and R2 was of $55.6 \pm 26.6\%$ (85.0
 360 $\pm 43.0 \text{ g-N/m}^3\cdot\text{d}^{-1}$) and $74.8 \pm 29.8\%$ ($49.1 \pm 44.1 \text{ g-N/m}^3\cdot\text{d}^{-1}$). The total treatment train

361 TAN removal of the nitrifying reactors in-series, R1-R2, was $88.3 \pm 18.0\%$ (134.14 ± 35.03
362 $\text{g-N}\cdot\text{m}^3\cdot\text{d}^{-1}$) (Figure 2 a, b, c). This performance is similar to results observed in a previous,
363 non-acclimatized ultra-low temperature MBBR nitrification study (Ahmed et al., 2019).
364 Alkalinity consumption in the two nitrifying reactors is observed in this study and is
365 proportional to the oxidation of ammonia in the reactors (Figure 2 d). TSS concentrations
366 did not increase significantly within the nitrification train. This is consistent with previous
367 work, as intrinsic cell yields for nitrifiers are low (Madigan et al., 2012; Forrest et al., 2016;
368 Young et al., 2016a). During this period, NO_2^- accounted for $11.5 \pm 14.2\%$ of NO_x in R1
369 and $57.2 \pm \%$ of NO_x in R2. This difference in NO_x is potentially due to the fact that R1
370 had a slightly lower average operating temperature than R2 ($3.9 \pm 1.9^\circ\text{C}$ and $3.4 \pm 2.2^\circ\text{C}$
371 respectively), causing nitrite accumulation (Delatolla et al., 2009; Wang and Li, 2015). It
372 has also been hypothesized that sulfides may also have bled through the reactor
373 treatment train inside the pilot treatment plant, which would have caused low-level
374 inhibition of nitrification rates, but this was not verified.

375 ***Full ice cover period (days 51-115)***

376 Full ice-cover operation (days 51-115) did not demonstrate a significant change in
377 average reactor temperature in the two nitrifying reactors in-series, R1 and R2 ($2.9 \pm$
378 1.0°C and $3.5 \pm 1.5^\circ\text{C}$ respectively). The average TAN percent removal and removal rate
379 in R1 and R2 was of $45.8 \pm 27.8\%$ ($54.2 \pm 33.2 \text{ g-N}/\text{m}^3\cdot\text{d}^{-1}$) and $41.2 \pm 27.4.0\%$ ($24.7 \pm$
380 $21.0 \text{ g-N}/\text{m}^3\cdot\text{d}^{-1}$). The total treatment train performance of both nitrifying reactors in series
381 (R1 + R2) was slightly lower than the ice-forming period (day 1-50), with TAN percent
382 removal of $70.5 \pm 20.2\%$ ($66.6 \pm 43.2 \text{ g-N}/\text{m}^3\cdot\text{d}^{-1}$). Nitrification in the first nitrifying reactor

383 (R1) was slightly lower in the full ice cover period (days 51-115) as compared to the ice-
384 forming period (days 1-50). Water temperatures did not change significantly between the
385 ice-forming and full ice cover periods, and it is therefore believed that the slight decrease
386 in observed TAN removal rates in the first nitrifying reactor (R1) may have been caused
387 by hydrogen sulfide bleed-through into R1, impacting the performance of the TAN
388 removal. No notable changes were observed in TSS or alkalinity concentrations within
389 the nitrification train. During this period, NO_2^- accounted for $60.2 \pm 21.9\%$ of NO_x in R1
390 and $57.2 \pm 21.4\%$ of NO_x in R2. The nitrite accumulation is believed to occur because of
391 NOB temperature inhibition (Wang and Li, 2015). Another possible contributing factor
392 could be that low-level sulfide bleed-through preferentially inhibits the (NOB) compared
393 to the ammonia-oxidizing-bacteria (AOB) (Erguder et al., 2008) in nitrifying biofilm
394 communities.

395 ***Spring thaw period (days 116-165)***

396 The spring thaw operation period of the study was characterized by increasing
397 water temperatures and thawing of lagoon ice cover. Average temperatures in the
398 nitrifying treatment train (R1 + R2) rapidly increased from day 116 to day 125, from 3.1°C
399 to 12.5°C and 3.1°C to 12.7°C , respectively. Temperatures further increased until the end
400 of the spring thaw period, reaching 20.6°C in R1 and 20.7°C in R2. The spring thaw period
401 removal performance increased in the nitrifying treatment train (R1 + R2) as compared to
402 the full ice cover period, demonstrated by higher TAN percentage removal and TAN
403 removal rates of $92.1 \pm 7.7\%$ ($65.1 \pm 16.0 \text{ g-N/m}^3\cdot\text{d}^{-1}$) and $46.8 \pm 28.7\%$ ($3.3 \pm 4.1 \text{ g-}$
404 $\text{N/m}^3\cdot\text{d}^{-1}$) during the spring thaw period. With increasing temperatures and decreasing

405 reactor loading due to drop in influent TAN concentrations, the majority of TAN was
 406 removed by the first nitrifying reactor in-series (R1), with the second nitrifying reactor in-
 407 series (R2) not performing significant removal. The spring thaw period was also
 408 characterized by icemelt and snowmelt, leading to some potential dilution effects on some
 409 of the water quality parameters, including TP and alkalinity. The TAN removal
 410 performance of the complete nitrifying treatment train (R1 + R2) was of $95.7 \pm 4.2\%$ (65.6
 411 $\pm 17.8 \text{ g-N}\cdot\text{m}^3\cdot\text{d}^{-1}$). TSS concentrations in R2 increased throughout the period to reach
 412 effluent concentrations of $58.5 \pm 16.3 \text{ mg/L}$, likely due to the start of reactor starvation
 413 and sloughing of some attached biofilm.

414 ***Effects of temperature on nitrifying treatment train performance***

415 Throughout the study, the nitrifying treatment train achieved a TAN percent
 416 removal of $82.1 \pm 19.6\%$, with an average TAN effluent concentration of $3.42 \pm 3.78 \text{ mg/L}$.
 417 The maximum nitrifying removal rate observed during this period was $157.25 \text{ g-N/m}^3\cdot\text{d}$.
 418 During spring-melt, it is possible that the nitrifying treatment train (R1 + R2) may have
 419 been TAN limited, especially during the end of the study. Near complete conversion of
 420 TAN to NO_x in the first nitrifying reactor (R1) was possible at higher temperatures

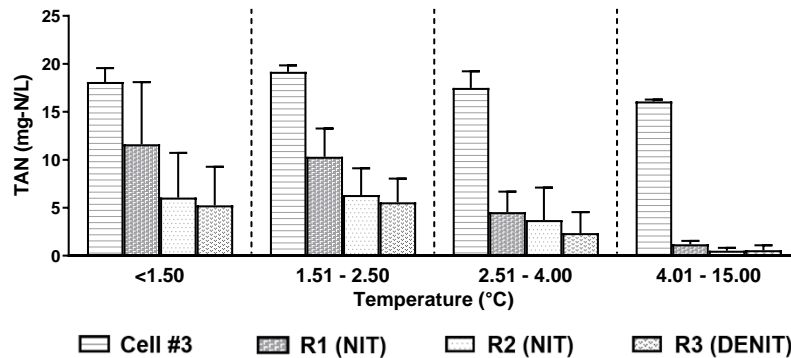


Figure 3. Ranges of TAN concentrations in the MBBR treatment train, by temperature.

421 observed during the spring thaw period (day 115-165), however, at lower temperatures
422 only partial nitrification was achieved in the first nitrifying reactor (R1), with the remainder
423 occurring in the second nitrifying reactor in-series (R2) (Figure 3). Over the duration of
424 the study, R1's TAN removal performance was on average 15.6% higher at operational
425 temperatures ranging between 1.5 – 2.5°C versus <1.5°C, outlining noticeable
426 operational differences occurring over the span of less than 2.5°C. At temperatures below
427 1.5°C, the average TAN removal rate of the complete nitrifying treatment train (R1 + R2)
428 was $52.34 \pm 0.52 \text{ g-N/m}^3\cdot\text{d}^{-1}$.

429 **3.2.2. Denitrifying MBBR reactor**

430 The denitrifying reactor, R3, in the pilot system was installed and operated
431 downstream of the two nitrifying reactors. In an attempt to again identify the effects of
432 temperature on process performance within the MBBR treatment train, results are
433 separated and presented based on the operational conditions of the upstream lagoon
434 system, over the three distinct operational periods: i) ice-forming period (days 1-50), ii)
435 full ice cover period (days 51-115) and iii) spring thaw period (days 116-165).

436 ***Ice-forming period (days 1-50)***

437 The ice-forming period was characterized by low wastewater temperatures. The
438 period average temperature was of $3.2 \pm 2.6^\circ\text{C}$. The ice-forming period NO_x removal
439 performance in the denitrifying reactor, R3, was $20.8 \pm 23.2\%$ ($38.0 \pm 49.9 \text{ g-N}\cdot\text{m}^3\cdot\text{d}^{-1}$).
440 During this period, R3 produced alkalinity at an average rate of $1.97 \text{ mg as CaCO}_3/\text{mg}$ of
441 N denitrified. This is slightly lower than values observed by other in studies of $2.85 - 3.93$
442 $\text{mg-CaCO}_3/\text{mg-N}$ (Jeris and Owens, 1975; Hamlin et al., 2008), but may be explained by

443 the partial denitrification of nitrite and nitrate, hence producing less alkalinity to the system
444 (Asano et al., 2007; Chung et al., 2014). TSS concentrations increased slightly in the
445 denitrifying reactor (R3) due to the relatively high cell yield of denitrifiers compared the
446 nitrifiers (Strohm et al., 2007), leading to ice-forming period-average effluent TSS
447 concentrations of 39.8 ± 30.5 mg/L.

448 ***Full ice cover period (days 51-115)***

449 The full ice cover period (days 51-115) was characterized by no significant change
450 in average reactor temperature in the denitrifying reactor, R3, ($4.4 \pm 2.0^\circ\text{C}$). The period
451 NO_x removal performance in R3 was $90.8 \pm 18.9\%$ (66.9 ± 26.8 g-N·m³·d⁻¹). During this
452 period, the denitrifying reactor (R3) produced alkalinity at an average rate of 2.01 mg as
453 CaCO_3 /mg of N denitrified. This is slightly lower than values observed by other in studies
454 of 2.85 – 3.93 mg- CaCO_3 /mg-N due to the relatively large proportion of NO_x being in the
455 NO_2^- instead of the NO_3^- form in R2 ($57.2 \pm 21.4\%$ for NO_2^-). Similarly to what was
456 observed during the first period, TSS concentrations increased slightly in the denitrifying
457 reactor (R3), leading to a period-average effluent TSS concentration of 36.8 ± 28.3 mg/L.

458 ***Spring thaw period (days 116-165)***

459 The spring thaw period of the study was characterized by increasing water
460 temperatures and thawing of lagoon ice cover. The average temperature in the
461 denitrifying reactor (R3) rapidly increased from 1.6°C to 13.0°C between days 116-125.
462 Temperatures further increased until the end of the period, reaching 20.6°C by the end of
463 the study. The period NO_x removal performance in the denitrifying reactor (R3) was $91.4 \pm$

464 14.7% ($62.2 \pm 18.3 \text{ g-N}\cdot\text{m}^3\cdot\text{d}^{-1}$). Lagoon icemelt led to a global dilution effect of most
465 water quality parameters, including nitrogen and alkalinity. During this period, the
466 denitrifying R3 produced alkalinity at an average rate of 3.68 mg as CaCO_3 /mg of N
467 denitrified. Stoichiometrically, heterotrophic denitrification theoretically yields
468 approximately 3.57 mg- CaCO_3 /mg-N (Jeris and Owens, 1975; Asano et al., 2007). The
469 closeness of observed alkalinity yield to theoretical values is explained by the near total
470 proportion of NO_x as NO_3^- ($82.9 \pm 28.4\%$). TSS concentrations in the denitrifying reactor
471 R3 increased throughout the period to reach effluent concentrations of $60.9 \pm 78.0 \text{ mg/L}$
472 TSS. TSS concentrations were notably higher than in the first two periods (ice-forming
473 and full ice cover), likely due to biofilm thinning in warmer operational temperatures,
474 causing sloughing (Bjornberg et al., 2012). It is important to note that the denitrifying
475 reactor removal rate was limited by the system's influent TAN concentration, and
476 therefore no optimization (increase in removal rate) could be performed. The authors also
477 wish to indicate that denitrifying reactor were dosed with COD in excess, to ensure that
478 the system was never COD limited. It is also noted that with such an approach, an
479 additional reactor might need required for polishing and prevent bleed-through of COD.

480 ***Effects of temperature on denitrifying reactor performance***

481 Throughout the study, there was significant removal of NO_x in the denitrifying
 482 reactor (R3), with an average removal efficiency of 94.3 ± 0.1% of NO_x at temperatures
 483 from <1.5°C to 4°C. At higher temperatures (4.0°C to 15.0°C), removal efficiencies
 484 decreased to 80.3 ± 19.0% (Figure 4). This decrease in removal efficiency was due to an
 485 upset in the operation of the denitrifying reactor over a period of approximately one and
 486 a half weeks (failure of a carbon source pump tubing). At temperatures below 1.5°C, the
 487 average NO_x removal rate of R3 was of 69.75 ± 24.49 g-N/m³·d⁻¹.

488

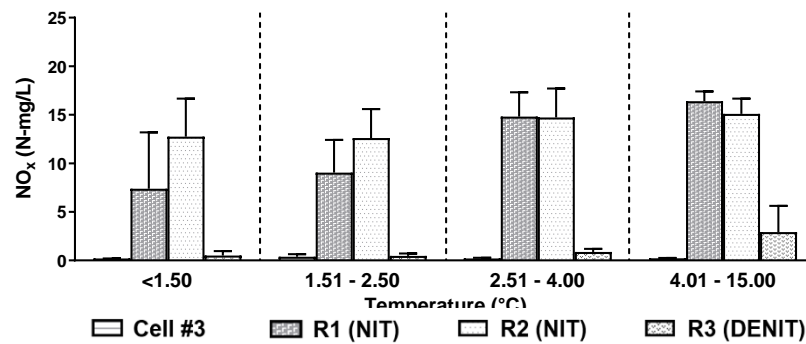


Figure 4. Ranges of NO_x concentrations in the MBBR treatment train, sorted by temperature.

489

490

491 3.2.3. Phosphorus cycling within reactors

492 TP concentrations within the lagoon's cell #3, the nitrifying reactors in-series (R1
493 + R2) remained similar throughout the ice-forming (day 1-50) and full ice cover (day 51-
494 115) periods, with average TP concentrations of 0.33 ± 0.09 mg-P/L, 0.29 ± 0.08 mg-P/L
495 and 0.29 ± 0.02 mg-P/L, respectively (Figure 5). The denitrifying reactor, R3, exhibited an
496 interesting trend throughout the study, as the denitrifying organisms assimilated TP,
497 substantially decreasing R3's TP concentrations, rarely exceeding 0.10 mg P/L. TP
498 concentrations within the denitrifying reactor (R3) remained relatively constant throughout
499 the study, with the denitrifying reactor maintaining an average TP removal efficiency of
500 $76.6 \pm 6.2\%$ in reference to R2. This behaviour is well understood and common for post-
501 nitrification denitrifying treatment (Boltz et al., 2012; Mases et al., 2012; Wilson et al.,
502 2012), outlining an additional benefit of deploying denitrifying MBBR systems as an end-
503 of-pipe phosphorus polishing system in constant-discharge or seasonally-discharged
504 lagoon treatment systems due to the phosphorus requirements of heterotrophic
505 denitrifying bacteria to sustain metabolic activity (Kuba et al., 1996; Aspegren et al.,

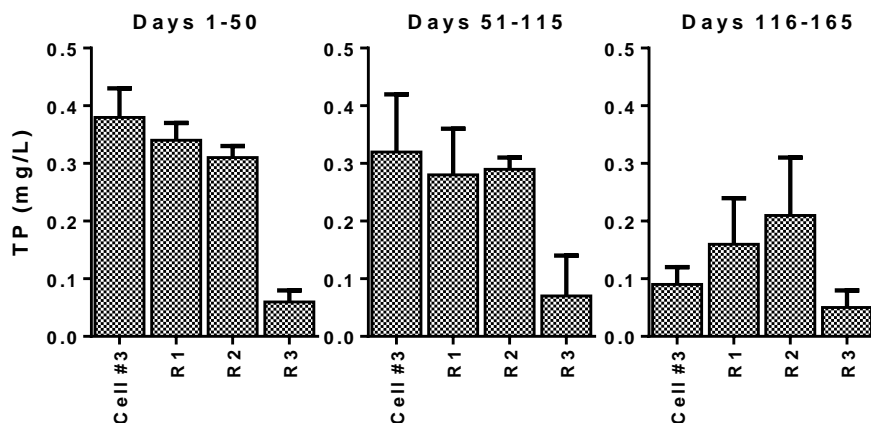


Figure 5. Progression of influent and in-reactor TP concentrations throughout the ice-forming period (days 1-50), full ice cover period (day 51-115) and spring thaw period (116-165).

506 1998). Meanwhile, minimal decreases in TP were observed in the two nitrifying reactors
507 (R1 + R2), likely due to the intrinsically low cell yield of autotrophic nitrifying organisms
508 (Gee et al., 1990).

509 As water temperatures increased during spring-melt, influent TP concentrations
510 decreased significantly. It is believed that the decrease was due to reaeration of the
511 lagoon cells, the inhibition of sulfate-reduction and strong dilution effects. Simultaneously,
512 TP concentrations in the nitrifying reactors in-series (R1 + R2) increased. TP
513 concentrations in the first nitrifying reactor in-series (R1) increased to 0.15 ± 0.08 mg-P/L,
514 peaking at 0.36 mg-P/L. Simultaneously, TP concentrations in the second nitrifying
515 reactor (R2) increased to 0.22 ± 0.09 mg-P/L, peaking at 0.47 mg-P/L. It is hypothesized
516 that the increase of TP concentrations in R1 and R2 may have been caused by a biofilm
517 response to changes in ambient TP concentrations. This “homeostasis” response had
518 previously been reported in eukaryotic cells (Wild et al., 2016) via inositol polyphosphate
519 signaling molecules (InsPs). A similar pathway was recently demonstrated for *E. coli*
520 (McCleary, 2017; Chande and Bergwitz, 2018). A similar response may have occurred in
521 the nitrifying and denitrifying biofilm in this study. The release of phosphorus initiated in
522 the nitrifying treatment train (R1 + R2) is thought to have occurred due to a stress
523 response to a rapid decrease in influent phosphorus concentrations. It is therefore
524 suggested that nitrifying communities within biofilms may be capable of regulating bulk
525 liquid phosphorus concentrations during stress events (Nordeidet et al., 1994). It has
526 been shown in previous studies that nitrifying bacterial communities are regarded as
527 phosphorus limited at concentrations below 0.10 mg-P/L (Helness et al., 1999). Studies
528 have also demonstrated a similar trend in denitrifying bacterial communities, where they

529 are considered phosphorus limited at concentrations between 0.03 to 0.10 mg-P/L.
530 Heterotrophic denitrification in attached-growth systems has a relatively high cell yield
531 and requires a minimum TP concentration to occur uninhibited (Nordeidet et al., 1994;
532 deBarbadillo et al., 2014).

533 4. Conclusions and recommendations

534 Following the completion of this study, the conclusions outlined are as follow:

- 535 1. The MBBR treatment technology has emerged as a suitable and sustainable upgrade
536 technology for existing lagoon and waste stabilization pond facilities operating in
537 temperate, northern, and cold climate countries.
- 538 2. Implementation of nitrifying and denitrifying MBBRs in-series at the outlet of a lagoon
539 in a NIT-NIT-DENIT configuration provided treatment robustness in terms of both total
540 nitrogen and phosphorus removal at temperatures as low as 0.6°C.
- 541 3. Throughout the 165-day study, the lagoon effluent treated by the pilot treatment plant
542 demonstrates that TN concentrations decreased by an average of $69.0 \pm 24.5\%$,
543 where the average TN concentration exiting the pilot treatment plant were 7.60 ± 5.70
544 mg-N/L.
- 545 4. The nitrifying MBBR reactors achieved TAN removal of $82.1 \pm 19.6\%$, reducing TAN
546 concentrations to an average of 3.42 ± 3.78 mg/L. The denitrifying MBBR reactor
547 achieved NO_x removal of $69.5 \pm 37.8\%$, reducing NO_x to an average of 3.63 ± 5.44
548 mg/L.
- 549 5. It is hypothesized that sulfide toxicity in lagoons might be another important limitation
550 to cold-temperature wintertime nitrification, leading to the hypothesis that nitrification
551 in lagoons situated in northern or temperate climates may not only be inhibited by
552 temperature effects, but also by significant sulfide inhibition events.

- 553 6. The use of the denitrification technology treat lagoon effluent demonstrates significant
554 phosphorus removal due to phosphorus assimilation concomitantly occurring with
555 total nitrogen removal through denitrification.
- 556 7. Throughout the 165-day study, the lagoon effluent treated by the pilot treatment plant
557 saw TP concentrations decrease by $74.7 \pm 20.1\%$. In addition, the average TP
558 concentrations exiting the pilot treatment plant were 0.05 ± 0.02 mg-P/L.
- 559 8. It is hypothesized that the embedded biofilm microorganisms can possibly perform
560 phosphorus homeostasis and increasing bulk-water phosphorus concentrations in the
561 reactors, as a response to low influent phosphorus stress induced by cessation of in-
562 lagoon sulfate-reduction and dilution effects.
- 563 9. The results of the study suggest that nitrifying + denitrifying MBBR systems can be
564 installed at the outlets of traditional facultative lagoons to achieve reliable TAN, TN
565 and TP treatment, thereby providing a pathway for substantially reducing the
566 environmental impact of municipal wastewater lagoon discharge on receiving waters
567 in Canada and other northern communities during wintertime operation.

568 **Declaration of competing interests**

569 The authors confirm that there are no known conflicts of interest associated with
570 this publication and there has been no significant financial support for this work that could
571 have influenced its outcome.

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578 **Data availability statement**

579 The data that support the findings of this study are available from the
580 corresponding author upon reasonable request.

581 **References**

- 582 Ahmed, W., Tian, X., Delatolla, R., 2019. Nitrifying moving bed biofilm reactor:
583 Performance at low temperatures and response to cold-shock. *Chemosphere* 229,
584 295–302. <https://doi.org/10.1016/j.chemosphere.2019.04.176>
- 585 Almomani, F. a., Delatolla, R., Örmeci, B., 2014. Field study of moving bed biofilm reactor
586 technology for post-treatment of wastewater lagoon effluent at 1°C. *Environ. Technol.*

587 35, 1596–1604. <https://doi.org/10.1080/09593330.2013.874500>

588 Andreottola, G., Foladori, P., Ragazzi, M., 2000. Upgrading of a small wastewater
589 treatment plant in a cold climate region using a moving bed biofilm reactor (MBBR)
590 system. *Water Sci. Technol.* 41, 177–185.

591 APHA, WEF, A., 2012. Standard methods for the examination of water and wastewater,
592 22nd ed. APHA, WEF, AWWA, Washington, D.C.

593 Archer, H.E., O'Brien, B.M., 2005. Improving nitrogen reduction in waste stabilisation
594 ponds. *Water Sci. Technol.* 51, 133–138. <https://doi.org/10.2166/wst.2005.0446>

595 Asano, T., Burton, F.L., Leverenz, H.L., Tsuchihashi, R., Tchobanoglous, G., 2007. *Water*
596 *Reuse Issues, Technologies, and Applications*, 1st editio. ed, Journal of Chemical
597 Information and Modeling. McGraw-Hill.

598 Aspegren, H., Nyberg, U., Andersson, B., Gotthardsson, S., Jansen, J.L.C., 1998. Post
599 denitrification in a moving bed biofilm reactor process. *Water Sci. Technol.* 38, 31–
600 38. [https://doi.org/10.1016/S0273-1223\(98\)00387-4](https://doi.org/10.1016/S0273-1223(98)00387-4)

601 Atkinson, T., Sherwood, R.F., Barton, L.L., 1995. *Biotechnology Handbooks 8 Sulfate-*
602 *reducing bacteria*, *Biotechnology Handbooks*.

603 Avsar, Y., Tarabeah, H., Kimchie, S., Naamneh, H., Ozturk, I., 2008. Rehabilitation of an
604 available facultative pond unit using a trickling biofilter. *Environ. Eng. Sci.* 25, 106–
605 113. <https://doi.org/10.1089/ees.2007.0034>

606 Bejarano Ortiz, D.I., Thalasso, F., Cuervo López, F. de M., Texier, A.C., 2013. Inhibitory
607 effect of sulfide on the nitrifying respiratory process. *J. Chem. Technol. Biotechnol.*
608 88, 1344–1349. <https://doi.org/10.1002/jctb.3982>

609 Bjornberg, C., Lin, W., Zimmerman, R., 2012. Effect of Temperature on Biofilm Growth
610 Dynamics and Nitrification Kinetics in a Full-Scale MBBR System. *Proc. Water*
611 *Environ. Fed.* 2009, 4407–4426. <https://doi.org/10.2175/193864709793954051>

612 Boltz, J.P., Morgenroth, E., Daigger, G.T., deBarbadillo, C., Murthy, S., Sørensen, K.,
613 Stinson, B., 2012. Process Control to Achieve Simultaneous Low-Level Effluent
614 Nitrogen and Phosphorus Concentrations with Post-Denitrification Moving Bed
615 Biofilm Reactor (MBBR) and Biologically Active Filter (BAF) Systems. *Proc. Water*
616 *Environ. Fed.* 2010, 4172–4178. <https://doi.org/10.2175/193864710798182286>

617 Boström, B., Andersen, J.M., Fleischer, S., Jansson, M., 1988. Exchange of phosphorus
618 across the sediment-water interface. *Hydrobiologia* 170, 229–244.
619 <https://doi.org/10.1007/BF00024907>

620 Brettle, M., Berry, P., Paterson, J., Yasvinski, G., 2016. Determining Canadian water
621 utility preparedness for the impacts of climate change. *Chang. Adapt. Socio-*
622 *Ecological Syst.* 2, 124–140. <https://doi.org/10.1515/cass-2015-0024>

623 Bruce, 2011, 2009. Lagoon Construction Guidelines. Alaska Dep. Environ. Conserv.
624 <https://doi.org/10.1017/CBO9781107415324.004>

625 Butterworth, E., Richards, A., Jones, M., Brix, H., Dotro, G., Jefferson, B., 2016. Impact

626 of aeration on macrophyte establishment in sub-surface constructed wetlands used
627 for tertiary treatment of sewage. *Ecol. Eng.* 91, 65–73.
628 <https://doi.org/10.1016/j.ecoleng.2016.01.017>

629 Caffrey, J.M., Bonaglia, S., Conley, D.J., 2019. Short exposure to oxygen and sulfide alter
630 nitrification, denitrification, and DNRA activity in seasonally hypoxic estuarine
631 sediments. *FEMS Microbiol. Lett.* 366, 1–10. <https://doi.org/10.1093/femsle/fny288>

632 Cameron, K., Madramootoo, C., Crolla, A., Kinsley, C., 2003. Pollutant removal from
633 municipal sewage lagoon effluents with a free-surface wetland. *Water Res.* 37,
634 2803–2812. [https://doi.org/10.1016/S0043-1354\(03\)00135-0](https://doi.org/10.1016/S0043-1354(03)00135-0)

635 Canadian Council of Ministers of the Environment, 2009. Canada-wide Strategy for the
636 Management of Municipal Wastewater Effluent. Whitehorse, YK.

637 Chande, S., Bergwitz, C., 2018. Role of phosphate sensing in bone and mineral
638 metabolism. *Nat. Rev. Endocrinol.* 14, 637–655. [https://doi.org/10.1038/s41574-018-](https://doi.org/10.1038/s41574-018-0076-3)
639 [0076-3](https://doi.org/10.1038/s41574-018-0076-3)

640 Chen, L., Delatolla, R., D'Aoust, P.M., Wang, R., Pick, F., Poulain, A., Rennie, C.D., 2017.
641 Hypoxic conditions in stormwater retention ponds: potential for hydrogen sulfide
642 emission. *Environ. Technol. (United Kingdom)* 1–12.
643 <https://doi.org/10.1080/09593330.2017.1400112>

644 Chung, J., Amin, K., Kim, S., Yoon, S., Kwon, K., Bae, W., 2014. Autotrophic
645 denitrification of nitrate and nitrite using thiosulfate as an electron donor. *Water Res.*

646 58, 169–178. <https://doi.org/10.1016/j.watres.2014.03.071>

647 D'Aoust, P.M., Delatolla, R., Poulain, A., Guo, G., Wang, R., Rennie, C., Chen, L., Pick,
648 F.R., 2017. Emerging investigators series: hydrogen sulfide production in municipal
649 stormwater retention ponds under ice covered conditions: a study of water quality
650 and SRB populations. *Environ. Sci. Water Res. Technol.* 3, 686–698.
651 <https://doi.org/10.1039/C7EW00117G>

652 D'Aoust, P.M., Pick, F.R., Wang, R., Poulain, A., Rennie, C., Chen, L., Kinsley, C.,
653 Delatolla, R., 2018. Sulfide production kinetics and model of Stormwater retention
654 ponds. *Water Sci. Technol.* 77, 2377–2387. <https://doi.org/10.2166/wst.2018.150>

655 Dale, C., Laliberte, M., Oliphant, D., Ekenberg, M., 2015. Proceedings of Mine Water
656 Solutions in Extreme Environments Wastewater treatment using MBBR in cold
657 climates 1–17.

658 deBarbadillo, C., Rectanus, R., Canham, R., Schauer, P., 2014. Tertiary Denitrification
659 and Very Low Phosphorus Limits: A Practical Look at Phosphorus Limitations on
660 Denitrification Filters. *Proc. Water Environ. Fed.* 2006, 3454–3465.
661 <https://doi.org/10.2175/193864706783751366>

662 Delatolla, R., Tufenkji, N., Comeau, Y., Gadbois, A., Lamarre, D., Berk, D., 2012. Effects
663 of long exposure to low temperatures on nitrifying biofilm and biomass in wastewater
664 treatment. *Water Environ. Res.* 84, 328–338.
665 <https://doi.org/10.2175/106143012x13354606450924>

666 Delatolla, R., Tufenkji, N., Comeau, Y., Gadbois, A., Lamarre, D., Berk, D., 2010.
667 Investigation of laboratory-scale and pilot-scale attached growth ammonia removal
668 kinetics at cold temperature and low influent carbon. *Water Qual. Res. J.* 45, 427–
669 436. <https://doi.org/10.2166/wqrj.2010.042>

670 Delatolla, R., Tufenkji, N., Comeau, Y., Gadbois, A., Lamarre, D., Berk, D., 2009. Kinetic
671 analysis of attached growth nitrification in cold climates. *Water Sci. Technol.* 60,
672 1173–1184. <https://doi.org/10.2166/wst.2009.419>

673 Delatolla, R.A., Babarutsi, S., 2005. Parameters Affecting Hydraulic Behavior of Aerated
674 Lagoons. *J. Environ. Eng.* 131, 1404–1413. [https://doi.org/10.1061/\(ASCE\)0733-
675 9372\(2005\)131:10\(1404\)](https://doi.org/10.1061/(ASCE)0733-9372(2005)131:10(1404))

676 Ding, S., Sun, Q., Xu, D., Jia, F., He, X., Zhang, C., 2012. High-resolution simultaneous
677 measurements of dissolved reactive phosphorus and dissolved sulfide: The first
678 observation of their simultaneous release in sediments. *Environ. Sci. Technol.* 46,
679 8297–8304. <https://doi.org/10.1021/es301134h>

680 Directive Council of the European Union, 1996. Council Directive 96/61/EC of 24
681 September 1996 concerning integrated pollution prevention and control, *Official
682 Journal of the European Communities*. Brussels.

683 Directive Council of the European Union, 1991. Council Directive 91/271/EEC of 21 May
684 1991 concerning urban waste-water treatment, *Official Journal of the European
685 Communities*. Brussels.

686 Erguder, T.H., Boon, N., Vlaeminck, S.E., Verstraete, W., 2008. Partial nitrification
687 achieved by pulse sulfide doses in a sequential batch reactor. *Environ. Sci. Technol.*
688 42, 8715–8720. <https://doi.org/10.1021/es801391u>

689 Forrest, D., Delatolla, R., Kennedy, K., 2016. Carrier effects on tertiary nitrifying moving
690 bed biofilm reactor: An examination of performance, biofilm and biologically produced
691 solids. *Environ. Technol. (United Kingdom)* 37, 662–671.
692 <https://doi.org/10.1080/09593330.2015.1077272>

693 Gan, C., Champagne, P., 2015. Evaluation of passive treatment technologies for septic
694 lagoon capacity expansion. *World Environ. Water Resour. Congr. 2015 Floods,*
695 *Droughts, Ecosyst. - Proc. 2015 World Environ. Water Resour. Congr. (Austin, TX)*
696 2403–2423. <https://doi.org/10.1061/9780784479162.236>

697 Gee, C.S., Suidan, M.T., Pfeffer, J.T., 1990. Modeling of Nitrification Under Substrate-
698 Inhibiting Conditions. *J. Environ. Eng.* 116, 18–31.
699 [https://doi.org/10.1061/\(ASCE\)0733-9372\(1990\)116:1\(18\)](https://doi.org/10.1061/(ASCE)0733-9372(1990)116:1(18))

700 Gieseke, A., Bjerrum, L., Wagner, M., Amann, R., 2003. Structure and activity of multiple
701 nitrifying bacterial populations co-existing in a biofilm 5, 355–369.

702 Hach Company, 2015. Nitrogen, Ammonia. Protok. Hach Co. Lange GmbH, 1989–2015.
703 All rights Reserv. 1997-2003. All rights Reserv. Print. U.S 1–6.

704 Hamlin, H.J., Michaels, J.T., Beaulaton, C.M., Graham, W.F., Dutt, W., Steinbach, P.,
705 Losordo, T.M., Schrader, K.K., Main, K.L., 2008. Comparing denitrification rates and

706 carbon sources in commercial scale upflow denitrification biological filters in
707 aquaculture. *Aquac. Eng.* 38, 79–92. <https://doi.org/10.1016/j.aquaeng.2007.11.003>

708 Hassard, F., Biddle, J., Cartmell, E., Jefferson, B., Tyrrel, S., Stephenson, T., 2015.
709 Rotating biological contactors for wastewater treatment - A review. *Process Saf.*
710 *Environ. Prot.* 94, 285–306. <https://doi.org/10.1016/j.psep.2014.07.003>

711 Heaven, S., Lock, A.C., Pak, L.N., Rspaev, M.K., 2003. Waste stabilisation ponds in
712 extreme continental climates: A comparison of design methods from the USA,
713 Canada, northern Europe and the former Soviet Union. *Water Sci. Technol.* 48, 25–
714 33.

715 Helness, H., Odegaard, H., Ødegaard, H., 1999. Biological phosphorus removal in a
716 sequencing batch moving bed biofilm reactor. *Water Sci. Technol.* 40, 161–168.

717 Hoang, V., 2013. MBBR ammonia removal: an investigation of nitrification kinetics, biofilm
718 and biomass response, and bacterial population shifts during long-term cold
719 temperature exposure. Univ. Ottawa.

720 Hoang, V., Delatolla, R., Laflamme, E., Gadbois, A., 2014. An Investigation of Moving
721 Bed Biofilm Reactor Nitrification during Long-Term Exposure to Cold Temperatures.
722 *Water Environ. Res.* 86, 36–42.
723 <https://doi.org/10.2175/106143013X13807328848694>

724 Jansson, M., 1987. Anaerobic dissolution of iron-phosphorus complexes in sediment due
725 to the activity of nitrate-reducing bacteria. *Microb. Ecol.* 14, 81–89.

726 <https://doi.org/10.1007/BF02011573>

727 Jeris, J.S., Owens, R.W., 1975. Pilot-scale, high-rate biological denitrification. *J. Water*
728 *Pollut. Control Fed.* 47, 2043–57.

729 Joye, S.B., Hollibaugh, J.T., 1995. Influence of sulfide inhibition of nitrification on nitrogen
730 regeneration in sediments. *Science* (80-). 270, 623–625.
731 <https://doi.org/10.1126/science.270.5236.623>

732 Krkosek, W.H., Ragush, C., Boutilier, L., Sinclair, A., Krumhansl, K., Gagnon, G.A.,
733 Jamieson, R.C., Lam, B., 2012. Treatment performance of wastewater stabilization
734 ponds in Canada's Far North. *Proc. Int. Conf. Cold Reg. Eng.* 612–622.
735 <https://doi.org/10.1061/9780784412473.061>

736 Kuba, T., Murnleitner, E., Van Loosdrecht, M.C.M., Heijnen, J.J., 1996. A metabolic model
737 for biological phosphorus removal by denitrifying organisms. *Biotechnol. Bioeng.* 52,
738 685–695. [https://doi.org/10.1002/\(SICI\)1097-0290\(19961220\)52:6<685::AID-
739 BIT6>3.3.CO;2-M](https://doi.org/10.1002/(SICI)1097-0290(19961220)52:6<685::AID-BIT6>3.3.CO;2-M)

740 Kumar, N., Lezama Pacheco, J., Noël, V., Dublet, G., Brown, G.E., 2018. Sulfidation
741 mechanisms of Fe(iii)-(oxyhydr)oxide nanoparticles: A spectroscopic study. *Environ.*
742 *Sci. Nano* 5, 1012–1026. <https://doi.org/10.1039/c7en01109a>

743 Lewis, W.M., Wurtsbaugh, W.A., Paerl, H.W., 2011. Rationale for control of
744 anthropogenic nitrogen and phosphorus to reduce eutrophication of inland waters.
745 *Environ. Sci. Technol.* 45, 10300–10305. <https://doi.org/10.1021/es202401p>

746 Macdonald, G., Holley, E.R., Goudey, J.S., 1991. Gas transfer measurements on an ice-
747 covered river, in: Air-Water Mass Transfer. ASCE, pp. 347–361.

748 Madigan, M.T.M., Martinko, J.M., Stahl, D.A., Clark, D.P., 2012. Brock Biology of
749 Microorganisms, 14th Editi. ed, International Microbiology. Peason Education,
750 Glenview, IL. <https://doi.org/10.1007/s13398-014-0173-7.2>

751 Mara, D., 2009. Waste stabilization ponds: Past, present and future. Desalin. Water Treat.
752 4, 85–88. <https://doi.org/10.5004/dwt.2009.359>

753 Mara, D.D., Johnson, M.L., 2006. Aerated Rock Filters for Enhanced Ammonia and Fecal
754 Coliform Removal from Facultative Pond Effluents. J. Environ. Eng. 132, 574–577.
755 [https://doi.org/10.1061/\(ASCE\)0733-9372\(2006\)132:4\(574\)](https://doi.org/10.1061/(ASCE)0733-9372(2006)132:4(574))

756 Mases, M., Dimitrova, I., Nyberg, U., Gruvberger, C., Andersson, B., 2012. Experiences
757 from MBBR Post-Denitrification Process in Long-term Operation at two WWTPs.
758 Proc. Water Environ. Fed. 2010, 458–471.
759 <https://doi.org/10.2175/193864710798208791>

760 Mattson, R.R., Wildman, M., Just, C., 2018. Submerged attached-growth reactors as
761 lagoon retrofits for cold-weather ammonia removal: Performance and sizing. Water
762 Sci. Technol. 78, 1625–1632. <https://doi.org/10.2166/wst.2018.399>

763 McCleary, W.R., 2017. Molecular Mechanisms of Phosphate Homeostasis in *Escherichia*
764 *coli*, in: Escherichia Coli - Recent Advances on Physiology, Pathogenesis and
765 Biotechnological Applications. InTech, p. 13. <https://doi.org/10.5772/67283>

766 Melcer, H., Evans, B., Nutt, S.G., Ho, A., 1995. Upgrading effluent quality for lagoon-
767 based systems. *Water Sci. Technol.* 31, 379–387.
768 <https://doi.org/10.2166/wst.1995.0506>

769 Ministry of Ecology and Environment of the People's Republic of China, 1996. Maximum
770 Allowable Discharge Concentrations for Other Pollutants in China.

771 Mittal, G.S., 2006. Treatment of wastewater from abattoirs before land application - A
772 review. *Bioresour. Technol.* 97, 1119–1135.
773 <https://doi.org/10.1016/j.biortech.2004.11.021>

774 MOE Ontario, 2008. Design guidelines for sewage works, Ontario Ministry of the
775 Environment. Canada.

776 Morse, J.W., Millero, F.J., Cornwell, J.C., Rickard, D., 1987. The chemistry of the
777 hydrogen sulfide and iron sulfide systems in natural waters. *Earth-Science Rev.* 24,
778 1–42. [https://doi.org/http://dx.doi.org/10.1016/0012-8252\(87\)90046-8](https://doi.org/http://dx.doi.org/10.1016/0012-8252(87)90046-8)

779 Muga, H.E., Mihelcic, J.R., 2008. Sustainability of wastewater treatment technologies. *J.*
780 *Environ. Manage.* 88, 437–447. <https://doi.org/10.1016/j.jenvman.2007.03.008>

781 Narasiah, K.S., Morasse, C., Lemay, J., 1994. Phosphorus removal from aerated lagoons
782 using alum, ferric chloride and lime. *Water Qual. Res. J.* 29, 1–18.

783 Nordeidet, B., Rusten, B., Ødegaard, H., 1994. Phosphorus requirements for tertiary
784 nitrification in a biofilm. *Water Sci. Technol.* 29, 77–82.
785 <https://doi.org/10.2166/wst.1994.0748>

786 Nürnberg, G., 1984. Iron and hydrogen sulfide interference in the analysis of soluble
787 reactive phosphorus in anoxic waters. *Water Res.* 18, 369–377.

788 Otawa, K., Asano, R., Ohba, Y., Sasaki, T., Kawamura, E., Koyama, F., Nakamura, S.,
789 Nakai, Y., 2006. Molecular analysis of ammonia-oxidizing bacteria community in
790 intermittent aeration sequencing batch reactors used for animal wastewater
791 treatment 8, 1985–1996. <https://doi.org/10.1111/j.1462-2920.2006.01078.x>

792 Painter, H.A., Loveless, J.E., 1983. Effect of temperature and pH value on the growth-
793 rate constants of nitrifying bacteria in the activated-sludge process. *Water Res.* 17,
794 237–248. [https://doi.org/10.1016/0043-1354\(83\)90176-8](https://doi.org/10.1016/0043-1354(83)90176-8)

795 Patry, B., Lessard, P., Vanrolleghem, P.A., 2019. Nitrification in a biofilm-enhanced highly
796 loaded aerated lagoon. *Water Environ. Res.* 1–8. <https://doi.org/10.1002/wer.1234>

797 Preston, S.D., Alexander, R.B., Schwarz, G.E., Crawford, C.G., 2011. Factors Affecting
798 Stream Nutrient Loads: A Synthesis of Regional SPARROW Model Results for the
799 Continental United States. *J. Am. Water Resour. Assoc.* 47, 891–915.
800 <https://doi.org/10.1111/j.1752-1688.2011.00577.x>

801 Ragush, C.M., Schmidt, J.J., Krkosek, W.H., Gagnon, G.A., Truelstrup-Hansen, L.,
802 Jamieson, R.C., 2015. Performance of municipal waste stabilization ponds in the
803 Canadian Arctic. *Ecol. Eng.* 83, 413–421.
804 <https://doi.org/10.1016/j.ecoleng.2015.07.008>

805 Reardon, J., Foreman, J.A., Searcy, R.L., 1966. New reactants for the colorimetric

806 determination of ammonia. *Clin. Chim. Acta* 14, 403–405.
807 [https://doi.org/10.1016/0009-8981\(66\)90120-3](https://doi.org/10.1016/0009-8981(66)90120-3)

808 Senga, Y., Mochida, K., Fukumori, R., Okamoto, N., Seike, Y., 2006. N₂O accumulation
809 in estuarine and coastal sediments: The influence of H₂S on dissimilatory nitrate
810 reduction. *Estuar. Coast. Shelf Sci.* 67, 231–238.
811 <https://doi.org/10.1016/j.ecss.2005.11.021>

812 Shao, M.F., Zhang, T., Fang, H.H.P., 2010. Sulfur-driven autotrophic denitrification:
813 Diversity, biochemistry, and engineering applications. *Appl. Microbiol. Biotechnol.* 88,
814 1027–1042. <https://doi.org/10.1007/s00253-010-2847-1>

815 Sharma, B., Ahlert, R.C., 1977. Nitrification and nitrogen removal. *Water Res.* 11, 897–
816 925. [https://doi.org/10.1016/0043-1354\(77\)90078-1](https://doi.org/10.1016/0043-1354(77)90078-1)

817 Shin, H.K., Polprasert, C., 1988. Ammonia nitrogen removal in attached-growth ponds. *J.*
818 *Environ. Eng. (United States)* 114, 846–863. [https://doi.org/10.1061/\(ASCE\)0733-](https://doi.org/10.1061/(ASCE)0733-9372(1988)114:4(846))
819 [9372\(1988\)114:4\(846\)](https://doi.org/10.1061/(ASCE)0733-9372(1988)114:4(846))

820 Smith, V.H., Tilman, G.D., Nekola, J.C., 1999. Eutrophication: Impacts of excess nutrient
821 inputs on freshwater, marine, and terrestrial ecosystems. *Environ. Pollut.* 100, 179–
822 196. [https://doi.org/10.1016/S0269-7491\(99\)00091-3](https://doi.org/10.1016/S0269-7491(99)00091-3)

823 Srinivas, D., 2007. Nitrification enhancement in lagoons using fixed film media.

824 Statistics Canada, 2018. Canada's Core Public Infrastructure Survey : Wastewater and
825 solid waste assets, 2016. Dly.

826 Statistics Canada, 2016. Canada's Core Public Infrastructure Survey : Wastewater and
827 solid waste assets, 2016.

828 Strohm, T.O., Griffin, B., Zumft, W.G., Schink, B., 2007. Growth yields in bacterial
829 denitrification and nitrate ammonification. *Appl. Environ. Microbiol.* 73, 1420–1424.
830 <https://doi.org/10.1128/AEM.02508-06>

831 Swanson, G.R., Williamson, K.J., 1980. Rock Filters for Removal of Algae from Lagoon
832 Effluents. Municipal Environmental Research Laboratory, Office of Research and

833 U.S. Environmental Protection Agency, 1978. Methods for Chemical Analysis of Water
834 and Wastes, Method 375.4 - Sulfate (Turbidimetric). Environmental Monitoring and
835 Support Laboratory, Washington, D.C.

836 USEPA, 2011. Principles of design and operations of wastewater treatment pond systems
837 for plant operators, engineers, and managers, 2nd ed. US Environmental Protection
838 Agency Cincinnati, OH, Cincinnati, OH.

839 USEPA, 2002. Federal Water Pollution Control Act 234. [https://doi.org/00024720-](https://doi.org/00024720-200608000-00007)
840 [200608000-00007](https://doi.org/00024720-200608000-00007) [pii]

841 USEPA, 2000. Wastewater Technology Fact Sheet Facultative Lagoons. *Environ. Prot.*
842 *Agency* 1–7. [https://doi.org/EPA 832-F-99-062](https://doi.org/EPA%20832-F-99-062)

843 Van Dyke, S., Jones, S., Ong, S.K., 2003. Cold weather nitrogen removal deficiencies of
844 aerated lagoons. *Environ. Technol. (United Kingdom)* 24, 767–777.
845 <https://doi.org/10.1080/09593330309385613>

846 Wang, L., Li, T., 2015. Effects of seasonal temperature variation on nitrification, anammox
847 process, and bacteria involved in a pilot-scale constructed wetland. *Environ. Sci.*
848 *Pollut. Res.* 22, 3774–3783. <https://doi.org/10.1007/s11356-014-3633-x>

849 Wang, W., 1980. Fractionation of sediment oxygen demand. *Water Res.* 14, 603–612.
850 [https://doi.org/10.1016/0160-4120\(84\)90232-0](https://doi.org/10.1016/0160-4120(84)90232-0)

851 Wessman, F.G., Johnson, C., 2006. Cold Weather Nitrification of Lagoon Effluent Using
852 a Moving Bed Biofilm Reactor (MBBR) Treatment Process, in: WEFTEC 2006. Dallas,
853 Texas, pp. 4738–4750.

854 Wild, R., Gerasimaite, R., Jung, J., Truffault, V., Pavlovic, I., Schmidt, A., Jessen, H.J.,
855 Poirier, Y., Hothorn, M., Mayer, A., 2016. Control of eukaryotic phosphate
856 homeostasis by inositol polyphosphate sensor domains. *Science* (80-.). 9858, 1–9.

857 Wilson, T., Toth, Z., Anderson, G., McSweeney, R., McGettigan, J., 2012. Using MBBRs
858 to Meet ENR Nitrogen Levels for Over 8 Years. *Proc. Water Environ. Fed.* 2008,
859 3622–3630. <https://doi.org/10.2175/193864708788733233>

860 Young, B., Banihashemi, B., Forrest, D., Kennedy, K., Stintzi, A., Delatolla, R., 2016a.
861 Meso and micro-scale response of post carbon removal nitrifying MBBR biofilm
862 across carrier type and loading. *Water Res.* 91, 235–243.
863 <https://doi.org/10.1016/j.watres.2016.01.006>

864 Young, B., Delatolla, R., Abujamel, T., Kennedy, K., Laflamme, E., Stintzi, A., 2017a.
865 Rapid start-up of nitrifying MBBRs at low temperatures: nitrification, biofilm response

866 and microbiome analysis. *Bioprocess Biosyst. Eng.* 40, 731–739.
867 <https://doi.org/10.1007/s00449-017-1739-5>

868 Young, B., Delatolla, R., Abujamel, T., Kennedy, K., Laflamme, E., Stintzi, A., 2017b.
869 Rapid start-up of nitrifying MBBRs at low temperatures: nitrification, biofilm response
870 and microbiome analysis. *Bioprocess Biosyst. Eng.* 40, 731–739.
871 <https://doi.org/10.1007/s00449-017-1739-5>

872 Young, B., Delatolla, R., Ren, B., Kennedy, K., Stintzi, A., Young, B., Delatolla, R., Ren,
873 B., Kennedy, K., Laflamme, E., 2016b. Pilot-scale tertiary MBBR nitrification at 1 ° C :
874 characterization of ammonia removal rate , solids settleability and biofilm
875 characteristics. *Environ. Technol.* 37, 2124–2132.
876 <https://doi.org/10.1080/09593330.2016.1143037>

877

878 **Captions for Figures and Tables in text (as ordered).**

879 Figure 1: Schematic of the Casselman municipal facultative lagoon WRRF and MBBR nitrogen
880 removal MBBR pilot systems.

881 Table 1. Average lagoon effluent concentrations of water quality parameters throughout the study
882 period.

883 Table 2. Average \pm SD of TAN, nitrate, TS and TP concentrations in cell #3 throughout the study.

884 Figure 2. Influent and effluent concentrations across three distinct operational periods: ice-forming
885 period (days 1-50); full ice-covered period (days 51-115) and spring thaw period (days 116-165)).
886 a) TAN; b) NO_x; c) TN, d) alkalinity and e) TSS.

887 Figure 3. Ranges of TAN concentrations in the MBBR treatment train, by temperature.

888 Figure 4. Ranges of NO_x concentrations in the MBBR treatment train, sorted by temperature.

889 Figure 5. Progression of influent and in-reactor TP concentrations throughout the ice-forming
890 period (days 1-50), full ice cover period (day 51-115) and spring thaw period (116-165).