

# Good modeling practice for the calibration of ion exchange breakthrough prediction

Daniel Illana González<sup>a,b</sup> \*, Mariane Yvonne Schneider<sup>a,b</sup> , Juan Pablo Gallo<sup>a,b</sup>, Ingmar Nopens<sup>a,b</sup>, Elena Torfs<sup>c</sup>

<sup>a</sup> BIOMATH, Department of Data Analysis and Mathematical Modelling. Ghent University, Coupure Links 653, 9000 Gent, Belgium.

<sup>b</sup> CAPTURE, Centre for Advanced Process Technology for Urban REsource recovery, Frieda Saeystraat 1, 9052 Gent

<sup>c</sup> modelEAU, Département de génie civil et de génie des eaux, Université Laval, pavillon Adrien-Pouliot, 1065, av. de la Médecine, Québec, G1V 0A6, Canada.

\* Corresponding author. E-mail: [Daniel.IllanaGonzalez@UGent.be](mailto:Daniel.IllanaGonzalez@UGent.be)

## Abstract

Ion exchange (IX) is a key technology in resource recovery processes for demineralization and fit-for-purpose water production due to its inherent ion-selective recovery properties. A major bottleneck in the optimization of the IX process is the accurate prediction of ion breakthrough times, which has the potential to save on regeneration chemicals by maximizing resin utilization. However, the models used to predict ion breakthrough times are often unreliable due to poor calibration methods and significant uncertainty in parameter estimates. A well-calibrated ion breakthrough prediction model could provide valuable insights into the process, enabling optimization and model-based control with the aim of improving the overall efficiency and sustainability of the process. Consequently, we conducted local and global sensitivity analyses to identify the design and equilibrium parameters that contribute most to the prediction of breakthrough curves. The global sensitivity analysis allowed us to select a limited subset of parameters for calibration, which showed that only two parameters, namely the maximum adsorption capacity isotherm parameter and the resin particle size, need to be thoroughly calibrated to obtain an accurate prediction of the breakthrough curve. We also showed that uncertainty quantification for model calibration is important to establish the reliability of the predictions. The validation of the model was carried out using experimental data. Hence, we propose a robust calibration procedure, based on good modeling practice, that encompasses both sensitivity and uncertainty analyses, and therefore provides a basis for the optimization of the IX process with the aim of improving the accuracy of breakthrough prediction.

## Keywords

Calibration protocol, fixed-bed column, global sensitivity analysis, ion exchange, uncertainty analysis

## Highlights

1. Model parameters most affecting breakthrough are revealed by sensitivity analysis.
2. Breakthrough prediction improved by calibrating resin capacity and particle size.
3. Proper calibration of parameter subsets is reliant on uncertainty of the estimates.
4. Uncertainty quantification provides reliability bands for breakthrough prediction.
5. Calibration protocol is applicable to other physico-chemical adsorption processes.

## Abbreviations

AIC	Akaike Information Criterion
CI	Confidence Interval
FMI	Fisher Information Matrix
GSA	Global Sensitivity Analysis
IX	Ion Exchange
LSA	Local Sensitivity Analysis
ODE	Ordinary Differential Equation
PDE	Partial Differential Equation
RMSE	Root Mean Square Error
WSSE	Weighted Sum of Square Error

## 1. Introduction

The global issue of water scarcity is placing increasing pressure on the supply and management of water resources. As a result, resource recovery has emerged as a strategy to increase the circularity of resources and water supplies. Ion exchange (IX) is a key technology in this endeavor, as it enables fit-for-purpose water demineralization through its ion-selective recovery properties (Kabdaşlı and Tünay, 2018; Jegatheesan et al., 2021; Taghvaie Nakhjiri et al., 2022). IX, like other adsorption processes, is typically operated as a continuous process in fixed-bed packed columns and used as an end-of-pipe removal treatment (Inglezakis and Zorpas, 2012). Notable applications include the recovery of nutrients and by-products in the food production industry (Kammerer et al., 2011); the removal of heavy metals such as nickel from urban wastewater (Ma et al., 2019), petrochemical wastewater (Cechinel et al., 2018) and other industrial waters such as chromium-rich textile waste (Wang et al., 2015); the recovery of precious metals from industrial wastewater (Taghvaie Nakhjiri et al., 2022); the removal of silica for the production of ultrapure water in various industries, including paper mills and electronics (Chen et al., 2022); or the removal of pharmaceuticals (Chu and Hashim, 2023a) and micropollutants such as PFAS in drinking water production (Smith et al., 2023).

The challenges of the IX technology include the optimization of the cost of chemicals associated with resin regeneration and the disposal of generated waste streams (Crittenden et al., 2012), as well as the accurate prediction of target ion breakthrough times for downstream compliance during operation and control (Inglezakis and Zorpas, 2012). Consequently, the optimization of IX processes would allow for a reduction in the energetic and material impacts of water treatments, thereby enhancing their environmental and economic sustainability and further increasing their potential for resource recovery.

A number of mechanistic and empirical models have been developed with the objective of improving the understanding of the IX process and addressing the aforementioned challenges by identifying the breakthrough time. A myriad of these models can be found in the literature, including transport and dispersion mechanisms derived from conservation laws, such as surface diffusion (Ma et al., 2019), pore diffusion (Zhang et al., 2015), and other intraparticle diffusion kinetic models (Wang and Guo, 2022). Similarly, numerous equilibrium isotherm models have been developed beyond the classical Langmuir and Freundlich models. A comprehensive review of the most commonly used models can be found in LeVan and Carta (2008), and more recently in Wang and Guo (2023) and Wang et al. (2024).

Therefore, in view of the considerable number of existing models, no unified modeling procedure exists, and as a result, a standardized calibration protocol for adsorption models is still missing. This is evidenced by the numerous existing reports on inconsistencies, mistakes, and misconceptions in the modeling of the IX process (Chu, 2023; Hauptert et al., 2021; Hu et al., 2021; Lima et al., 2021; Mudhoo and Pittman, 2023; Tran et al., 2017; Xiao et al., 2018), which result in models with limited reliability and prediction power. The following five common misconceptions in modeling tasks were identified: (i) overfitting, (ii) use of linearized models, (iii) false sensitivity analyses, performed locally and for single parameters, (iv) absence of uncertainty quantification, and (v) confusion between the goal of sensitivity analyses and uncertainty analyses, as previously highlighted by Saltelli et al. (2019).

A review of the above articles from literature essentially shows that in IX, models can be effectively used to predict ion breakthrough in a multitude of applications. However, the calibration methodologies employed by various authors exhibit inconsistencies in rigor resulting in overfitting, limited reproducibility and high uncertainty regarding the predictions of these models. Consequently, these models frequently fail when subjected to validation or extrapolation to future time series, due to the considerable uncertainty associated with the calibrated parameters. The limited extrapolation capability, coupled with the complexity and non-linear nature of the operation, restricts the utility of these models as a monitoring and control tool for water treatment applications. A standardized model calibration procedure is currently missing.

In order to achieve reliable and powerful prediction capabilities and establish good modeling practice for the calibration and reproducibility of IX models, we followed general recommendations by Saltelli et al. (2019) regarding sensitivity and uncertainty analysis, which are valid across disciplines. Similar guidelines have been successfully established for the calibration of diverse processes in the wastewater field (Rieger et al., 2012; Vanrolleghem et al., 2003). In a recent study, Chu and Hashim (2023b) employed rigorous model selection techniques to evaluate the performance of competing models with varying numbers of fitting parameters. Nevertheless, essential considerations in the calibration process, such as verification of the model structure, a detailed description and uncertainty quantification of the parameter estimation procedure, and the analysis of the data quality and resulting

performance, are still absent in numerous studies. Our work is based on the model proposed by Zhang et al. (2015), which considered an advection-diffusion-reaction model for fixed-bed ion exchange columns. This allowed us to address a few of the aforementioned limitations and to propose a framework for model calibration based on good modeling practice.

This work is accompanied by open-access code (<https://github.com/UGentBiomath/IX-GMP>) and presents a framework for model calibration based on good modeling practice. The work includes a description of a simple one-component IX model for fixed-bed operation and employs sensitivity and uncertainty analyses to verify the model structure and facilitate rigorous calibration of an identifiable parameter set. The resulting protocol is applicable to other IX modeling studies and allows for scenario analysis and optimization of the system under study.

## 2. Materials and Methods

### 2.1 Model definition

We implemented a dynamic model describing the transport of ions through a fixed-bed IX column reactor. We selected a two one-dimensional partial differential equation (PDE) model by Zhang et al. (2015) in order to develop our model. The first PDE describes the liquid phase, while the second PDE describes the solid diffusion. As the goal of this modeling study is to optimize the process, computational efficiency is a significant factor. Consequently, we simplified the model with the following modifications: the simplified model considers the same one-dimensional set of equations, comprising an advection-dispersion-reaction partial differential equation (PDE) for the liquid phase, but proposes a linear driving force mass transfer ordinary differential equation (ODE) for the solid diffusion, as proposed by LeVan and Carta (2008). Therefore, we replaced the second PDE with an ODE.

Both models compute the concentration profile of ions in the liquid and solid phases along the column, thereby predicting the breakthrough of these ions following saturation of the resin. A visual inspection of Figure 7 shows that our simplified model using parameters from Zhang et al. (2015) (designated as “uncalibrated” in the figure) does indeed produce the anticipated breakthrough curve of the data presented by Zhang et al. (2015). Therefore, the simplified model can be employed as a fast, accurate tool to improve the efficiency of IX operations. This is made possible by the fast concurrent computation of both scales of the process, namely the ion concentration in the liquid and solid phases, which are respectively referred to as the macro and micro-scale by Zhang et al. (2015). Subsequently, both scales are translated into a single output, namely the ion transport in the fluid along the column. Similarly, the solid phase can be analyzed for the purposes of regeneration, although this is beyond the scope of the present work.

#### 2.1.1 Model structure

Figure 1 depicts the IX process as a fixed-bed resin-packed column, illustrating the main variables involved in the dynamic transport of ions along the column and through the liquid and solid phases.

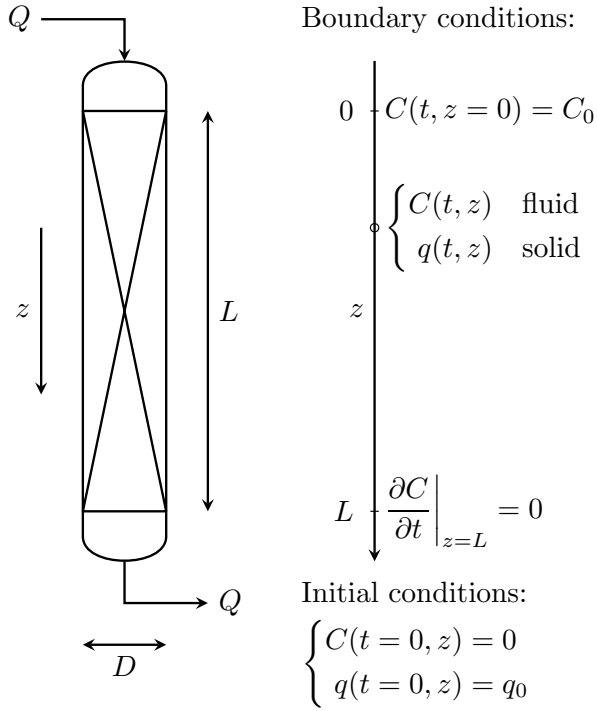


Figure 1. Schematic representation of a typical fixed-bed IX column and description of the main variables of the 1-D model with appropriate initial and boundary conditions. The parameters are detailed in Table 1.

The evolution of the concentration profiles of a given ion in the column can be obtained by performing a mass balance. This results in the PDE (1), which must be solved after the appropriate initial and boundary conditions have been defined. Furthermore, the solution involves the discretization of the space and time domains:

$$\frac{\partial C}{\partial t} = D_z \frac{\partial^2 C}{\partial z^2} - u \frac{\partial C}{\partial z} + \frac{\rho_b}{\varepsilon} \frac{\partial q}{\partial t} \quad (1)$$

In the above expression,  $C$  represents the concentration of ions present in the liquid phase at any given point. The variable  $q$  denotes the concentration of ions in the solid phase. The term  $u$  is the fluid velocity, which is assumed to be constant.  $D_z$  is the axial liquid dispersion coefficient.  $\rho_b$  is the bulk bed density, while  $\varepsilon$  denotes the bed porosity. In this context, the independent variables  $t$  and  $z$  are used to refer to time and column height, respectively. Furthermore, the following assumptions have been made: a uniform velocity profile and intraparticle diffusion in the column; and a controlling equilibrium between the liquid and solid phases (negligible resistance to mass transfer in the boundary layer, as indicated by Zhang et al., 2015). The velocity can be calculated by assuming a constant inlet flow and a constant void bed cross-section in the column, according to the following equation:  $u = \frac{Q}{A\varepsilon}$ . The last term in Equation (1) denotes the change in ion concentration due to adsorption or desorption in the solid phase. Assuming a linear driving force between the liquid and solid phases (LeVan and Carta, 2008), the second equation of the model can be expressed as an ordinary differential equation at each discretized point within the spatial domain:

$$\frac{\partial q}{\partial t} = k_{ldf}(q^* - q) \quad (2)$$

where  $k_{ldf}$  is the mass transfer coefficient and  $q^*$  is the concentration in the resin phase surface in equilibrium with the liquid phase concentration,  $C$ , which can be calculated using an equilibrium isotherm model. Equations (1) and (2) must be integrated and solved concurrently.

The two most frequently utilized equilibrium isotherm models in literature are the Freundlich and Langmuir models (O'Neal and Boyer, 2013). The Freundlich isotherm model (Freundlich, 1907) assumes multilayer adsorption described by a power law:

$$q^* = K_f \cdot C^{n_f} = K_f \cdot C^{1/n} \quad (3)$$

where  $K_f$  and  $n_f$  are parameters to be calibrated. The Langmuir model (Langmuir, 1918) assumes monolayer adsorption, with  $q_{max}$  representing the maximum adsorption capacity, and  $K_L$  a second parameter to be calibrated:

$$q^* = q_{max} \frac{K_L C}{1 + K_L C} \quad (4)$$

In this study, we used the Langmuir isotherm model due to the greater interpretability of its parameters. Nevertheless, the Freundlich model has also been successfully applied to describe the equilibrium between two phases (Sengupta and Pandit, 2011; O'Neal and Boyer, 2013). It should be noted that other, more complex models exist; however, the inclusion of additional parameters complicates the calibration process and is therefore not considered here.

The mass transfer coefficient  $k_{ldf}$  in Equation (2) is a function of the intraparticle diffusion coefficient,  $D_p$ , and the particle radius,  $r_p$ , according to a pore or solid diffusion mechanism (LeVan and Carta, 2008). This can be expressed as follows:

$$k_{ldf} = 15 \frac{D_p}{r_p^2} \quad (5)$$

The solid and liquid phases are related through the bed density given by  $\rho_b = \rho_p(1 - \varepsilon)$ , where  $\rho_p$  is the resin density and  $\varepsilon$  is the void fraction in the column left by the settled resin, also known as bed porosity.

### 2.1.2 Model parameters and variables

Table 1 lists the variables considered for the present model structure, classified according to their function: output variable, input variable, or parameter. The latter can be further subdivided into the following categories: design parameters, which include column dimensions and the physicochemical properties of the resin; operational parameters; and equilibrium parameters. The last column indicates the source of the values: whether they were measured experimentally, calculated numerically, fixed by

the process or equipment specifications related to design decisions and reported in manufacturers' data sheets, or obtained from the literature. Parameters for which there is no direct measurement or other means of obtaining a value are estimated from output measurements in the calibration process. For each parameter, the values reported in the literature are listed in Table 2, together with the ranges and sources. Even those parameters which have been assumed to be constant for design reasons (manufacturer's data sheets, operating rules), are considered uncertain for the purposes of the sensitivity analysis, in order to provide a comprehensive overview of potential model uncertainties.

The liquid phase concentration is frequently determined at the inlet ( $C_0$ ) and outlet ( $C$ ) of the column. Direct measurement of the concentration profile within the column is not feasible without introducing multiple sample extractions, which would affect the total column volume. However, these profiles are predicted by the model. The measurement of solid-phase concentrations is challenging; however, they can be calculated through a mass balance by means of Equation (2). The initial solid phase concentration,  $q_0$ , is assumed to be zero for fresh resin. As the original resin capacity is never fully restored, this initial condition could also be regarded as a variable in cases where regeneration is being investigated.

Table 1. Variables and parameters considered in the simplified IX model. The symbols are in accordance with the commonly used nomenclature in the literature. For detailed values, see Table 2.

Name	Symbol	Unit	Type	Source
<b>Outputs</b>				
Liquid-phase concentration	$C$	mmol/L		Computed by Eq. (1)
Solid-phase concentration	$q$	mmol/g		Computed by Eq. (2)
<b>Inputs</b>				
Inlet liquid-phase concentration	$C_0$	mmol/L		Directly measured*
Initial solid-phase concentration	$q_0$	mmol/g		Initial condition in Eq. (2)
Volumetric flow	$Q$	L/min		Manipulated/fixed*
<b>Parameters</b>				
Bed length	$L$	m	Design	Directly measured*
Bed diameter	$D$	m	Design	Column manufacturer*
Resin bead particle size	$r_p$	m	Design	Resin manufacturer*
Resin bead particle density	$\rho_p$	g/L	Design	Resin manufacturer*
Bed porosity (void fraction)	$\varepsilon$	–	Operation	Literature*
Axial dispersion coefficient	$D_z$	m <sup>2</sup> /s	Operation	Literature*
Intraparticle diffusion coefficient	$D_p$	m <sup>2</sup> /s	Operation	Literature*
Sorption capacity	$q_{max}$	mmol/g	Equilibrium	Literature*
Langmuir constant	$K_L$	L/mmol	Equilibrium	Literature*

\*Determination of these parameters is uncertain and therefore could be considered for calibration.

The design parameters include the bed dimensions, which determine the volume available for both resin and effluent in the column, as well as the particle size and the density of the solid phase (inversely proportional to the specific volume occupied by the resin). The effective volume available for the liquid fraction is a function of the bed porosity, which is in turn determined by the compaction of the resin in the column and the swelling induced by the liquid. This parameter has been extensively studied, and a

range of variability can be found in the literature depending on the packing shape and size (see Table 2). The actual liquid capacity of the bed is calculated as follows:  $V = \varepsilon LA = \varepsilon L \frac{\pi}{4} D^2$ , with the height or length ( $L$ ) of the bed directly measured, as it is not always the case that the full column length is utilized. In the literature, the bed volume is typically referred to as a measure of the time elapsed during the process, calculated as the treated effluent volume per unit time divided by the effective bed volume. In addition, dispersion coefficients are frequently encountered in the literature in the form of empirical correlation functions of the Reynolds number and bed configuration. The determination of intraparticle coefficients is an experimental process, and they are therefore considered to be uncertain.

Table 2. Values and ranges of variability reported in the literature for the parameters listed in Table 1.

	Reported value	Variability range	Source
<b>Design parameters</b>			
Bed diameter, $D$	0.1 m	(±20%)	Zhang et al., 2015
Bed length, $L$	$10^{-2}$ m	(±20%)	Zhang et al., 2015
<b>Operational parameters</b>			
Inlet P concentration, $C_0$	20 mmol/L	(±20%)	Zhang et al., 2015
Flow, $Q$	3.1 cm/min	(±20%)	Zhang et al., 2015
Porosity, $\varepsilon$	0.37	0.3–0.44 (±20%)	Yoshida et al., 1985
Resin size (diameter), $d_p = 2r_p$	$7.5 \cdot 10^{-4}$ m	$3\text{--}12 \cdot 10^{-4}$ m (±60%)	Sengupta and Pandit, 2011
Resin density, $\rho_p$	389 g/L	(±20%)	O’Neal and Boyer, 2013
Intraparticle diffusivity, $D_p$	$5.3 \cdot 10^{-10}$ m <sup>2</sup> /s*	(±20%)	Sengupta and Pandit, 2011
Maximum capacity, $q_{max}$	0.291 mmol/g*	(±20%)	O’Neal and Boyer, 2013
Langmuir constant, $K_L$	1.18 L/mmol*	(±20%)	O’Neal and Boyer, 2013

\* Values for synthetic fresh urine.

### 2.1.3 Model implementation and numerical solution

The model equations have been formulated as a set of one-dimensional differential equations, thereby enabling the model to be employed as a fast optimization tool for the IX process. It is assumed that the velocity profiles of the fluid across the column are uniform and that there is no significant existence of wide preferential channels due to the low diameter-to-length ratio ( $D/L$ ) of the column.

In order to predict the one-dimensional spatial variation of pollutant concentration along the IX column, a discretization of the column length was considered. The solution of the nonlinear system of equations can be computationally intensive. Therefore, a discretization error analysis was conducted (see Appendix for a detailed discussion) and an appropriate discretization step of 100 grid points and a time step of 0.1 seconds was selected to achieve sufficient accuracy without substantial computational effort.

The model was implemented in Python 3 (Van Rossum and Drake, 2009) and makes use of available scientific packages *Numpy* (Harris et al., 2020), *Scipy* (Virtanen et al., 2020), *Pandas* (McKinney, 2010; The pandas development team, 2020), and *Matplotlib* (Hunter, 2007). The code will be made available at <https://github.com/UGentBiomath/IX-GMP>.

The equations were solved with the *scikit-finite-diff* package (Cellier and Ruyer-Quil, 2019) using the finite difference method and the method of lines for the spatial and temporal discretization of the PDE. Figure 2a illustrates the concentration profiles along the length of the column as a function of time in relation to the inlet concentration,  $C_0$ . The arrow indicates the typical temporal evolution of the profiles resulting from the transport of ions along the column and into the solid phase. The breakthrough curve (Figure 2b) reflects the evolving ion concentration in the effluent at the column outlet, illustrating the gradual depletion of the resin and the breakthrough of the ion, which is adsorbed onto the solid phase until the resin is completely saturated.

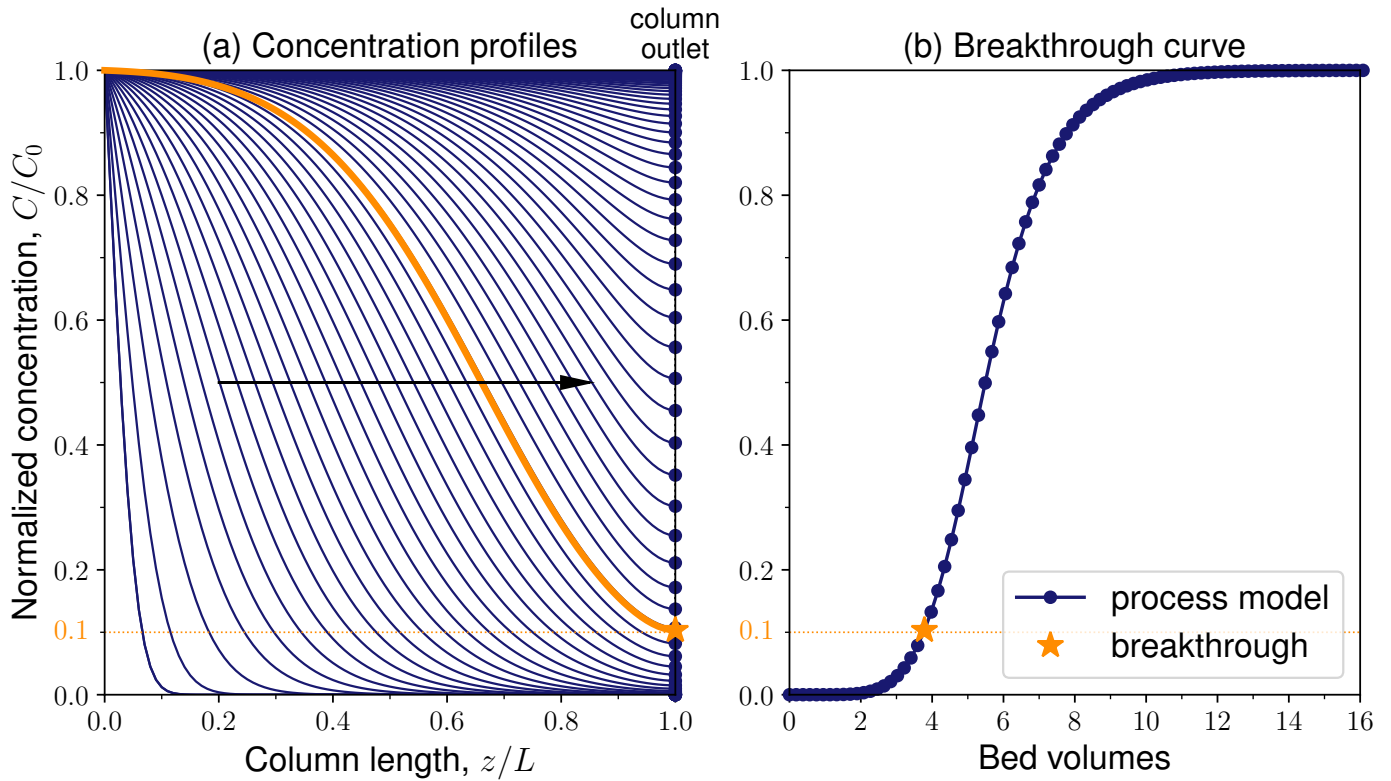


Figure 2. Concentration profiles (a) and breakthrough curve (b) obtained by solving the model equations (1) to (4) with the appropriate initial and boundary conditions as specified by the scheme in Figure 1. The breakthrough time (highlighted) is defined as the time for 10% of initial concentration in the outlet.

## 2.2 Sensitivity analysis

The objective of the sensitivity analysis is to identify those parameters that exert the greatest influence on model output, with the aim of reducing the variability or uncertainty in the model output by accurately determining their values. By identifying which parameters are sensitive, it is possible to reduce the cost of experiments by focusing efforts on measuring those parameters (Saltelli et al., 2007).

### 2.2.1 Local sensitivity analysis

A local sensitivity analysis (LSA) quantifies which model parameters have a greater influence on the model output(s) for a given value of each parameter (Saltelli et al., 2007). The sensitivity function is defined as the change in the model output resulting from a small change or perturbation in the value of a single parameter. A numerical approximation of the sensitivity function, commonly referred to as the derivative method, is the first-order forward finite approximation:

$$S_{\theta_i}^j = \frac{\partial y_j}{\partial \theta_i} \cdot \frac{\theta_i}{y_j} \approx \frac{y_j(\theta_i + \Delta\theta_i) - y_j(\theta_i)}{\Delta\theta_i} \cdot \frac{\theta_i}{y_j} \quad (6)$$

where  $y_j$  represents the model output,  $\theta_i$  denotes the parameter subject to variation, and  $\Delta_i$  is the perturbation value. In the present study, a perturbation value of  $10^{-5} \cdot \theta$  was used for all parameters. The sensitivity function is normalized to accommodate disparate scales or magnitudes for both the output and the parameters. It is important to note that the results of a local sensitivity analysis may vary significantly depending on the specific location within the parameter space under investigation. Hence, the analysis may be repeated at different locations to obtain further information, or a global sensitivity analysis may be performed (see section 2.2.2).

The local sensitivity analysis is employed as an initial model check on the parameters' behavior, requiring minimal computational efforts and offering insight into the extent to which varying parameters influence the model output. Furthermore, it identifies potential correlations between parameters and operational regions where parameter sensitivity is higher and correlation minimal. These regions are of particular interest for experimental data collection.

### 2.2.2 Global sensitivity analysis

A global sensitivity analysis (GSA) aims to quantify the relative importance of parameters in determining the variability of a model output over a wide range of parameter values (Saltelli et al., 2007). In contrast to a local analysis, a systematic exploration of the parameter space can provide a more comprehensive understanding of the overall influence of the different parameters on the model output. To this end, sensitivity indices can be calculated in order to identify these effects.

First-order indices describe the main effect of parameters on the variance of model outputs. They can be calculated as the variability in model output resulting from the variation of a single parameter in isolation, relative to the total variability attributable to changes in all parameters, expressed as follows:

$$S_i = \frac{V[E(Y|X_i)]}{V(Y)} \quad (7)$$

where  $V$  represents the variability of model output  $Y$ ,  $E$  denotes the expected value, and  $X_i$  is a specific parameter value. In contrast, total effects comprise the cumulative impact of a parameter on the model output, accounting for both first-order and higher-order effects stemming from interactions or non-linearities between parameters. A disparity between total and first-order effects indicates the existence of higher-order interactions:

$$S_{T_i} = 1 - \frac{V[E(Y|X_{\sim i})]}{V(Y)} \quad (8)$$

The GSA is calculated at a specific time point. In the present study, the time at which 10% breakthrough is observed was selected as the most informative for GSA. The sensitivity indices for the 10%

concentration breakthrough times with respect to the model parameters listed in Table 2 were calculated by Sobol uniform sampling using the *SALib* sensitivity analysis library (Herman and Usher, 2017). Furthermore, second-order sensitivity indices were calculated and employed to ascertain the existing correlation between parameters. The Sobol sampling of parameters consisted of 5,632 uniform values, selected within the ranges of variability outlined in Table 2. A 20% variability was considered for all parameters, including  $r_p$ . Subsequently, a Monte Carlo-type of simulation was employed to ascertain the variability in model output derived from the simulation results corresponding to each parameter subset sampling. The primary disadvantage of GSA is its high computational cost, which is a consequence of the substantial number of model evaluations required. One advantage of an LSA over a GSA is that it can facilitate the preliminary screening and reduction of the parameter set prior to a more comprehensive global analysis.

Once the most sensitive model parameters have been identified, they can be estimated through the process known as model calibration. In order to achieve this, it is necessary to obtain experimental data.

### 2.3 Experimental data for calibration and validation

The experimental data used for the calibration of our model are derived from measurements of breakthrough curves for ion exchange of phosphorus recovery from fresh urine, as documented by O'Neal and Boyer (2015). The data set describes the breakthrough of phosphate ions from synthetic fresh urine as it traverses a fixed-bed column containing a specific phosphate-selective HAIX-Fe resin. Further details regarding the column tests can be found in the referred work.

The authors acknowledge a limitation in the data set, namely the absence of measurements of sulphate and other ions present in the urine effluent. These ions compete with phosphates in their adsorption onto the resin; however, the analysis of competition and affinity effects is beyond the scope of the present work and not captured by the model. Therefore, the influence of these ions on the predictive power of the model will not be addressed. The required parameters for the modeling of this system are enumerated in Table 2, accompanied by the sources from which they were derived. As illustrated by Figure 7, the predicted breakthrough curve exhibits a high degree of agreement with the measured data. Further details can be found in the Results and Discussion section.

### 2.4 Model calibration

The goal of model calibration is to identify the optimal set of parameter values that fit the experimental data, which can be considered as an optimization problem. The quality of the fit is quantified by an objective function, which is then minimized. The weighted sum of squared errors (WSSE) was selected as the objective function for model calibration:

$$J(\theta) = \sum_{i=1}^N (\hat{y}_i(\theta) - y_i)^T W (\hat{y}_i(\theta) - y_i) \quad (9)$$

where  $\hat{y}_i(\theta)$  represents the model prediction of the output  $y_i$  corresponding to the parameter set of values  $\theta$ ,  $y_i$  denotes the measurement of the output  $y$ ,  $W$  is the square matrix of weights associated

with different outputs or time moments, and  $N$  is the number of measurements. In this analysis, we assumed  $W$  to be the identity matrix, thereby assigning equal weight to each time point of the single measured model output.

There are numerous techniques for minimizing the objective function, including the well-known Nelder-Mead simplex method (Nelder and Mead, 1965). The Levenberg-Marquardt method (Levenberg, 1944; Marquardt, 1963) was selected as a robust and efficient method well suited for unconstrained nonlinear least squares fitting problems. Further details regarding the implementation can be found in Gavin (2019). A recent overview of calibration methods for computer simulation is provided by Sung and Tuo (2024).

Unconstrained optimization methods may suggest values for the parameters that are outside of their range of physical validity. Consequently, we also employed a robust constrained trust region method (Branch et al., 1999) to evaluate the performance of constrained parameter subsets in comparison to that of the unconstrained Levenberg-Marquardt algorithm. Both constrained and unconstrained methods are readily available in the optimization library of the *Scipy* Python package (Virtanen et al., 2020). For the constrained optimization method, the parameter values were bounded by their ranges of validity. The initial parameter estimates, along with their respective ranges of variability, are presented in Table 2. The relative tolerance, which serves as a stopping criterion for the iterative optimization algorithm, was set to  $10^{-4}$  in order to reduce the number of model evaluations. This value was selected following a verification process, during which it was determined that it produced results that were not significantly different from those obtained with lower tolerance. Furthermore, the scale or magnitude of each parameter was provided to the algorithm, thereby reducing the number of iterations. The accuracy of the solution was evaluated by comparing the model output to the experimental data provided by Zhang et al. (2015).

## 2.5 Uncertainty of the parameter estimation

The estimated parameters have an associated uncertainty due to the presence of noise in the experimental data set used for calibration, and the degree of uncertainty is dependent on the relative importance of this imperfect information in the model structure. Accordingly, parameter estimates are only meaningful when accompanied by their confidence region, within which the actual true value is situated at a specified confidence level (Donckels, 2009). The confidence region can be based on an approximation of the contour of the WSSE objective function, as this provides a measure of the fit to the experimental data. In the case of linear models, the contour is exact. However, for nonlinear models, it is common practice to employ a linear approximation of the parameter estimation covariance matrix, providing a lower bound for this region (Marsili-Libelli et al., 2003):

$$\{\theta : WSSE(\theta) \leq c \cdot WSSE(\hat{\theta})\} \tag{10}$$

A linear approximation results in the Fisher Information Matrix (FIM), whose inverse is employed for the approximation of the error covariance matrix, can be computed as follows (Dochain and Vanrolleghem, 2001):

$$FIM = \sum_{i=1}^N S_{\theta}' \cdot W^{-1} \cdot S_{\theta} = \sum_{i=1}^N \left( \frac{\partial \hat{y}}{\partial \theta}(t_i) \right)' \cdot W^{-1} \cdot \left( \frac{\partial \hat{y}}{\partial \theta}(t_i) \right) \quad (11)$$

where  $W$  is the covariance matrix of the measurement errors associated with the measured variables, and  $S_{\theta}$  denotes the parameter sensitivity matrix of all outputs with respect to each parameter, as calculated by Equation (5) for  $N_p$  parameters and  $N$  experimental time points. The approximated error covariance matrix can be used to construct a confidence region for the parameter estimates,  $\delta_i$ , with a specified level of confidence  $1-\alpha$  (Marsili-Libelli et al., 2003):

$$\delta_i = t_{N-N_p}^{\alpha/2} \cdot \sqrt{\sigma_{i,i}^2} \quad (12)$$

where  $t$  represents the  $\alpha/2$  quartile of the Student's  $t$  distribution for a given confidence level  $\alpha$  and  $N - N_p$  degrees of freedom, with  $N$  denoting the number of data points,  $N_p$  the number of estimated parameters, and  $\sigma_{i,i}^2$  the variance of parameter  $i$  taken from the error covariance matrix. The diagonals of the covariance matrix thus provide the variances of the errors in the parameter estimates, whereas the off-diagonal elements are the covariances of the errors and offer a measure of the correlation between the different parameters. The linear correlation between two parameters can be estimated based on the following equation:

$$r_{i,j} = \frac{\sigma_{i,j}}{\sqrt{\sigma_{i,i}^2 \cdot \sigma_{j,j}^2}} \quad (13)$$

The linear correlation is approximately -1 (negative) or 1 (positive) for pairs of parameters with a high degree of correlation, whereas a value of approximately zero indicates a low correlation.

Once the confidence region for the parameter values has been established, the Monte Carlo technique can be employed to sample the parameters within the region and thereby obtain the expected variability in the model output. A normal distribution was assumed for all parameters, and a total of 640 values were sampled for use in the Monte Carlo simulations. The plotting of the most frequent values of the model output can assist in establishing an uncertainty band around the mean value, thereby providing a confidence interval band for the prediction of the breakthrough curve.

## 2.6 Fitness comparison

The root mean square error (RMSE) was employed as a metric for assessing the goodness of fit, or the discrepancy between the experimental data and the calculated breakthrough concentrations. This was due to the fact that the RMSE represents a more accurate average of the distance between the data and the model prediction than the WSSR of the objective function. The RMSE can be calculated as follows:

$$RMSE = \sqrt{\frac{\sum(y_i - \hat{y}_i)^2}{N}} = \sqrt{\frac{WSSR}{N}} \quad (14)$$

The RMSE offers a straightforward and readily understandable representation of the overall model's error, using the same units as the measured variable, even for unitless comparisons. On the other hand, the RMSE is sensitive to both outliers and overfitting; consequently, it diminishes when additional parameters are incorporated into the model. In order to facilitate a comparative analysis of model fitness for different numbers of calibrated parameters, it is possible to consider criteria that balance the goodness of fit to experimental data with the number of model parameters. This approach allows to penalize overfitting when increasing parameters are incorporated into the model structure. One such established method is Akaike's Information Criterion or AIC (Akaike, 1974):

$$AIC = N \cdot \log\left(\frac{WSSR}{N}\right) + 2 \cdot N_p \quad (15)$$

In Equation (15), the first term will decrease for overparametrized candidate models due to overfitting, while the second term will penalize the added complexity when more parameters are considered. A lower value of AIC is indicative of a superior model, whereas a low WSSR value is indicative of overfitting.

## 2.7 Model validation

To confirm the predictive power of the calibrated model, a new data set was selected from the same column breakthrough experiments as the calibration tests (see Section 2.3 for a description of the data). For calibration, the parameter  $q_0$  in our model was assumed to be zero, corresponding to fresh resin. The second cycle was chosen to validate the model and therefore the measured data correspond to a regenerated column. Hence, we assumed a regeneration efficiency of 95% for the resin, which was calculated as the amount of phosphorus recovered from the total adsorbed in the column, and the remaining amount was assumed to be retained or fouled in the resin, as also reported by the authors (O'Neal and Boyer, 2015; Zhang et al., 2015). From the adsorption data, fouling is estimated at 1.7 mg out of the total 31.9 mg adsorbed, resulting in a 5.3% fouled amount or 94.7% regeneration efficiency. We then assumed an initial concentration profile in the resin, parameterized by  $q_0$  in our model.

# 3. Results and Discussion

## 3.1 Sensitivity analysis

### 3.1.1 Local sensitivity analysis

The specific set of model parameters to be analyzed is presented in Table 2. The parameter  $q_0$  was omitted from the list, as it is assumed to be equal to zero in this calibration data set. Figure 3(a) shows the variation over time, expressed in bed volumes, of the relative sensitivity values for the breakthrough concentration with respect to all parameters in the selected subset, with the most sensitive parameters highlighted. The less sensitive parameter  $D_p$  is also highlighted as a reference. As can be seen, the

sensitivity to changes in parameter values is practically zero at the start of the column operation and increases exponentially when the first breakthrough is detected, around two bed volumes. A peak in sensitivity is reached for all parameters around four to six bed volumes and decreases again exponentially until the end of the operation when the resin is saturated. The occurrence of peaks for several parameters at approximately the same time is a clear indication of a correlation between these parameters. This correlation will make it more challenging to obtain reliable values when these parameters are estimated simultaneously. Since some sensitivities appear to change sign over time, an averaged sensitivity allows to compare the overall trend. Figure 3(b) shows the averaged local sensitivity values over 16 bed volumes for all parameters with their corresponding signs.

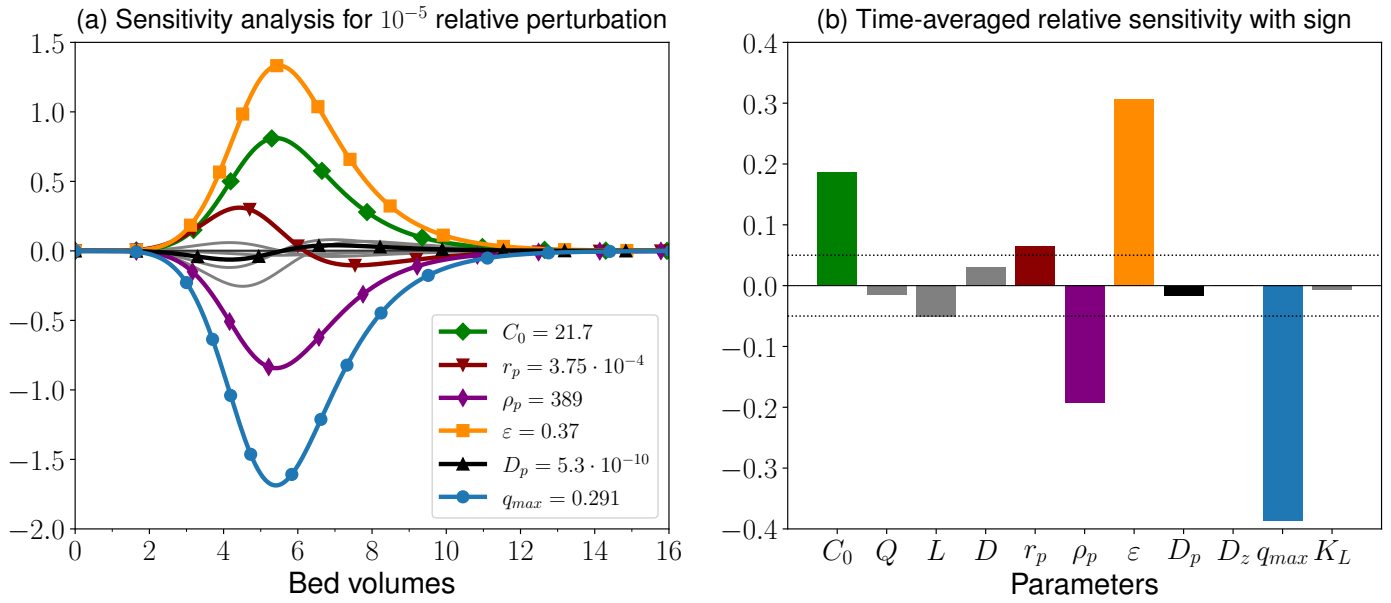


Figure 3. Local sensitivity analysis (LSA) of the breakthrough curve for the parameters listed in Table 1: time variation in bed volumes for a  $10^{-5}$  relative perturbation (a), and time average with sign (b). The most sensitive parameters ( $q_{max}$ ,  $\epsilon$ ,  $\rho_p$ ,  $C_0$ ,  $r_p$ ) are highlighted, with  $r_p$  and  $D_p$  negatively correlated.

In descending order of sensitivity, the most sensitive parameters are  $q_{max}$ ,  $\epsilon$ ,  $\rho_p$ , and  $C_0$ . Most of these parameters exhibit positive or negative sensitivity to varying degrees, indicating that they affect the model output in a consistent manner but with differing magnitudes. However, parameters  $r_p$  and  $D_p$  exert an inverse influence on breakthrough, as evidenced by their inverse correlation, illustrated in Figure 3(a) and in accordance with Equation (5). The particle size is more than twice as sensitive as the intraparticle diffusivity due to the quadratic influence, with the greatest effect observed around four bed volumes, where both parameters are most sensitive.

Changes in sensitivities are also indicative of nonlinearity. In order to illustrate the impact of varying sensitivities, it is possible to plot the effect of a change in different parameters on the model output. Figure 4 shows the impact of a 10% increase in four of the most sensitive parameters and its effect on the breakthrough concentration. An increase in a parameter with negative sensitivity (e.g.,  $q_{max}$ ) will result in a delay in breakthrough, as it has a negative influence on the concentration. Conversely, a positive sensitivity (e.g., with respect to  $\epsilon$  or  $C_0$ ) will result in an earlier occurrence of breakthrough for an increase in the parameter value. Figure 4 also confirms that  $C_0$  is a less sensitive parameter than  $\epsilon$ , indicating that a 10% increase in the latter will have a more pronounced effect on breakthrough. An

analogous analysis may be performed for the remaining, less sensitive parameters. As can be observed, a change in the less sensitive parameter  $r_p$  is most pronounced around four bed volumes, where the parameter exhibits the greatest sensitivity to changes, as illustrated in Figure 3. However, the overall effect of this one parameter is minor in comparison to an increase in the adsorption capacity,  $q_{max}$ , as demonstrated by Figure 3: an increase in the bed height (parameter  $L$ ) would result in greater resin availability too, which in turn delays breakthrough. However, this increase has overall less effect than a comparable change in the adsorption capacity. This serves to illustrate once more the nonlinear nature of the process and the importance of taking into account these effects for the estimation of parameters.

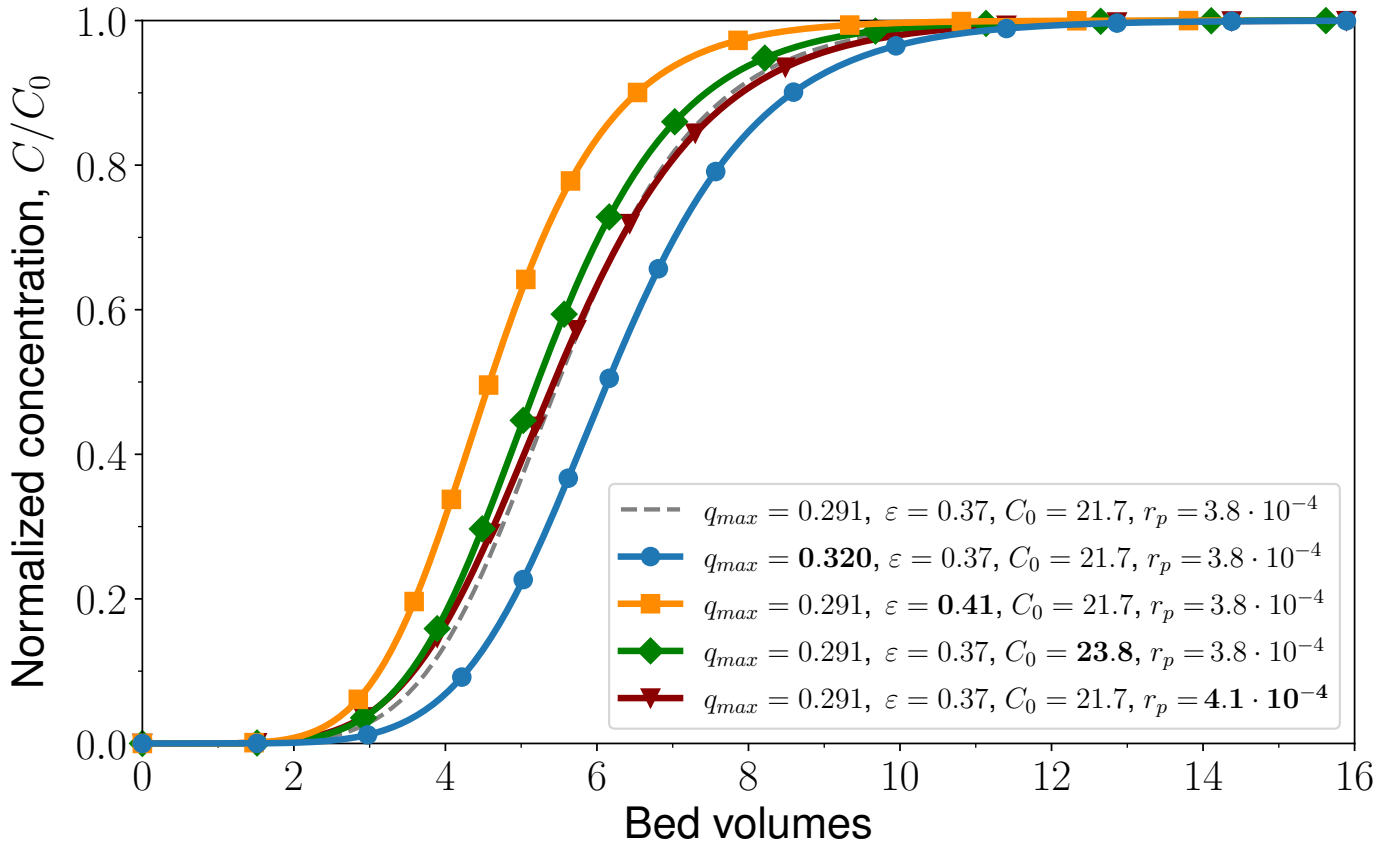


Figure 4. Effect of a 10% increase in the most sensitive parameters ( $q_{max}$ ,  $\varepsilon$ ,  $C_0$  and  $r_p$ ) on the breakthrough curve: the adsorption capacity ( $q_{max}$ ) has a marked negative impact on the breakthrough concentration, while the other three parameters have a positive effect, albeit with varying magnitudes.

Following Figure 3 and Figure 4, we can establish a local ranking of the parameter sensitivities for the model structure considered. It should be noted, however, that this is only a local ranking and should therefore be interpreted and used with care. In the case of nonlinear models, parameter sensitivities may vary considerably when evaluated in different regions of the parameter space. Accordingly, a global sensitivity analysis is conducted in the following section to ascertain an overall parameter ranking for the model calibration. The results of the LSA serve two distinct purposes. First and foremost, the LSA results offer insights into the parameter sensitivities as a function of time, thereby enabling the identification of an optimal time point for conducting a GSA analysis. Based on the LSA analysis, the time at which a 10% breakthrough occurs is identified as an informative time instant to perform the GSA.

Secondly, the LSA results are employed to conduct an initial screening of the parameters, whereby parameters exhibiting low sensitivity can be excluded from a subsequent analysis. As the GSA analysis is considerably more computationally intensive, a preliminary screening through LSA can significantly accelerate the global analysis. The parameters exhibiting the lowest local sensitivity are  $K_L$ ,  $Q$ ,  $D_p$  and  $D_z$ . As the inlet flow rate  $Q$  is a variable that can be manipulated, it was decided that it should be retained for a comprehensive analysis in order to ascertain its influence. Furthermore, the isotherm parameter  $K_L$  is frequently calibrated in conjunction with  $q_{max}$ , and thus it was also determined that it should be included in the GSA. However, in the event of computational constraints, this category of parameters may be excluded from the subsequent analysis, given their negligible impact. It is therefore possible to assign a fixed value within their range of variability to those parameters that have little influence on the output and produce little variance, without compromising the estimation process for the remaining parameters.

A sensitivity analysis may also serve to identify the experimental conditions that will yield the most accurate parameter estimation by maximizing the effect on the model output, which is the goal of *optimal experimental design* or OED (Donckels, 2009). The objective may be to generate experimental data with high information content, thereby further reducing the uncertainty of the parameter estimates. The LSA analysis conducted in the present study indicates that experiments where the breakthrough concentration surpasses at least 50% of the initial concentration represent the most informative experiments, exhibiting the highest parameter sensitivity. Nevertheless, the optimization of the experimental design for parameter estimation falls outside of the scope of the present work.

### 3.1.2 Global sensitivity analysis

#### *Parameter subset selection for GSA*

Based on the LSA results, parameter  $D_z$  was excluded from the GSA: its quasi-zero sensitivity corresponds to the common plug-flow assumption and the reason why the diffusion term in Equation (1) is often neglected in practice. Figure 5 illustrates the Sobol sensitivity indices for 10% concentration breakthrough times across the ten remaining parameters. Both the total and first-order sensitivity indices are plotted for each parameter, accompanied by 95% confidence bands. First-order sensitivity indices are analogous to local sensitivities, but they are calculated over the entire parameter space. As can be observed,  $\varepsilon$  and  $q_{max}$  are the parameters that exert the greatest influence on the breakthrough curve based on both the total and first-order indices. The maximum adsorption capacity,  $q_{max}$ , has a smaller effect than anticipated by LSA when the full range of parameter values is considered. In contrast, the bed porosity,  $\varepsilon$ , has overall a more pronounced effect on the breakthrough concentration than anticipated by LSA. This is due to the fact that a slight increase in the value of  $\varepsilon$  results in a reduction of the resin available in the column, which in turn leads to a decrease in the overall adsorption efficiency. The other equilibrium parameter, the Langmuir constant,  $K_L$ , has a negligible effect on the breakthrough concentration across the entire range of values considered. Consequently, the equilibrium parameter  $q_{max}$ , which is considerably more sensitive, should be given precedence in model calibration. Lastly, we selected an arbitrary cut-off value of 0.1 for parameters exhibiting minimal global sensitivity. The design parameters,  $D$  and  $L$ , and the inlet flow,  $Q$ , have low indices, indicating that these parameters could be excluded from the calibration process initially. Nevertheless, the resin

density and particle size parameters demonstrated a notable degree of local sensitivity, as illustrated in Figure 3. Accordingly, the outcomes of the GSA should be interpreted with caution.

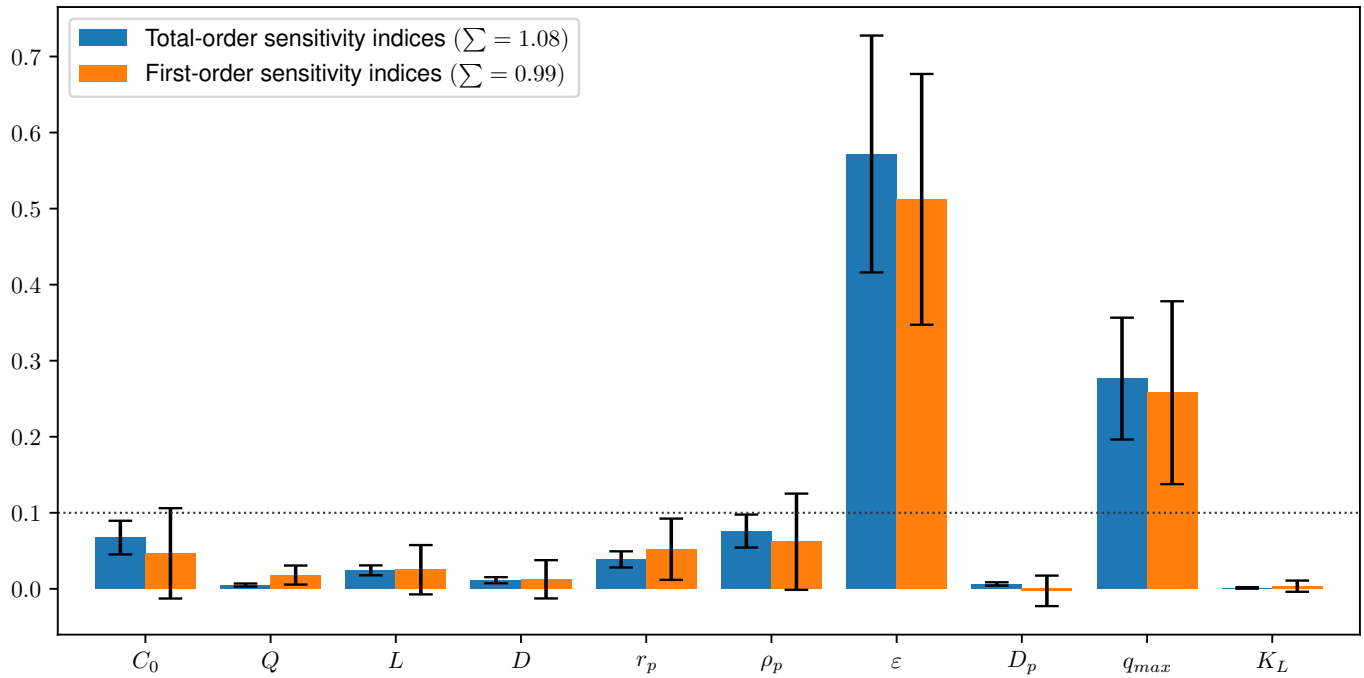


Figure 5. Sobol global sensitivity indices for the parameters listed in Table 2: total-order indices (left), and first-order indices (right). The analysis was performed for a 10% concentration breakthrough time. A variability of 20% for all parameters was considered.

Figure 5 additionally illustrates the total-order sensitivity indices for each parameter. The observation that the sum of the total and first-order indices is, respectively, 1.08 and 0.99, with an uncertainty indicated by the 95% confidence bands, suggests that this model exhibits some degree of nonlinearity. This is also evidenced by the discrepancy between total-order and first-order effects, which suggests the presence of higher-order interactions or correlations between parameters. The parameters exhibiting the most significant discrepancy between total and first-order indices indicate the highest degree of interaction with other parameters. The *SALib* library further allows the calculation of second-order sensitivity indices with supplementary parameter values sampled, thus necessitating additional computational resources and effort. Figure 6 depicts these second-order sensitivity indices, which illustrate the interactions between pairs of parameters. The largest interactions are observed for both isotherm parameters and the intraparticle diffusivity with the bed porosity, the particle size, and the resin density. This indicates that all these parameters have a high degree of correlation and may exert a particularly strong influence on the adsorption process. This strong correlation between parameters was also evident in the single point of the parameter space depicted in Figure 3, with the majority of parameters exhibiting a peak in sensitivity at approximately the same time instant, following breakthrough and preceding saturation. A strong correlation complicates the simultaneous estimation of multiple parameters, since correlated parameters are non-identifiable (Dochain and Vanrolleghem, 2001). Nevertheless, an example of parameter exhibiting comparatively less correlation with the remaining parameters is that of the intraparticle diffusion,  $D_p$ , with the notable exception of  $r_p$  and  $\epsilon$ , with which it displays a significant degree of correlation. Furthermore, the remaining parameters also exhibit some degree of interaction, particularly the inlet concentration,  $C_0$ , although to a lesser extent.

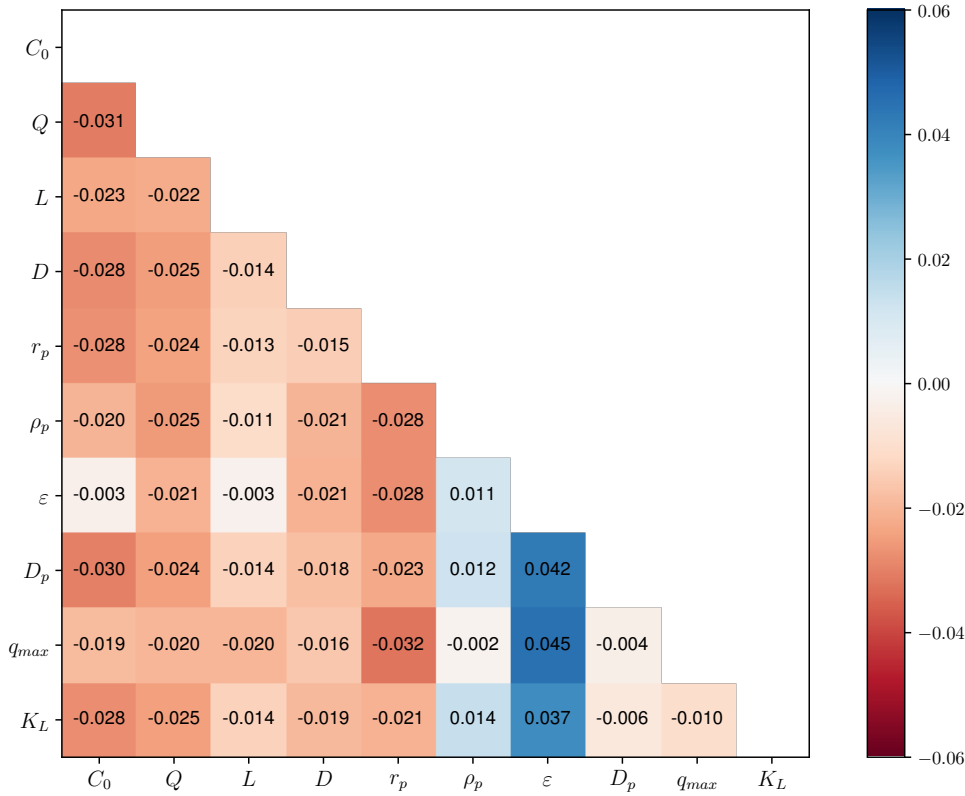


Figure 6. Second-order sensitivity indices. Parameters  $\varepsilon$ ,  $q_{max}$ ,  $K_L$ ,  $D_p$ ,  $r_p$  show the largest interactions.

### Parameter subset selection for calibration

In light of the results of the GSA presented in Figure 5 and Figure 6, it becomes evident that only a limited subset of parameters will undergo calibration. A subset of parameters for calibration can be selected based on the most sensitive parameters. Consequently, the first parameter to be estimated would be  $\varepsilon$ , as this exerts the greatest influence on the breakthrough concentration. Furthermore, it can be observed that  $q_{max}$  exerts a considerable impact on the breakthrough concentration. A calibration subset comprising the two most sensitive parameters could be formed. Both  $\rho_p$  and  $r_p$  have low indices, yet Figure 3 indicates that both parameters exhibit high local sensitivities. Accordingly, both parameters could also be included in a calibration subset. The parameters  $D_p$  and  $K_L$  exert an overall negligible influence on the breakthrough concentration and should therefore not be calibrated initially. Moreover, we assumed that the inlet concentration,  $C_0$ , remains constant in this study. The design parameters  $L$  and  $D$  can be excluded, given their low sensitivity indices and the fact that their values are frequently fixed for existing columns. With regard to the manipulable variable  $Q$ , its value is typically fixed for operational reasons. Therefore, only the parameters  $\varepsilon$ ,  $q_{max}$ ,  $\rho_p$  and  $r_p$  will be considered for calibration.

## 3.2 Model calibration

In accordance with the findings of the GSA, we conducted a stepwise calibration for the model parameters, beginning with the calibration of the most sensitive parameter and subsequently incorporating an additional parameter into the calibration set. For instance, we initially calibrated  $\varepsilon$ , followed by the simultaneous calibration of  $\varepsilon$  and  $q_{max}$ . This process was then repeated for subsequent parameters. For each calibration exercise, the confidence intervals of the parameters were calculated. Both the RMSE and AIC values are provided as fitness criteria. The results are summarized in Table 3.

Table 3. Results of the parameter estimation for up to four parameters in different calibration subsets. The values indicate the estimated parameter values, the corresponding 95% confidence intervals (relative percentage) calculated with FIM, RMSE and AIC as fitness criteria. Initial estimates given by  $\theta_0$ .

$N_p$ <sup>a</sup>	$N_f$ <sup>b</sup>	Min. <sup>c</sup>	$\varepsilon$	$q_{max}$	$\rho_p$	$r_p$	RMSE	AIC
1	22, 10	LM, TR	<b>0.35 ± 1.8%</b>				<b>0.0233</b>	<b>-148</b>
	19, 12	LM, TR		<b>0.323 ± 1.4%</b>			<b>0.0261</b>	<b>-144</b>
	23, 10	LM, TR			463 ± <b>25.8%</b>		<b>0.0300</b>	<b>-138</b>
	9, 8	LM, TR				3.4·10 <sup>-4</sup> ± <b>34.8%</b>	<b>0.0857</b>	<b>-96</b>
2	31, 21	LM, TR	<b>0.31 ± 30.6%</b>	<b>0.243 ± 22.1%</b>			0.0207	-151
	25	LM	<b>0.30 ± 30.0%</b>		<b>248 ± 42.9%</b>		0.0208	-151
	24	TR	<b>0.32 ± 32.5%</b>		<b>311 ± 48.0%</b>		0.0213	-150
	15	LM, TR	<b>0.35 ± 1.9%</b>			<b>4.1·10<sup>-4</sup> ± 8.1%</b>	<b>0.0207</b>	<b>-151</b>
	21	LM <sup>d</sup>		0.280 ± <b>10<sup>5</sup>%</b>	<b>517 ± 10<sup>5</sup>%</b>		<b>0.0261</b>	<b>-142</b>
	15	TR <sup>d</sup>		0.312 ± <b>10<sup>5</sup>%</b>	417 ± <b>10<sup>5</sup>%</b>		<b>0.0261</b>	<b>-142</b>
	15	LM, TR		<b>0.326 ± 1.4%</b>		<b>4.3·10<sup>-4</sup> ± 8.4%</b>	<b>0.0206</b>	<b>-151</b>
3	<b>44</b>	TR	0.31 ± <b>11.7%</b>	0.258 ± <b>10<sup>6</sup>%</b>	344 ± <b>10<sup>6</sup>%</b>		0.0207	<b>-149</b>
	<b>35</b>	LM	0.33 ± <b>10<sup>2</sup>%</b>	0.275 ± <b>10<sup>2</sup>%</b>		4.0·10 <sup>-4</sup> ± <b>10<sup>2</sup>%</b>	0.0206	<b>-149</b>
	<b>39</b>	TR	0.36 ± <b>10<sup>2</sup>%</b>	0.312 ± <b>10<sup>2</sup>%</b>		4.2·10 <sup>-4</sup> ± <b>10<sup>2</sup>%</b>	0.0205	<b>-149</b>
4	<b>71</b>	TR <sup>d</sup>	0.35 ± <b>10<sup>7</sup>%</b>	0.301 ± <b>10<sup>7</sup>%</b>	389 ± <b>10<sup>7</sup>%</b>	4.2·10 <sup>-4</sup> ± <b>10<sup>7</sup>%</b>	0.0207	<b>-147</b>
	<b>35</b>	TR <sup>d</sup>	0.35 ± <b>10<sup>7</sup>%</b>	0.301 ± <b>10<sup>7</sup>%</b>	466 ± <b>10<sup>7</sup>%</b>	4.2·10 <sup>-4</sup> ± <b>10<sup>7</sup>%</b>	0.0207	<b>-147</b>
$\theta_0$			0.37	0.291	389	3.8·10 <sup>-4</sup>	0.0864	-
LM			-	-	-	-		
TR			0.30-0.44 (±20%)	0.233-0.349 (±20%)	311-467 (±20%)	3.0·10 <sup>-4</sup> -4.5·10 <sup>-4</sup> (±20%)		

<sup>a</sup> Number of model parameters in the calibration subset.

<sup>b</sup> Number of respective function evaluations of the minimization algorithm.

<sup>c</sup> Minimization algorithm. LM: Levenberg-Marquardt (unconstrained); TR: trust region method (constrained).

<sup>d</sup> The substantial CIs are attributable to the inability to estimate the covariance matrix (FIM matrix is singular).

The calibration of only the most sensitive parameter, namely  $\varepsilon$ , resulted in a 73% improvement in fit (RMSE of 0.0233 vs 0.0864) compared to the initial uncalibrated value of 0.37 given by  $\theta_0$  at the bottom of Table 3. The 95% confidence interval (CI) was estimated at ±1.8%, expressed as a relative percentage of the optimal value. After approximately 10 to 20 model evaluations, both the Levenberg-Marquardt method (unconstrained parameter values) and the trust reflective region method (parameter values are constrained to the bounds indicated at the bottom of Table 3) yielded identical results. In comparison, the calibration of the following less sensitive parameter,  $q_{max}$ , yielded a confidence interval of ±1.4%, which can be attributed to the fact that it has a higher local sensitivity compared to  $\varepsilon$ . Moreover, the determined value of 0.323 is more closely aligned with the total phosphate loading on the resin of 10.2 mg P/g or 0.329 mmol P/g, as reported by O'Neal and Boyer (2015). The remaining, less sensitive parameters  $\rho_p$  and  $r_p$  yielded a comparatively inferior fit when calibrated separately, as evidenced by the increased values for the confidence intervals and both RMSE and AIC fitness criteria.

Subsequently, a second parameter was incorporated into the calibration set. This allowed for the simultaneous estimation of both parameters, beginning with their initial values as presented at the bottom of Table 3. The estimation of both  $\varepsilon$  and  $q_{max}$  resulted in a further reduction in the root mean square error (RMSE) to 0.0207, representing an 11% decrease. However, both minimization methods

yielded considerably low values in comparison to those reported in the literature, and the quality of the estimation also declined, as evidenced by the increased confidence intervals. This can be attributed to the significant correlation between both parameters, which was calculated to be as high as 99.8%, as illustrated in Figure 3. Moreover, estimating both  $\varepsilon$  and  $\rho_p$  did not yield enhanced results, as this last parameter is considerably less sensitive than  $q_{max}$ . This was demonstrated in both Figure 3 and Figure 5, and is further corroborated by the calculated correlation coefficient, which reached a value of 99.8%. However, when estimating both  $\varepsilon$  and  $r_p$  simultaneously, despite the calculated correlation of 44.5%, the estimated value and uncertainty for  $\varepsilon$  remained largely unchanged. Furthermore, the calculated 95% confidence interval for this second parameter was 8.1%, indicating that both parameters can be estimated with a high degree of confidence. In comparison, the simultaneous calibration of both  $q_{max}$  and  $r_p$  resulted in an uncertainty of 1.4% and 8.4%, respectively. An alternative calibration subset comprising parameters  $q_{max}$  and  $\rho_p$  yielded unreliable estimates, likely due to their correlation.

As illustrated in Table 3, the calibration of additional parameters results in considerably larger confidence intervals for all parameters, consequently leading to a notable increase in the uncertainty associated with the estimated values. Such overparameterization does not result in a significant increase in the fit to the experimental data, as the introduction of additional, less sensitive parameters necessitates substantially larger alterations in their values to produce a change in the objective function. Moreover, this also dramatically increases the number of required iterations for convergence of the minimization algorithm. As a result, the estimation is rendered ill-conditioned. For instance, this is substantiated by the relatively higher (less negative) values of AIC, which are -149 and -147, with the lowest value of -151 corresponding to the calibration of only two parameters. The simultaneous calibration of three or four parameters results in a further deterioration in the precision of the estimation, as evidenced by the considerably broader confidence intervals for all parameters, including those that were previously estimated with a high degree of accuracy (11.7% for  $\varepsilon$  vs 1.8% when it is estimated together with  $q_{max}$  and  $\rho_p$ ). This phenomenon occurs when the selected minimization method yields disparate solutions within the parameter space, with less sensitive parameters estimated at the expense of the certainty in the value of the most sensitive ones. Furthermore, the estimation of less sensitive parameters provides only minimal information, resulting in unreliable estimates due to a poor approximation of the covariance matrix (Vugrin et al., 2007).

Figure 7(a) illustrates the fit of the model predictions to the experimental data of Zhang et al. (2015), showcasing both calibrated and uncalibrated parameter values. The calibrated values for parameter  $q_{max}$  from Table 3 are slightly higher than the initial reported values in the literature, resulting in a rightward shift of the breakthrough curve with respect to the uncalibrated model. Figure 7(b) depicts the residuals, defined as the absolute difference between the experimental data and both the calibrated and uncalibrated model predictions. As can be observed, calibration essentially reduces the residuals in the zone where the calibrated parameters are most sensitive, namely after breakthrough, around 4-10 bed volumes. This is the case for all parameter subsets, although beyond two calibrated parameters, the residuals are almost indistinguishable. The calibration of additional parameters does not significantly improve the fit to the experimental data, due to the phenomenon of overfitting or overparameterization. Furthermore, the uncalibrated model displays a tendency to overpredict the breakthrough concentration. However, all models exhibit a comparable limitation in accurately

reproducing the initial time instants of the experimental data. This is a consequence of the low sensitivity of the parameters in this zone, which renders the model less powerful in its ability to capture this aspect of the curve for the utilized data.

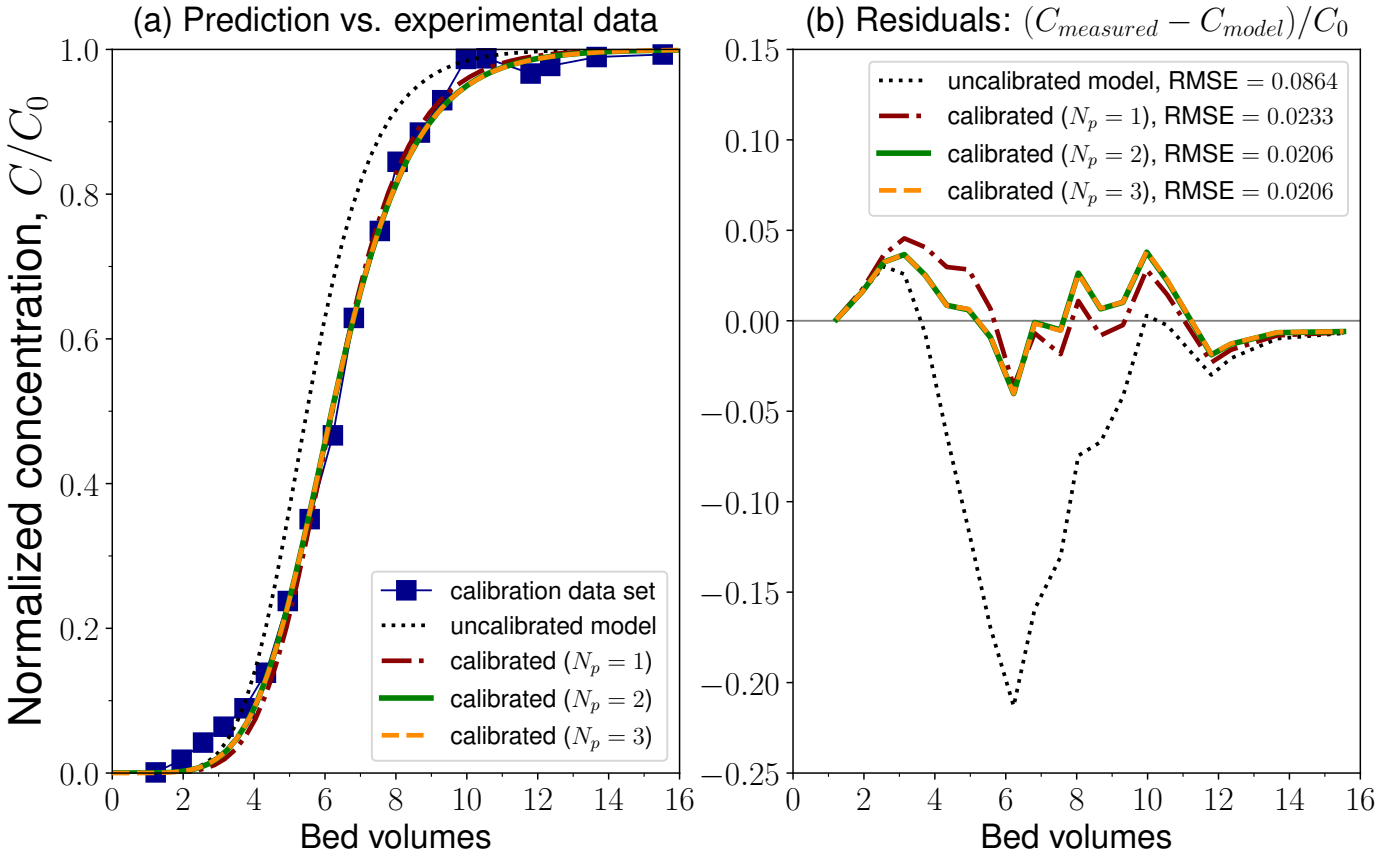


Figure 7. Calibration of up to three model parameters ( $q_{max}$ ,  $r_p$ ,  $\varepsilon$ ): model fit to experimental data (a) and residual calculation (b). Data for fresh urine by Zhang et al. (2015). Decreasing RSME is a better fit.

The optimal model, as determined by the lowest value of the Akaike Information Criterion (AIC) in Table 3, is obtained by calibrating only two of the most sensitive parameters. This may be, for example, the bed porosity,  $\varepsilon$ , and the particle size,  $r_p$ , or alternatively, the maximum adsorption capacity isotherm parameter,  $q_{max}$ , and the particle size,  $r_p$ , since  $\varepsilon$  and  $q_{max}$  are correlated. This example illustrates the importance of exercising caution when estimating parameters with low sensitivity, as for subsets of more than two parameters, there exist infinite combinations of parameters that produce the same fit, which significantly reduces the predictive capacity of the model. Hence, these results confirm that only the most sensitive model parameters should be included in the calibration process. Furthermore, a robust calibration protocol is essential, where sensitivity and correlation of parameters are evaluated prior to calibration, to ensure the development of a reliable model with minimal uncertainty and optimal predictive power.

### 3.3 Uncertainty analysis

In addition to the quality of the parameter estimation provided by the confidence regions from Table 3, an uncertainty analysis of the model with respectively two and three calibrated parameters is presented in Figure 8. The calibrated model output of the breakthrough curve is enclosed by a 95% confidence

band. As illustrated, the uncertainty in the prediction is minimal at the initial and final stages of the operation but increases after breakthrough at approximately 2-4 bed volumes and subsequently decreases near the saturation point at around 10-12 bed volumes. Consequently, the width of the uncertainty band is dependent upon the quality of the model calibration step and thus determines the reliability of the model prediction. For two calibrated parameters ( $q_{max}$  and  $r_p$ ,  $N_p = 2$ ), the uncertainty band is narrow and closely surrounds the calibration data set, indicating a highly accurate prediction. However, while the initial instants of the process were not accurately captured by the model, the prediction of breakthrough closely follows the calibration data. Further reasoning was provided in Section 3.2. In the case of calibrated parameters  $q_{max}$  and  $\varepsilon$ , the uncertainty band becomes broad due to the higher uncertainty resulting from the correlation of these two parameters. For three calibrated parameters ( $N_p = 3$ , as illustrated in Figure 8), the uncertainty band surrounding the breakthrough prediction is markedly broader, thereby demonstrating the inferior calibration. The lowest uncertainty is thus obtained with the calibration of parameters  $q_{max}$  and  $r_p$ .

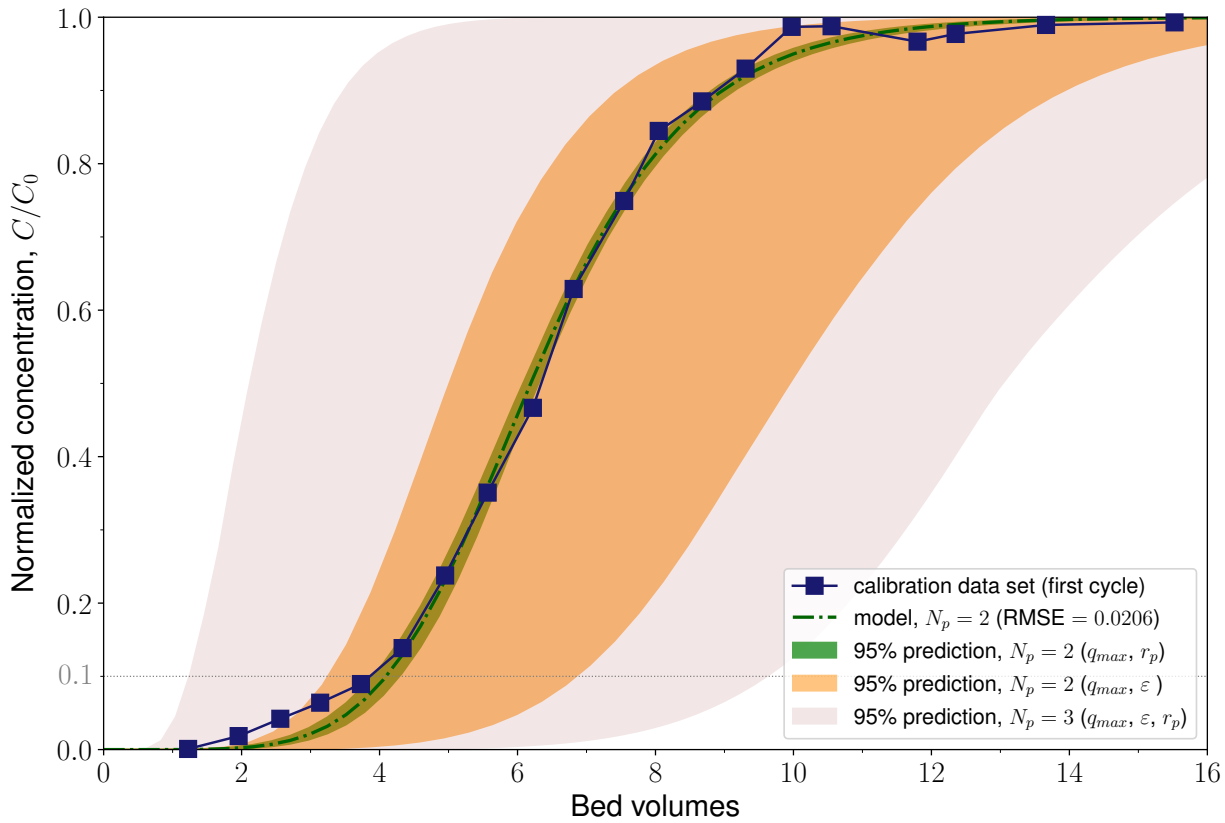


Figure 8. Uncertainty analysis of the breakthrough curve for a 95% confidence in the prediction interval. The uncertainty bands for two and three calibrated parameters are generated by the corresponding confidence intervals of Table 3. The lowest uncertainty is obtained with two parameters ( $q_{max}$  and  $r_p$ ).

### 3.4 Model validation

Once the model has been calibrated with the accurate determination of the most sensitive parameters, the predictive power of the model can be tested against a validation data set. Figure 9 depicts the predicted breakthrough curve of a second cycle of the process, which demonstrates substantial agreement with the experimental data as reported by Zhang et al. (2015).

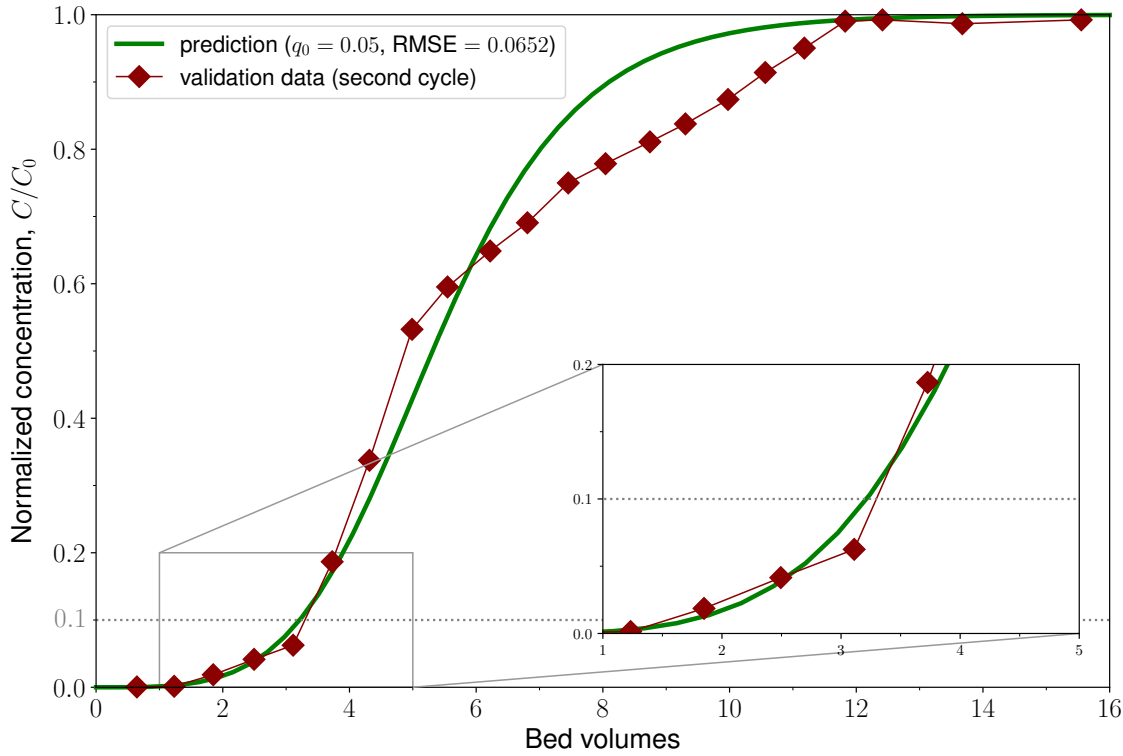


Figure 9. Model validation for the 95% regenerated resin using column data from (Zhang et al., 2015). The inset shows that the initial time instants around 10% breakthrough are well captured by the model.

As illustrated by the inset, the initial time instants of the experiment are accurately represented by the model, with a precise prediction (error of 1.0%) of the 10% breakthrough time occurring at approximately 3.2 bed volumes (see Table 4). For the calibration set, the predicted 10% breakthrough time was in good agreement with the measured value of 3.8 bed volumes, with an error of 7.8%. As expected, the breakthrough time decreases as a consequence of resin fouling. Moreover, the saturation time, defined as the time required for a breakthrough concentration of 90% of the initial concentration, also decreases with the accumulation of fouling. These trends were also observed in the prediction of a third cycle with a 50% fouled resin (results not shown), with good agreement between prediction and experimental data. However, the discrepancy between the predicted and actual values at the saturation point is more pronounced in comparison to the breakthrough times. The potential causes of this discrepancy include interactions between the adsorbed ions onto the resin, as postulated by O’Neal and Boyer (2015), which could result in an increased adsorption capacity when the resin is close to saturation. This may, in turn, result in a delayed breakthrough of the ions. It seems plausible that this trend may persist following additional regeneration cycles. However, further analysis is necessary to substantiate this hypothesis, which is beyond the scope of the present study.

Table 4. Breakthrough and saturation times (in bed volumes) from the model and the experimental data. The first cycle corresponds to the calibration data set and the second cycle to the validation data set.

	Breakthrough time, $t_{10}$			Saturation time, $t_{90}$		
	Data	Model	Rel. error	Data	Model	Rel. error
First cycle (calibration)	3.8	4.1	7.8%	8.8	9.0	2.2%
Second cycle (5% fouled, validation)	3.2	3.2	1.0%	10.3	8.1	21.0%
Third cycle (50% fouled, not illustrated)	2.4	2.2	6.7%	9.3	7.2	22.6%

## 4. Conclusions

We present a framework for calibrating a dynamic model for a fixed-bed column IX operation based on good modeling practice that can be used as a reference for future modeling studies and practical model implementations. The local and global sensitivity analyses allowed us to identify the design and operational parameters that contribute most to the prediction of breakthrough curves. Specifically, the local sensitivity analysis revealed which time intervals during IX operation provide the most information for model calibration, thus allowing the selection of the most informative time instances for further analysis as well as enabling an initial screening to identify highly sensitive parameters. Subsequently, the global sensitivity analysis allowed us to select a limited subset of parameters for calibration. This approach showed that only two parameters, namely the maximum adsorption capacity isotherm parameter and the resin particle size, which were identified as the most sensitive parameters based on the GSA, require comprehensive calibration to achieve an accurate prediction of the breakthrough curve. Moreover, our findings demonstrated that the inclusion of additional, less sensitive parameters results in a reduction in the reliability of the parameter estimates, since the parameters become less identifiable (as illustrated by the AIC model selection criterion). Additionally, we demonstrated the effect of parameter estimation uncertainty on the model output by propagating the parameter uncertainty through the model, which showed that the inclusion of insensitive parameters in the calibration significantly increased the uncertainty of the model output. The model was validated using experimental data. In light of these findings, we propose a robust calibration procedure, based on good modeling practice, that encompasses both sensitivity and uncertainty analyses, and therefore provides a basis for the optimization of the IX process with the aim of improving the accuracy of breakthrough prediction.

## Acknowledgements

We would like to express our gratitude to Benjamin Claessens for his input on the model development at the start of the project, and to Ivaylo Hitsov Plamenov for sharing his expertise on the ion exchange process.

## Funding

This work was supported by Ghent University. MYS was supported by the Swiss National Science Foundation (P500PT\_211132).

# Appendix

## Discretization analysis with respect to numerical parameters

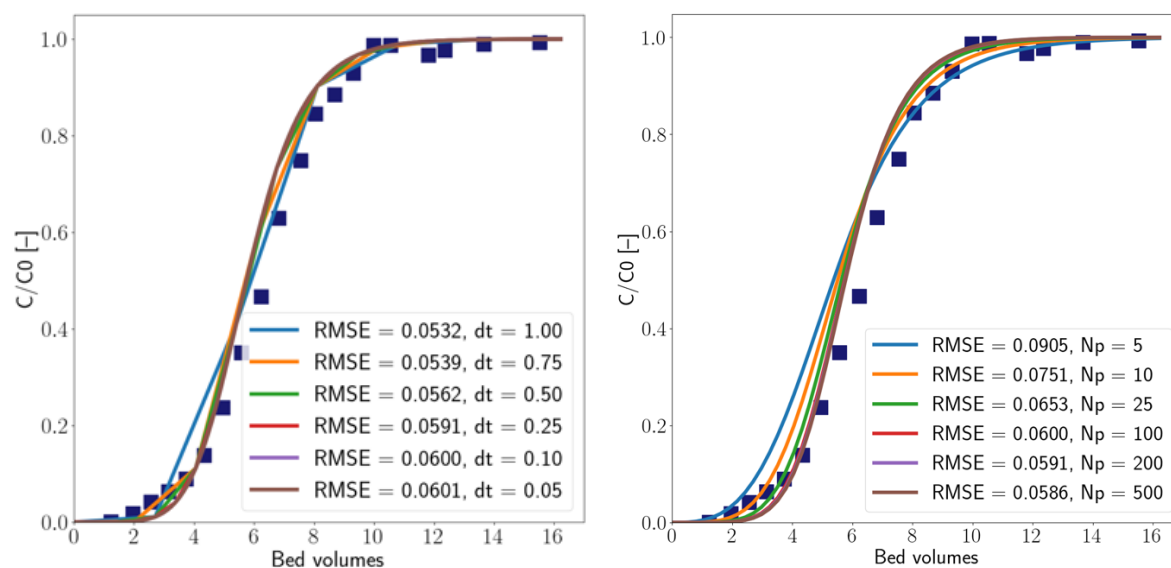


Figure S1. Discretization analysis of model output (breakthrough concentration) with respect to numerical parameters: time step (left) and number of discretized column elements (right). A time step  $dt = 0.1$  and  $N_p = 100$  elements provide sufficient accuracy in the numerical solution to predict breakthrough without significantly adding computational effort. This was assessed by subsequently performing simulations with time steps of 1, 0.75, 0.5, 0.25, 0.1 and 0.05 seconds with 100 grid points, followed by 5, 10, 25, 100, 200 and 500 grid points with a 0.1 second time step, and observing indistinguishable simulation results. All simulations require a computation time of under 1 minute.

## References

- Akaike, H., 1974. A new look at the statistical model identification. *IEEE Trans. Autom. Control* 19, 716–723. <https://doi.org/10.1109/TAC.1974.1100705>
- Branch, M.A., Coleman, T.F., Li, Y., 1999. A Subspace, Interior, and Conjugate Gradient Method for Large-Scale Bound-Constrained Minimization Problems. *SIAM J. Sci. Comput.* 21, 1–23. <https://doi.org/10.1137/S1064827595289108>
- Cechinel, M.A.P., Mayer, D.A., Mazur, L.P., Silva, L.G.M., Girardi, A., Vilar, V.J.P., de Souza, A.A.U., Guelli U. de Souza, S.M.A., 2018. Application of ecofriendly cation exchangers (*Gracilaria caudata* and *Gracilaria cervicornis*) for metal ions separation and recovery from a synthetic petrochemical wastewater: Batch and fixed bed studies. *J. Clean. Prod.* 172, 1928–1945. <https://doi.org/10.1016/j.jclepro.2017.11.235>
- Cellier, N., Ruyer-Quil, C., 2019. scikit-finite-diff, a new tool for PDE solving. *J. Open Source Softw.* 4, 1356. <https://doi.org/10.21105/joss.01356>
- Chen, S., Bai, S., Ya, R., Du, C., Ding, W., 2022. Continuous silicic acid removal in a fixed-bed column using a modified resin: Experiment investigation and artificial neural network modeling. *J. Water Process Eng.* 49, 102937. <https://doi.org/10.1016/j.jwpe.2022.102937>

- Chu, K.H., 2023. Fixed Bed Adsorption of Water Contaminants: A Cautionary Guide to Simple Analytical Models and Modeling Misconceptions. *Sep. Purif. Rev.* 52, 75–97. <https://doi.org/10.1080/15422119.2022.2039196>
- Chu, K.H., Hashim, M.A., 2023a. Adsorptive removal of pharmaceutical contaminants: Accurate description of tailing breakthrough curves. *J. Environ. Chem. Eng.* 11, 111025. <https://doi.org/10.1016/j.jece.2023.111025>
- Chu, K.H., Hashim, M.A., 2023b. Removal of antibiotics through fixed bed adsorption: Comparison of different breakthrough curve models. *J. Water Process Eng.* 56, 104512. <https://doi.org/10.1016/j.jwpe.2023.104512>
- Crittenden, J.C., Trussell, R.R., Hand, D.W., Howe, K.J., George Tchobanoglous, 2012. Ion Exchange, in: *MWH's Water Treatment: Principles and Design, Third Edition*. John Wiley & Sons, Ltd, pp. 1263–1334. <https://doi.org/10.1002/9781118131473.ch16>
- Dochain, D., Vanrolleghem, P.A., 2001. Dynamical modelling and estimation in wastewater treatment processes. IWA Publ, London.
- Donckels, B., 2009. Optimal experimental design to discriminate among rival dynamic mathematical models (dissertation). Ghent University.
- Freundlich, H., 1907. Über die Adsorption in Lösungen. *Z. Für Phys. Chem.* 57U, 385–470. <https://doi.org/10.1515/zpch-1907-5723>
- Gavin, H.P., 2019. The Levenberg-Marquardt algorithm for nonlinear least squares curve-fitting problems. *Dep. Civ. Environ. Eng. Duke Univ.* August 3.
- Harris, C.R., Millman, K.J., van der Walt, S.J., Gommers, R., Virtanen, P., Cournapeau, D., Wieser, E., Taylor, J., Berg, S., Smith, N.J., Kern, R., Picus, M., Hoyer, S., van Kerkwijk, M.H., Brett, M., Haldane, A., del Río, J.F., Wiebe, M., Peterson, P., Gérard-Marchant, P., Sheppard, K., Reddy, T., Weckesser, W., Abbasi, H., Gohlke, C., Oliphant, T.E., 2020. Array programming with NumPy. *Nature* 585, 357–362. <https://doi.org/10.1038/s41586-020-2649-2>
- Hauptert, L.M., Pressman, J.G., Speth, T.F., Wahman, D.G., 2021. Avoiding pitfalls when modeling removal of per- and polyfluoroalkyl substances by anion exchange. *AWWA Water Sci.* 3, e1222. <https://doi.org/10.1002/aws2.1222>
- Herman, J., Usher, W., 2017. SALib: An open-source Python library for Sensitivity Analysis. *J. Open Source Softw.* 2, 97. <https://doi.org/10.21105/joss.00097>
- Hu, Q., Pang, S., Wang, D., Yang, Y., Liu, H., 2021. Deeper Insights into the Bohart–Adams Model in a Fixed-Bed Column. *J. Phys. Chem. B* 125, 8494–8501. <https://doi.org/10.1021/acs.jpccb.1c03378>
- Hunter, J.D., 2007. Matplotlib: A 2D Graphics Environment. *Comput. Sci. Eng.* 9, 90–95. <https://doi.org/10.1109/MCSE.2007.55>
- Inglezakis, V.J., Zorpas, A., 2012. Fundamentals of Ion Exchange Fixed-Bed Operations, in: Dr., I., Luqman, M. (Eds.), *Ion Exchange Technology I: Theory and Materials*. Springer Netherlands, Dordrecht, pp. 121–161. [https://doi.org/10.1007/978-94-007-1700-8\\_4](https://doi.org/10.1007/978-94-007-1700-8_4)

- Jegatheesan, V., Shu, L., Jegatheesan, L., 2021. Producing fit-for-purpose water and recovering resources from various sources: An overview. *Environ. Qual. Manag.* 31, 9–28. <https://doi.org/10.1002/tqem.21780>
- Kabdaşlı, I., Tünay, O., 2018. Nutrient recovery by struvite precipitation, ion exchange and adsorption from source-separated human urine – a review. *Environ. Technol. Rev.*
- Kammerer, J., Carle, R., Kammerer, D.R., 2011. Adsorption and Ion Exchange: Basic Principles and Their Application in Food Processing. *J. Agric. Food Chem.* 59, 22–42. <https://doi.org/10.1021/jf1032203>
- Langmuir, I., 1918. THE ADSORPTION OF GASES ON PLANE SURFACES OF GLASS, MICA AND PLATINUM. *J. Am. Chem. Soc.* 40, 1361–1403. <https://doi.org/10.1021/ja02242a004>
- LeVan, M.D., Carta, G., 2008. Adsorption and ion exchange. McGraw-Hill, New York.
- Levenberg, K., 1944. A method for the solution of certain non-linear problems in least squares. *Q. Appl. Math.* 2, 164–168.
- Lima, E.C., Sher, F., Guleria, A., Saeb, M.R., Anastopoulos, I., Tran, H.N., Hosseini-Bandegharaei, A., 2021. Is one performing the treatment data of adsorption kinetics correctly? *J. Environ. Chem. Eng.* 9, 104813. <https://doi.org/10.1016/j.jece.2020.104813>
- Ma, A., Abushaikha, A., Allen, S.J., McKay, G., 2019. Ion exchange homogeneous surface diffusion modelling by binary site resin for the removal of nickel ions from wastewater in fixed beds. *Chem. Eng. J.* 358, 1–10. <https://doi.org/10.1016/j.cej.2018.09.135>
- Marquardt, D.W., 1963. An Algorithm for Least-Squares Estimation of Nonlinear Parameters. *J. Soc. Ind. Appl. Math.* 11, 431–441. <https://doi.org/10.1137/0111030>
- Marsili-Libelli, S., Guerrizio, S., Checchi, N., 2003. Confidence regions of estimated parameters for ecological systems. *Ecol. Model.* 165, 127–146. [https://doi.org/10.1016/S0304-3800\(03\)00068-1](https://doi.org/10.1016/S0304-3800(03)00068-1)
- McKinney, W., 2010. Data Structures for Statistical Computing in Python - SciPy Proceedings [WWW Document]. URL <https://proceedings.scipy.org/articles/Majora-92bf1922-00a> (accessed 9.26.24).
- Mudhoo, A., Pittman, C.U., 2023. Adsorption data modeling and analysis under scrutiny: A clarion call to redress recently found troubling flaws. *Chem. Eng. Res. Des.* 192, 371–388. <https://doi.org/10.1016/j.cherd.2023.02.033>
- Nelder, J.A., Mead, R., 1965. A Simplex Method for Function Minimization. *Comput. J.* 7, 308–313. <https://doi.org/10.1093/comjnl/7.4.308>
- O’Neal, J.A., Boyer, T.H., 2015. Phosphorus recovery from urine and anaerobic digester filtrate: comparison of adsorption–precipitation with direct precipitation. *Environ. Sci. Water Res. Technol.* 1, 481–492. <https://doi.org/10.1039/C5EW00009B>
- O’Neal, J.A., Boyer, T.H., 2013. Phosphate recovery using hybrid anion exchange: Applications to source-separated urine and combined wastewater streams. *Water Res.* 47, 5003–5017. <https://doi.org/10.1016/j.watres.2013.05.037>

- Rieger, L., Gillot, S., Langergraber, G., Ohtsuki, T., Shaw, A., Takacs, I., Winkler, S., 2012. Guidelines for using activated sludge models, First published. ed, Scientific and technical report series. IWA publishing, London New York.
- Saltelli, A., Aleksankina, K., Becker, W., Fennell, P., Ferretti, F., Holst, N., Li, S., Wu, Q., 2019. Why so many published sensitivity analyses are false: A systematic review of sensitivity analysis practices. *Environ. Model. Softw.* 114, 29–39. <https://doi.org/10.1016/j.envsoft.2019.01.012>
- Saltelli, A., Ratto, M., Andres, T., Campolongo, F., Cariboni, J., Gatelli, D., Saisana, M., Tarantola, S., 2007. *Global Sensitivity Analysis. The Primer*, 1st ed. Wiley. <https://doi.org/10.1002/9780470725184>
- Sengupta, S., Pandit, A., 2011. Selective removal of phosphorus from wastewater combined with its recovery as a solid-phase fertilizer. *Water Res.* 45, 3318–3330. <https://doi.org/10.1016/j.watres.2011.03.044>
- Smith, S.J., Wahman, D.G., Kleiner, E.J., Abulikemu, G., Stebel, E.K., Gray, B.N., Datsov, B., Crone, B.C., Taylor, R.D., Womack, E., Gastaldo, C.X., Sorial, G., Lytle, D., Pressman, J.G., Hauptert, L.M., 2023. Anion Exchange Resin and Inorganic Anion Parameter Determination for Model Validation and Evaluation of Unintended Consequences during PFAS Treatment. *ACS EST Water* 3, 576–587. <https://doi.org/10.1021/acsestwater.2c00572>
- Sung, C.-L., Tuo, R., 2024. A review on computer model calibration. *WIREs Comput. Stat.* 16, e1645. <https://doi.org/10.1002/wics.1645>
- Taghvaie Nakhjiri, A., Sanaeepur, H., Ebadi Amooghin, A., Shirazi, M.M.A., 2022. Recovery of precious metals from industrial wastewater towards resource recovery and environmental sustainability: A critical review. *Desalination* 527, 115510. <https://doi.org/10.1016/j.desal.2021.115510>
- The pandas development team, 2020. *pandas-dev/pandas: Pandas*. <https://doi.org/10.5281/zenodo.3509134>
- Tran, H.N., You, S.-J., Hosseini-Bandegharai, A., Chao, H.-P., 2017. Mistakes and inconsistencies regarding adsorption of contaminants from aqueous solutions: A critical review. *Water Res.* 120, 88–116. <https://doi.org/10.1016/j.watres.2017.04.014>
- Van Rossum, G., Drake, F.L., 2009. *Python 3 Reference Manual*. CreateSpace, Scotts Valley, CA.
- Vanrolleghem, P.A., Insel, G., Petersen, B., Sin, G., De Pauw, D., Nopens, I., Dovermann, H., Weijers, S., Gernaey, K., 2003. A comprehensive model calibration procedure for activated sludge models, in: *Weftec 2003*. Water Environment Federation, pp. 210–237.
- Virtanen, P., Gommers, R., Oliphant, T.E., Haberland, M., Reddy, T., Cournapeau, D., Burovski, E., Peterson, P., Weckesser, W., Bright, J., Van Der Walt, S.J., Brett, M., Wilson, J., Millman, K.J., Mayorov, N., Nelson, A.R.J., Jones, E., Kern, R., Larson, E., Carey, C.J., Polat, İ., Feng, Y., Moore, E.W., VanderPlas, J., Laxalde, D., Perktold, J., Cimrman, R., Henriksen, I., Quintero, E.A., Harris, C.R., Archibald, A.M., Ribeiro, A.H., Pedregosa, F., Van Mulbregt, P., SciPy 1.0 Contributors, Vijaykumar, A., Bardelli, A.P., Rothberg, A., Hilboll, A., Kloeckner, A., Scopatz, A., Lee, A., Rokem, A., Woods, C.N., Fulton, C., Masson, C., Häggström, C., Fitzgerald, C., Nicholson, D.A., Hagen, D.R., Pasechnik, D.V., Olivetti, E., Martin, E., Wieser, E., Silva, F., Lenders, F., Wilhelm, F., Young, G., Price, G.A., Ingold, G.-L., Allen, G.E., Lee, G.R., Audren, H., Probst, I., Dietrich, J.P., Silterra,

- J., Webber, J.T., Slavič, J., Nothman, J., Buchner, J., Kulick, J., Schönberger, J.L., De Miranda Cardoso, J.V., Reimer, J., Harrington, J., Rodríguez, J.L.C., Nunez-Iglesias, J., Kuczynski, J., Tritz, K., Thoma, M., Newville, M., Kümmerer, M., Bolingbroke, M., Tartre, M., Pak, M., Smith, N.J., Nowaczyk, N., Shebanov, N., Pavlyk, O., Brodtkorb, P.A., Lee, P., McGibbon, R.T., Feldbauer, R., Lewis, S., Tygier, S., Sievert, S., Vigna, S., Peterson, S., More, S., Pudlik, T., Oshima, T., Pingel, T.J., Robitaille, T.P., Spura, T., Jones, T.R., Cera, T., Leslie, T., Zito, T., Krauss, T., Upadhyay, U., Halchenko, Y.O., Vázquez-Baeza, Y., 2020. SciPy 1.0: fundamental algorithms for scientific computing in Python. *Nat. Methods* 17, 261–272. <https://doi.org/10.1038/s41592-019-0686-2>
- Vugrin, K.W., Swiler, L.P., Roberts, R.M., Stucky-Mack, N.J., Sullivan, S.P., 2007. Confidence region estimation techniques for nonlinear regression in groundwater flow: Three case studies. *Water Resour. Res.* 43. <https://doi.org/10.1029/2005WR004804>
- Wang, J., Guo, X., 2023. Adsorption kinetics and isotherm models of heavy metals by various adsorbents: An overview. *Crit. Rev. Environ. Sci. Technol.* 53, 1837–1865. <https://doi.org/10.1080/10643389.2023.2221157>
- Wang, J., Guo, X., 2022. Rethinking of the intraparticle diffusion adsorption kinetics model: Interpretation, solving methods and applications. *Chemosphere* 309, 136732. <https://doi.org/10.1016/j.chemosphere.2022.136732>
- Wang, W., Li, M., Zeng, Q., 2015. Adsorption of chromium (VI) by strong alkaline anion exchange fiber in a fixed-bed column: Experiments and models fitting and evaluating. *Sep. Purif. Technol.* 149, 16–23. <https://doi.org/10.1016/j.seppur.2015.05.022>
- Wang, Y., Wang, C., Huang, X., Zhang, Q., Wang, T., Guo, X., 2024. Guideline for modeling solid-liquid adsorption: Kinetics, isotherm, fixed bed, and thermodynamics. *Chemosphere* 349, 140736. <https://doi.org/10.1016/j.chemosphere.2023.140736>
- Xiao, Y., Azaiez, J., Hill, J.M., 2018. Erroneous Application of Pseudo-Second-Order Adsorption Kinetics Model: Ignored Assumptions and Spurious Correlations. *Ind. Eng. Chem. Res.* 57, 2705–2709. <https://doi.org/10.1021/acs.iecr.7b04724>
- Yoshida, H., Kataoka, T., Ikeda, S., 1985. Intraparticle mass transfer in bidispersed porous ion exchanger part I: Isotopic ion exchange. *Can. J. Chem. Eng.* 63, 422–429. <https://doi.org/10.1002/cjce.5450630310>
- Zhang, J., Amini, A., O’Neal, J.A., Boyer, T.H., Zhang, Q., 2015. Development and validation of a novel modeling framework integrating ion exchange and resin regeneration for water treatment. *Water Res.* 84, 255–265. <https://doi.org/10.1016/j.watres.2015.07.027>