

The dissolved oxygen ramp is immoral: facing global water challenges with mathematical analysis

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Abstract

The water sector is increasingly relying on data-driven approaches to create proxy measurements. These approaches are often trained with data covering only few situations, resulting in a lack of robustness and increasing the risk of false predictions. Therefore, robust approaches are needed for data-driven proxy measurements (i.e. soft sensors), especially for water recovery and reuse. In climate science and robotics, [Dynamical Systems Analysis \(DSA\)](#) is used to explore a wide range of system behaviour and uncover conditions of high uncertainty, and potential tipping points. [DSA](#) allows the systematic analysis of model dynamics, and the measurable features caused by these dynamics. We created a novel [DSA](#) workflow for soft-sensor development to measure water quality. Herein, we demonstrate that the integration of [DSA](#) into soft-sensor development adds robustness by revealing all mathematically possible combinations of state variables that lead to a feature. It can thus detect possible interferences and help design the soft sensor to avoid these. We used [DSA](#) for the falsification of a ramp-feature-based soft sensor and found that, despite a published, successful laboratory and real-world application, the ramp feature is immoral. Such an immorality implies that false predictions could occur. However, the [DSA](#) analysis uncovered under which operational conditions the ramp becomes a robust soft sensor feature. Targeted experiments will be necessary to confirm the boundary of the robust conditions in the real world.

Contents

1	Introduction	2
2	Results and Discussion	5
2.1	Model dependency graph	5
2.2	The dissolved oxygen ramp feature in detail	5
2.3	Identification of states leading to dissolved oxygen ramps	6
2.4	Immorality of the dissolved oxygen ramp	8
2.5	Ramp points distribution	8
2.6	Limitations and outlook	9
3	Conclusion	10

4	Materials and methods	10
4.1	Dissolved oxygen feature	10
4.2	Model signal features	11
4.3	Model structure	11
4.4	Ramp contingency	12
A	Supplementary information	17
A.1	Time derivatives	17
A.1.1	Single state actuation	18
B	Ramp points curve	18
B.1	Interpretability of the inflection point curve	20
B.2	Symbols name	20
B.3	ASM1 model	20
C	Causal diagram	25
C.1	Ammonium depletion soft-sensor based on the dissolved oxygen ramp feature	26

Glossary

ADM Anaerobic Digestion Model. 3, 4, 10, 12

ASM Activated Sludge Model. 3–6, 8–12, 18, 25

BSM Benchmark Simulation Model. 4, 25

DSA Dynamical Systems Analysis. 1, 3–11, 26

ODE Ordinary Differential Equation. 3

ramp A time-based signal feature. It is defined as the signal’s non-saddle (non-zero first time derivative) inflection point (zero second time derivative).. 2, 5–12, 18–21, 25, 26

SBR Sequencing Batch Reactor. 5, 10

1 Introduction

The impact of climate change on water cycles has a cascading effect on urban areas, agriculture, industry, and the natural environment, necessitating adaptation. A flexible, circular on-site infrastructure [1–3], is a promising adaptation strategy. Furthermore, on-site plants may allow cost savings by removing the need for sewer infrastructure [4].

Monitoring and control pose decisive challenges inherent to on-site water and wastewater infrastructure. This can be attributed, on the one hand, to the high capital cost required to purchase physical water quality sensors. One way to reduce these costs is developing soft sensors [5] based on inexpensive proxy sensors instead. On the other hand, monitoring is hampered by required sensor maintenance [6]. Most studies do not address the maintenance challenge even for on-site settings [e.g. 7, 8]. However, sensors expose a variety of often non-linear wearing effects such as damage, fouling, or deterioration [e.g. 9–12] and sensor faults that impact the treatment performance [13]. The wearing effects are especially challenging for data-driven approaches, despite accurate results [e.g. 14]. The reason is that wearing effects cause measurements that can lie outside the training dataset. Consequently, sensors should either be installed in an environment where nearly no maintenance is required, for example, a chlorination tank [15] or features for monitoring [16] and control [17] need to be robust to sensor wearing effects such as drift or noise by design.

Schneider et al. [16] directly compared a soft sensor to predict complete ammonia oxidation using a ramp feature (i.e. a non-saddle inflection point, see "Ramp" in fig. 7) in the dissolved oxygen signal based on data from maintained and unmaintained sensors. The ammonium-depletion detection based on the **ramp** feature are robust by design to most effects from noise, measurement uncertainties, or effects of sensor wearing by relying on the qualitative trend instead of absolute values and including a low-pass filter. Nevertheless, three time-consuming sampling campaigns revealed at least two alternative causes for the dissolved-oxygen **ramp** feature i.e. the aeration pattern and alkalinity limitation. Additionally, in silico experiments based on synthetic data generated with SUMO [18] revealed that short solids retention time [6] can cause a false predictions (see fig. 1. Hence, designing robust soft sensors is constrained by our ability to systematically explore the causes of the feature, e.g. due to the plethora of state combinations that can lead to it. Moreover, the presence of alternative causes for the feature (see the supplementary information sec. C) poses a risk of erroneous soft-sensor predictions, potentially affecting human and

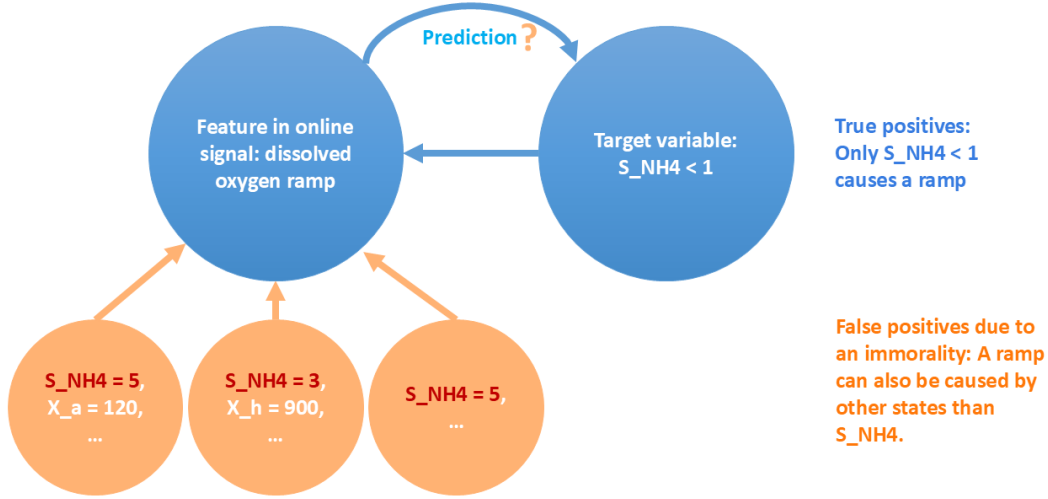


Figure 1: Schematic explanation of immortality, true and false positive predictions. S_{NH_4} represents the target variable in this case, the ammonium concentration. X_a (the autotrophic bacteria) and X_h (the heterotrophic bacteria) are states involved in an immortality and could lead to false predictions. This can occur with any other state variable, excluding the target variable, that is used to predict the feature.

environmental health. To avoid these undesired effects, a systematic identification of the alternative causes and the analysis of the plausibility of their occurrence is required.

For complex systems such as the human brain, global weather, or biochemical wastewater treatment processes, understanding the functional architecture is crucial for defining effective, rigorous, principled, and general methods, yet remains a challenge [19]. To find such a method, we systematically analyse sets of **Ordinary Differential Equations (ODE)** using an approach that we call **Dynamical Systems Analysis (DSA)** [see 20, for a joyful introduction]. The approach aims at understanding a set of **ODEs** without integrating them, usually leading to efficient workflows and adding a novel perspective to problem-solving. **DSA** encompasses techniques for characterising stability and bifurcations (cf. tipping points), and allows screening any dynamic model for a plethora of features. In other fields, such as robotics, **DSA** is frequently employed [21–23, e.g.] to gain mathematical insights into the engineering problem at hand, which more often than not leads to novel solutions. In climate science, **DSA** has been used to predict abrupt changes like tipping points [24]. **DSA** also provides an alternative perspective into model calibration [25, ch. 1], which focuses on validating structural features of the model with experimental data, as opposed to the more conventional approach of matching time series. Due to all these properties, we hypothesise that an improved understanding of model dynamics with **DSA** is crucial for engineering features for robust monitoring and control.

In wastewater treatment, **DSA** remains an uncommon approach. When **DSA** is employed, it is frequently referred to with ambiguous terms like "analysis", which hinders identifying relevant studies. We, therefore, believe that a review on **DSA** applied in biochemical treatment and recovery processes of wastewater and similar streams is timely, especially to the popular **Activated Sludge Model (ASM)** [26] and **Anaerobic Digestion Model (ADM)** [27]. Hence, we provide a brief one in tab. 1.

Reference	Model	DSA outcome	Implication/application
Vanrolleghem [28]	ASM1 and solid flux sedimentation model	Implication of degradation and settling for control design	Improved control design
Grognard and Bernard [29]	Two-population model for anaerobic processes	Stability analysis with non-trivial solutions	Avoiding instability e.g. acidification
Shen et al. [30]	Anaerobic digestion	Identified three stable points, and a saddle-node bifurcation under typical operation conditions	Identification of stable operating conditions
Nelson and Sidhu [31]	ASM1	Identified two branch points	Determination of optimal residence time
Volcke et al. [32]	Two-step biological conversion system	Identification of steady state multiplicity	Experimental design
Benyahia et al. [33]	Two-step ADM	Bifurcation and stability analysis	Experimental design for a soluble microbial products monitoring strategy
Bornhöft et al. [34]	ADM1	Bifurcation analysis	Model selection: Impact of model simplification
Dionisi et al. [35]	Two-population model in a batch process	Stability analysis	Accelerate calculations to reach steady state
Wade and Wolkowicz [36]	Impulsive annamox	Bifurcation analysis	Prediction of wash-out of nitrite-oxidising bacteria
Neto et al. [37]	Benchmark Simulation Model1 (BSM)	Structural controllability and observability	Stability, controllability and observability
Sampaio et al. [38]	ASM1 extended for greenhouse-gas emissions, two-step nitrification, four-step denitrification, and 10-layer settling	Structural analysis	Full-state controllability and observability of greenhouse gas emissions

Table 1: Literature review on DSA in anaerobic digestion and activated sludge modelling for wastewater treatment.

These successful examples from the water sector indicate the value of **DSA** for analysing steady states and stability. These analyses contribute to the identification of regions of optimal operation, control, or observability, and experimental design. However, the use in other fields shows, that the potential of **DSA** is far from exhausted in the water sector. We deduce that systematic detection of information-rich system dynamics (i.e. features) merits further attention, with applications in feature-based model calibration and feature engineering for soft-sensor development, the latter being the primary focus of this study.

2 Results and Discussion

Herein, we showcase **DSA** for qualitative trend feature-based soft-sensor development. To model the activated sludge process we used the mass balance-corrected version of **ASM1** [39] by Hauduc et al. [40]. We exemplify the use of **DSA** for feature engineering by systematically investigating factors affecting the **ramp** feature in the dissolved oxygen signal in a **Sequencing Batch Reactor (SBR)** to predict the effluent ammonium concentration. The current work complements previous data-driven, mechanistic, or hybrid feature-learning approaches [41] on a process familiar to the authors to introduce and discuss **DSA** as a potential standard procedure for soft-sensor development.

2.1 Model dependency graph

A worthwhile first step to get an overview before performing a **DSA** is a visual inspection of the dependencies. The dependency graph implies causation from the relations between states defined in the analysed model. Figure 2 shows the dependency graph of the 14 states of the mass balance-corrected **ASM1** Hauduc et al. [40]. The many circular dependencies reflect the high complexity of the system. Furthermore, all processes follow the pattern state→rate→state, which hints at the specific model structure presented in sec. 2.3. By looking at the arrows and their direction, we can identify that five of these states do not affect the dynamics of the other states, of which two are uncoupled (no children or parents in the graph, i.e. $X_{nb,in}$, S_{nb}), and three are measurements (no children, i.e. $X_{nb,e}$, S_{N2} , S_{alk}). Therefore, the dynamics of **ASM1** are determined by the following nine states (in brackets is the notation by Henze et al. [39]):

- S_b Soluble biodegradable organics (SS)
- X_{cb} Particulate and colloidal biodegradable organics (XS)
- X_h Ordinary heterotrophic organisms (XBH)
- X_a Autotrophic nitrifying organisms (NH4+ to NO3-, XBA)
- S_{O2} Dissolved oxygen (SO)
- S_{NOx} Nitrate and nitrite (NO3 + NO2) (considered to be NO3 only for stoichiometry, SNO)
- S_{NHx} Ammonia (NH4 + NH3) (SNH)
- S_{bN} Soluble biodegradable organic N (SND)
- X_{cbN} Particulate and colloidal biodegradable organic N (XND)

2.2 The dissolved oxygen ramp feature in detail

To illustrate the traditional way of finding inflection points (i.e. ramp features), we show in fig. 3 the time-series of the states of **ASM1** generated by one set of initial conditions. With S_{O2} and S_{NHx} in the top panel true and false positive (see fig. 1 for a definition) **ramp** features can be identified in this single batch run. The vertical lines represent time instances where the conditions of a ramp in the oxygen signal (defined in equations (9)-(10)) are satisfied.

The first **ramp** in the S_{O2} signal is particularly interesting. If the soft sensor does not identify this ramp correctly to stop the treatment process, the ammonium concentration starts rising. The relevant process is the ammonification during the decay of heterotrophic bacteria in the absence of sufficient autotrophic bacteria. The second **ramp** is a false positive one. The third **ramp** is what we generally would expect in a biochemical treatment process that recovers wastewater batch-wise. Remarkable is also that the two true positive **ramps** have a much steeper slope than the false positive one. The first objective of the present work is to use **DSA** to efficiently screen for potential causes of false positives in the **ramp** feature, in other words to identify when the soft sensor fails to estimate the ammonium concentration correctly. Another objective is to explore the potential of **DSA** to make the **ramp** feature more robust.

Formally, **ramps** are zeros of the second time derivative in which the first derivative is positive (i.e. non-saddle inflection point). The locus, i.e. the set of all points that satisfy the specified conditions, of **ramps** can be described mathematically by:

$$\text{ramps} := \{t : \dot{x}(t) > 0 \wedge \ddot{x} = 0\} \quad (1)$$

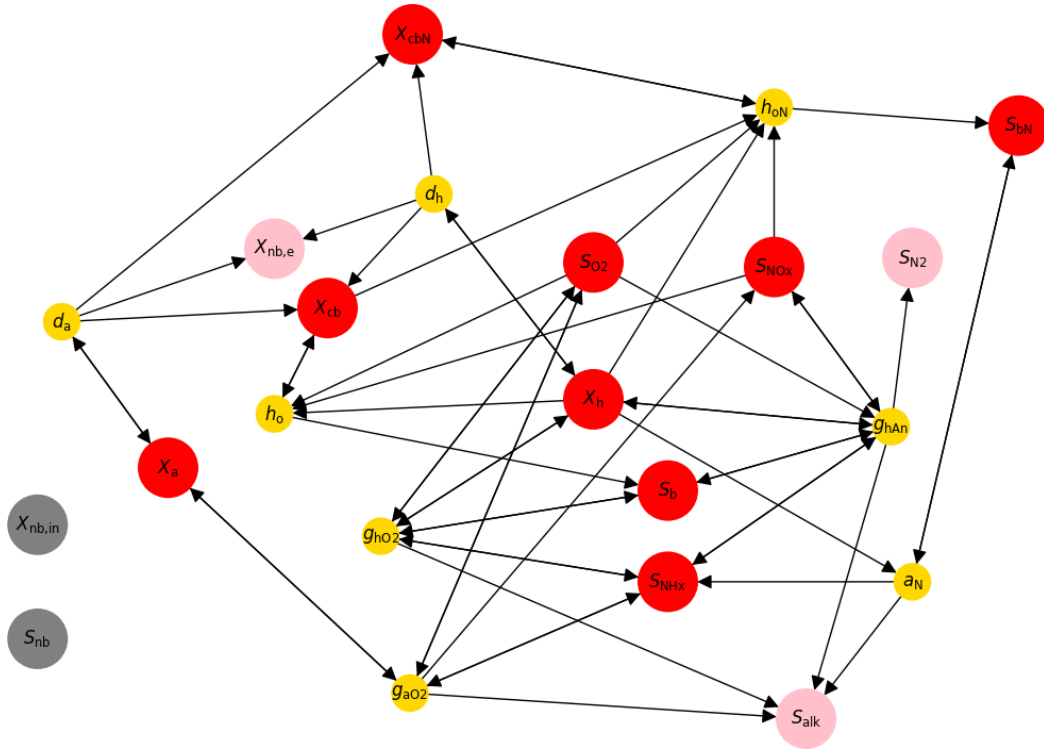


Figure 2: [ASM1](#) state dependency graph. The graph depicts the interactions between the states in the model. States (larger nodes) affect rates (smaller nodes), which in turn affect the states through their time derivative. The arrows indicate which states and rates affect each other. Pink nodes with no child are measurements which do not impact other states. Grey nodes with neither parent nor child are uncoupled (i.e. the non-biodegradable organics, S_{nb} , and particulate non-biodegradable organics from the influent, $X_{nb,in}$). The red nodes are the potential alternative causes for the [ramp](#) feature that would lead to a false prediction of the target variable.

where the signal is given by $x(t)$, the dotted symbols represent time derivatives, and the wedge (\wedge) is the logical conjunction (and) symbol, meaning that both criteria need to be fulfilled simultaneously.

2.3 Identification of states leading to dissolved oxygen ramps

To identify the states that lead to a [ramp](#) feature in the dissolved oxygen signal (represented by the state S_{O_2}), we need to compute the second time derivative of this state (i.e. \ddot{S}_{O_2}), see equation (13). This derivative can be computed directly from the model equations without having to simulate the full set of equations to generate a signal, making [DSA](#) a compelling and computationally efficient method. This computation can be done for any model structure, but for the [ASM](#) family the computations are simplified thanks to the structure of the model that can for example be identified from fig. 2:

$$\dot{\mathbf{x}} = \mathbf{M} \mathbf{r}(\mathbf{x}) + \mathbf{z} \quad (2)$$

$$\ddot{\mathbf{x}} = \mathbf{M} \dot{\mathbf{r}} + \dot{\mathbf{z}} = \mathbf{M} \mathbf{J}_r \dot{\mathbf{x}} + \dot{\mathbf{z}} \quad (3)$$

where we used boldface, e.g. \mathbf{x} , for vectors, and capital typewriter font, e.g. \mathbf{M} for matrices. In equation (2), \mathbf{x} is the vector of states of the model. The states are then transformed via the mapping $\mathbf{r}(\mathbf{x})$, into a new vector, which herein we will call *rates* vector. The matrix \mathbf{M} , such as the stoichiometric matrix Gujer and Henze [42], combines the rates to produce the time derivatives of the states. The additional term (\mathbf{z}) in equation (2) represents the *actuation* on the states of the system. More details and explanations are given in sec. 4.3. Therefore, if the autonomous parts of the model, \mathbf{M} and $\mathbf{r}(\mathbf{x})$, are given in tabular form, we can evaluate equations (2) and (3) directly, which allows to analyse any features based on the time derivatives of the model states.

Applying these formulae to [ASM1](#) (given in supplementary information B.3), we obtain equations for the first and second time derivative of the dissolved-oxygen state (S_{O_2}). For the first derivative, we get:

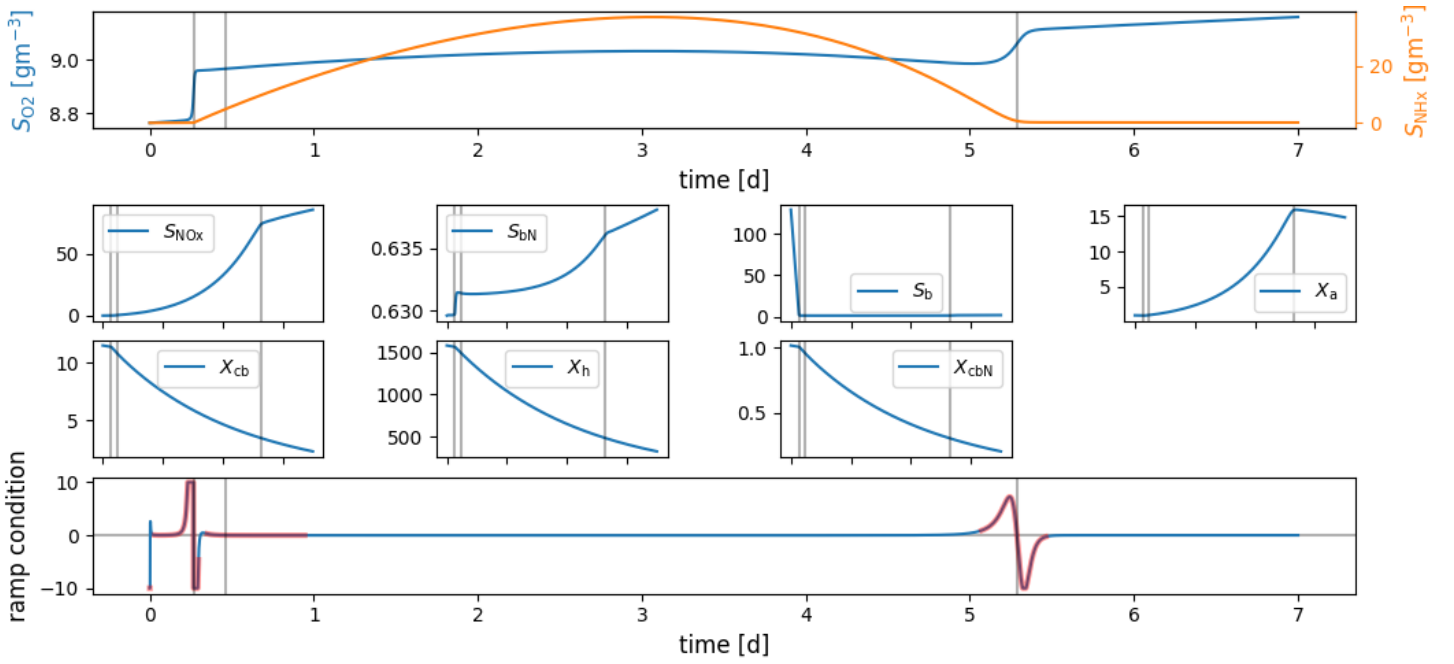


Figure 3: The top panel shows the dissolved oxygen (S_{O_2}) and the effluent ammonium (S_{NH_x}) concentration signals. The **ramp** time instances are marked on all panels as vertical lines and the time intervals on the x-axes are the same. The second **ramp** (close to 0.5 d) is a false positive for the effluent ammonium concentration estimation, the third one (close to 5.25 d) is a true positive feature, and the first one (close to 0.25 d) is a true positive that is not caused by the ammonium reaching a concentration below the threshold at the **ramp**-occurrence time, but the ammonium is already low before the **ramp**. Possible causes for the **ramp** feature can be identified by looking at the other panels, showing other state variables of the model (see list in page 5). Each of these three **ramp** observations corresponds with one point on an inflection point curve. The time series are generated for one single set of initial conditions, and are shown here to display the **ramp** feature detection. These time series would not be simulated when **DSA** is used to obtain the inflection point curves. The units of all state variables are $g\ m^{-3}$.

$$\dot{S}_{O_2} = m_{S_{O_2} g_{aO_2}} g_{aO_2}(S_{NH_x}, S_{O_2}, X_a) + m_{S_{O_2} g_{hO_2}} g_{hO_2}(S_{NH_x}, S_{O_2}, S_b, X_h) + z_{O_2} \quad (4)$$

$$m_{S_{O_2} g_{aO_2}} := \frac{\gamma_a + \iota_{COD,NO_3}}{\gamma_a} \quad m_{S_{O_2} g_{hO_2}} := \frac{\gamma_h - 1}{\gamma_h} \quad (5)$$

$$g_{aO_2}(S_{NH_x}, S_{O_2}, X_a) := \mu_{max,a} X_a \frac{S_{NH_x} S_{O_2}}{(S_{NH_x} + \kappa_{NH_x,a})(S_{O_2} + \kappa_{O_2,a})} \quad (6)$$

$$g_{hO_2}(S_{NH_x}, S_{O_2}, S_b, X_h) := \mu_{max,h} X_h \frac{S_{NH_x} S_{O_2} S_b}{(S_{NH_x} + \kappa_{NH_x,h})(S_{O_2} + \kappa_{O_2,h})(S_b + \kappa_b)} \quad (7)$$

Since the biological activity of heterotrophic and autotrophic bacteria consumes dissolved oxygen, the first two terms are always non-positive. The signs of $m_{S_{O_2} g_{aO_2}}$ and $m_{S_{O_2} g_{hO_2}}$ are determined by the coefficients in equation (5), and for plausible values of γ_a and γ_h (see tab. 4), these signs are negative.

In the equations above we explicitly show the dependency of the rates (states in parentheses) that we extracted from the model equations. We omit them in the formulae for the second derivative:

$$\begin{aligned} \ddot{S}_{O_2} = & \dot{S}_{NH_x} \partial_{S_{NH_x}} (g_{aO_2} m_{S_{O_2} g_{aO_2}} + g_{hO_2} m_{S_{O_2} g_{hO_2}}) \\ & + \dot{S}_{O_2} \partial_{S_{O_2}} (g_{aO_2} m_{S_{O_2} g_{aO_2}} + g_{hO_2} m_{S_{O_2} g_{hO_2}}) \\ & + \dot{S}_b \partial_{S_b} g_{hO_2} m_{S_{O_2} g_{hO_2}} + \dot{X}_a \partial_{X_a} g_{aO_2} m_{S_{O_2} g_{aO_2}} + \dot{X}_h \partial_{X_h} g_{hO_2} m_{S_{O_2} g_{hO_2}} \\ & + z_{O_2} \partial_{S_{O_2}} (g_{aO_2} m_{S_{O_2} g_{aO_2}} + g_{hO_2} m_{S_{O_2} g_{hO_2}}) + \dot{z}_{O_2} \end{aligned} \quad (8)$$

The conditions for the **ramp** (section 4.2) read:

$$\dot{S}_{O_2} > 0 \quad (9)$$

$$\ddot{S}_{O_2} = 0 \quad (10)$$

The first condition for a **ramp**, equation (9), requires a positive time derivative. This expresses that the rate at which oxygen is transferred to the liquid phase (z_{O_2}) needs to be bigger than the oxygen uptake rate by the bacteria, as there

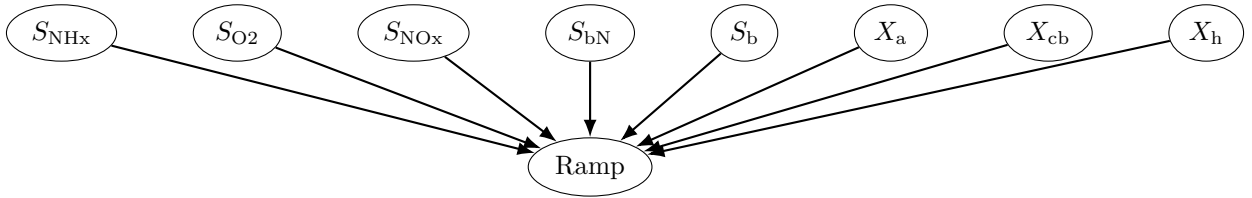


Figure 4: Causes of the dissolved oxygen (S_{O_2}) **ramp** feature. Obtained from the variables in equation (8). Several V-structures are identified from the causal relations. See list in page 5 for the variable names.

cannot be more oxygen consumed than is present. Therefore, in **ASM1** the dissolved oxygen **ramp** is a feature that can only be generated with a source of dissolved oxygen (aeration) as otherwise equation (9) cannot be fulfilled. This underlines the role of aeration for the detection and robustness of the **ramp** feature, which in a different context, was observed in a real-world case in [6].

Using **DSA** shows that the conditions (9) and (10) depend on other states besides S_{NH_x} and S_{O_2} , i.e. S_{NO_x} , S_{bN} , S_b , X_a , X_{cb} , X_h . So, even if we kept S_{NH_x} constant at a value above the prediction threshold for a full ammonium depletion, changes to these other states could generate a **ramp** that is not due to a S_{NH_x} below this threshold. This can potentially lead to false positive predictions with a **ramp**-based soft sensor.

2.4 Immorality of the dissolved oxygen ramp

From the nine states in the dependency graph (fig. 2), **DSA** shows that eight states are actual causes of a **ramp**, X_{cbN} is not a cause. These eight states form several V-structures within the causal diagram of the **ramp** feature in fig. 4; known as *immoralities* [43]. The states (parents) in a V-structure, will be associated (correlated) when data is selected conditional on the presence of the feature (child). These correlations can be exploited by a data-driven machine learning model to infer difficult-to-monitor states, e.g. S_{NH_x} from S_{O_2} , or to alleviate an effluent ammonium prediction task. However, since pure data-driven approaches are limited to exploiting correlations without guaranteeing causality; adding causal relations, as we do here, can enhance the performance and information extracted from the method, e.g. to include root cause analysis and therewith improve the robustness of the sensor.

DSA allows the systematic evaluation of conditions that lead to a **ramp** in a search process by sampling the states independently to determine the causes (parents) of the ramp feature, see fig. 4. This is illustrated in fig. 5a which shows all ramps found on the (S_{O_2} , S_{NH_x}) plane for 10,000 uniformly distributed random values of all the other six states that can cause **ramps** (see section B for explanations on how to read these curves). Each line is a 0-level set curve of equation (10) that satisfies equation (9). Figure 5a further shows that many of these **ramps** are below an arbitrary threshold of $1 \text{ g m}^{-3} S_{NH_x-N}$. All the ramps below the threshold would lead to the correct conclusion that the ammonium oxidation process is complete (true positive), hence the feature is informative. However, many other **ramps** are clearly above the threshold, and would lead to an erroneous conclusion (false positive).

To further investigate these false positives, fig. 5b shows all **ramps** found on the (S_{O_2} , X_a) plane for the autotrophic bacteria. We chose the autotrophic bacteria because they are responsible for the removal of nitrogen compounds such as ammonium. Therefore, we have only sampled values of S_{NH_x} above the arbitrary threshold, between 1 g m^{-3} to 200 g m^{-3} . For all other states, we used the same ranges as before. Hence, only false positive **ramps** are shown in this plot. This allows us to evaluate the causes of false positive results and identify measures to increase accuracy. The pattern clearly shows that a high concentration of X_a could drastically reduce the false positive results.

In summary, fig. 5a and fig. 5b illustrate how **DSA** can be used to systematically scan a feature space for the occurrence of features and their causes. Analysing the equations allows us to identify causal relations, which is very different from checking data for correlation. For an extended discussion of these results concerning experimental work in [6, 16] see sec. C.1.

2.5 Ramp points distribution

Figure 6 shows the distribution of states that cause a **ramp** feature (satisfy equations (9)- (10)). Blue and orange dots indicate effluent ammonium concentrations below and above an arbitrary threshold (T), respectively. Blue dots are thus true positives for the ammonium soft sensor and orange dots represent false positives (compare fig. 1). The plot also shows the dissolved oxygen time derivative (\dot{S}_{O_2}) at these points. We observed in fig. 3 that true **ramps** are associated with higher derivatives than false **ramps**. The top right panel of fig. 6 (S_{NH_x} vs \dot{S}_{O_2}) supports this observation, as almost no false **ramps** are found above dissolved oxygen time derivatives of $400 \text{ g m}^{-3} \text{ d}^{-1}$. Therefore, using time derivative information (slope in a time series plot) together with the detection of **ramps** constitutes a more accurate feature.

This observation also provides a plausible explanation for the improved accuracy of an ammonium depletion classifier when slope information was included. Without a restriction on the slope the feature was not informative [6, 16]. In these previous studies this slope restriction was found with trial and error, while **DSA** offers a systematic approach. **ASM1** already captures this relation, despite missing processes and its relative simplicity compared to other activated sludge models. Identifying such

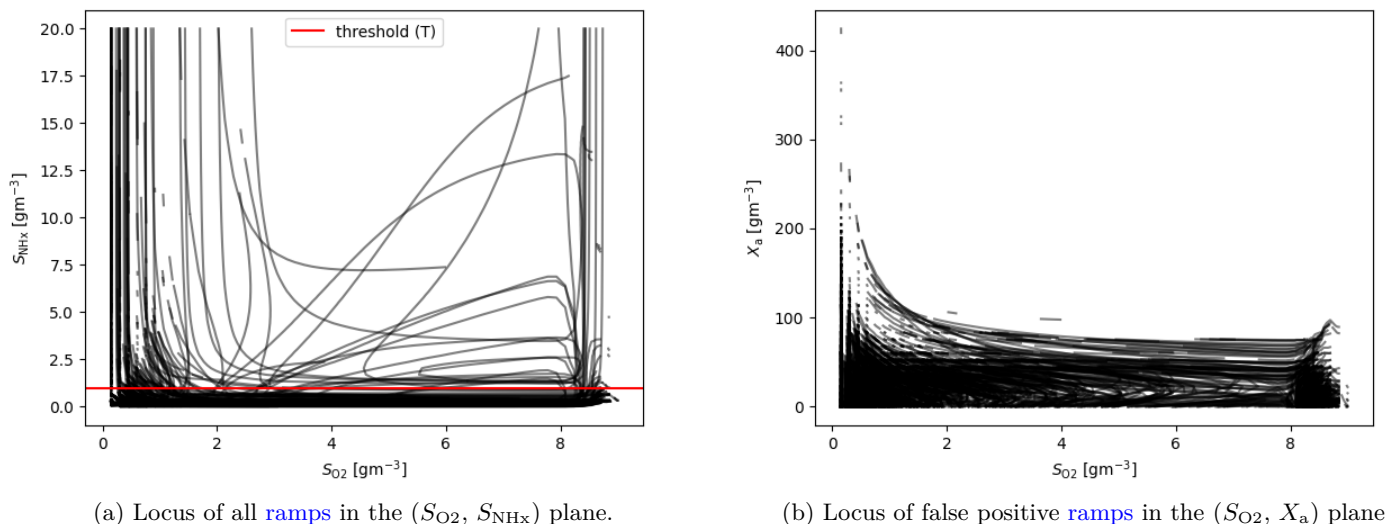


Figure 5: Inflection point curves in two planes. The lines show the combinations of S_{O_2} and S_{NHx} or S_{O_2} and X_a that lead to a ramp feature. 10,000 simulations with different initial conditions for each panel were conducted. The darker the area, the more overlapping inflection points are in the region. The ammonium concentrations for b) were restricted to only show panels with false positive ramps i.e. the ammonium concentration is above an arbitrary threshold of 1 g m^{-3} .

a slope dependence shows that for the ramp-feature design **ASM1** is structurally useful. Therefore, we demonstrate that and how **DSA** can be used for structural model verification.

In the top right panel of fig. 6 we further observe that eliminating the false positives by defining a minimal slope and rejecting all ramps below the chosen slope (e.g. $< 400 \text{ g m}^{-3} \text{ d}^{-1}$) may result in many false negatives. The coloured shaded regions for different levels of X_a indicate that with a higher X_a the slope tolerance could be considerably lowered to reduce the false negatives. Hence, finding another feature or sensor to estimate X_a would increase the accuracy of the ramp-based soft sensor.

Further patterns can be recognised in fig. 6- For example if S_{bN} is above approximately 7 g m^{-3} , no correct predictions may be possible. It is reasonable to think that if the soluble biodegradable organic nitrogen is high, too much ammonium is formed, and its concentration drop is limited. The experimental confirmation of this discovery is part of future work. Furthermore, observing multiple additional patterns in fig. 6 to the ones discussed in this article shows great potential for future studies, e.g., to further improve the soft sensor with an estimate X_a or to identify an early warning feature for loss of nitrifying biomass. Additionally, the results of the **DSA** hint at which regions could be used by data-driven models.

Herein, we demonstrated how **DSA** can be used to improve the feature, e.g. by adding further constraints such as the slope of S_{O_2} . **DSA** could also be used to discover feature sets from signals of other sensors than dissolved oxygen. These feature sets could further improve the accuracy and robustness of an ammonium-depletion soft sensor or soft sensors for different quantities of interest.

We provide all the code under a GNU free public license in Carbajal and Schneider [44], hence readers can adapt it to their own analyses and for different models, as long as these models have the structure described in equation (3).

2.6 Limitations and outlook

As illustrated herein, **DSA** is applied to a specific model, and is restricted by the model's structure. For example, there is another known state variable that could cause a ramp: the alkalinity [45], which does not influence other states in the definition of **ASM1**. Hence, no **DSA** performed on **ASM1** will inform us of the influence of alkalinity. However, the presented method for **DSA** can be applied to other models such as **ASM3** to assess the relationship between alkalinity and the ramp feature. In the present work, it was decided to develop a proof of concept of the method based on **ASM1** as this is still the most commonly used model in the **ASM** family.

DSA deals with deterministic dynamics, therefore process noise cannot be handled easily with the current approach. Measurement noise does not affect the dynamics, and therefore correlations between state variables and features based on them are not affected by measurement noise. Detection of these features in real systems, by means of sensors and state estimation, is however affected by measurement noise. Examining the impact of measurement noise and the feasibility of applying **DSA** results in practical applications is not the focus of this work, nor is it an inherent part of **DSA** analysis; these are additional analyses conducted subsequently.

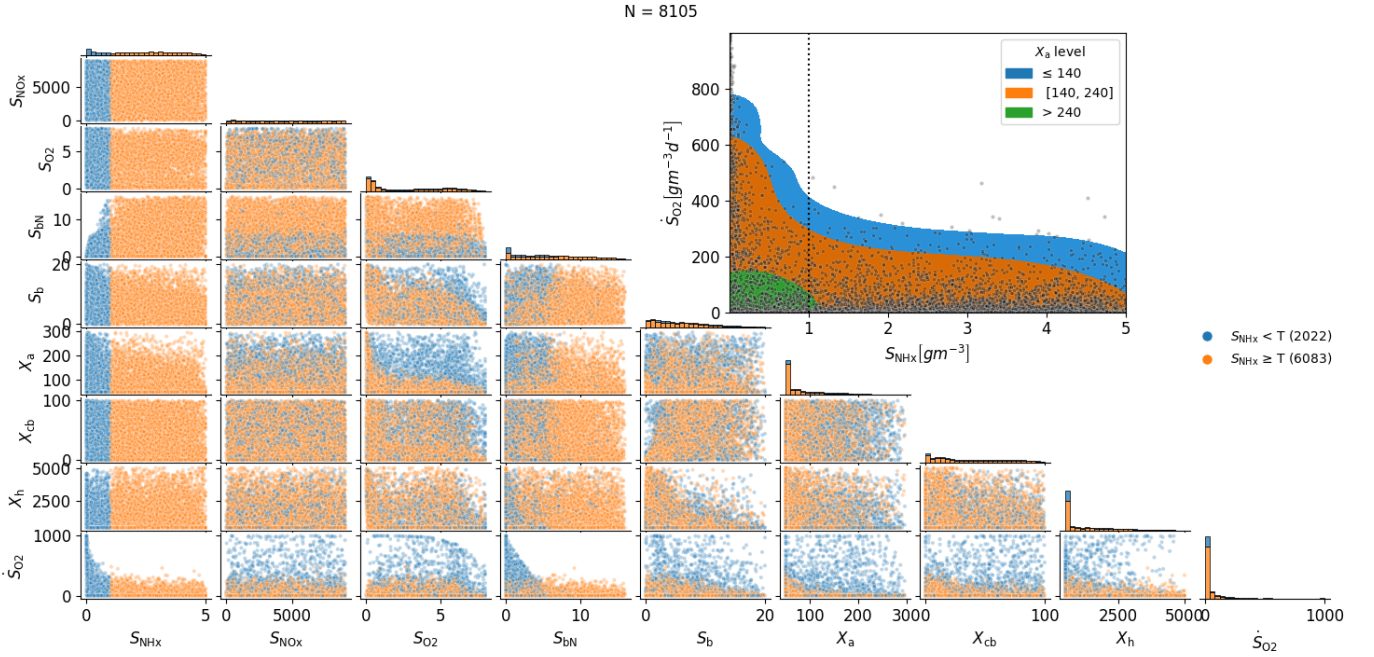


Figure 6: **ASM1** ramp points distribution. Distribution of states fulfilling the ramp conditions (9) and (10). The points are colored by whether the value of S_{NH_x} is above (orange) or below (blue) an arbitrary threshold (T). The number of samples (N) in each category is shown in the legend. The panels on the diagonal always show the density of points. The bottom row of the panels shows the value of the dissolved oxygen time derivative (ramp's slope). The top-right panel shows the relation between the ramp's slope and the ammonium concentration. The coloured regions on that panel show the level of autotrophic bacteria (X_a), illustrating a further correlation that can be exploited for increasing the accuracy of a full-ammonium-depletion detection soft sensor.

3 Conclusion

1. We used **DSA** to systematically evaluate all the mathematically possible solutions leading to a **ramp** feature in the dissolved oxygen signal in **ASM1**. The dissolved oxygen ramp is clearly immoral, meaning that not only a low value of the state S_{NH_x} , i.e. an ammonium depletion, can cause the **ramp** feature, but also eight alternative causes, i.e. S_{O_2} , S_{NO_x} , S_{bN} , S_{b} , X_a , X_{cb} , X_{h} , X_{cbN} .
2. With **DSA** we identified that the slope of the dissolved oxygen at the locus of the **ramp** is relevant and can increase the accuracy of the **ramp** feature.
3. Many models (e.g. **ASM** family or **ADM1**) follow the same matrix structure which means that the formulae presented here can be applied "as-is" for a plethora of models in the water and resource recovery domain.
4. The analysis underlines how important systematic falsification studies are, especially when methods are developed for setups that can directly impact human and environmental health such as water recovery facilities for reuse purposes. With **DSA**, we suggest a concrete method for such a systematic analysis by increasing the robustness of soft sensors through the exposure of immoralities that can then be mitigated by multi-feature approaches.

4 Materials and methods

4.1 Dissolved oxygen feature

As introduced in section 1, a feature can be used to monitor a biochemical wastewater treatment process. From the two measurement signals pH and dissolved oxygen from a previous study [16], we here chose the latter, as S_{O_2} is a state in the **ASM1** while pH is not a state variable in any of the **ASM** models. This allowed us to start with a model with a small number (eight) of equations to implement. The feature of the dissolved oxygen is a **ramp** (see fig. 7) that can be used to predict the effluent ammonium (S_{NH_x}) concentration. Olsson and Andrews [46] describe the ramp feature for a plug flow reactor, hence w.r.t. the location. Following the same principle in our study, we use an **SBR**, thus the ramp is w.r.t. time, i.e. whenever an inflection point is observed during the aeration phase, we assume that the ammonium is fully oxidised so the ammonium nitrogen is below or equal to an arbitrary threshold of 1 g m^{-3} [6]. If there is no ramp feature the effluent

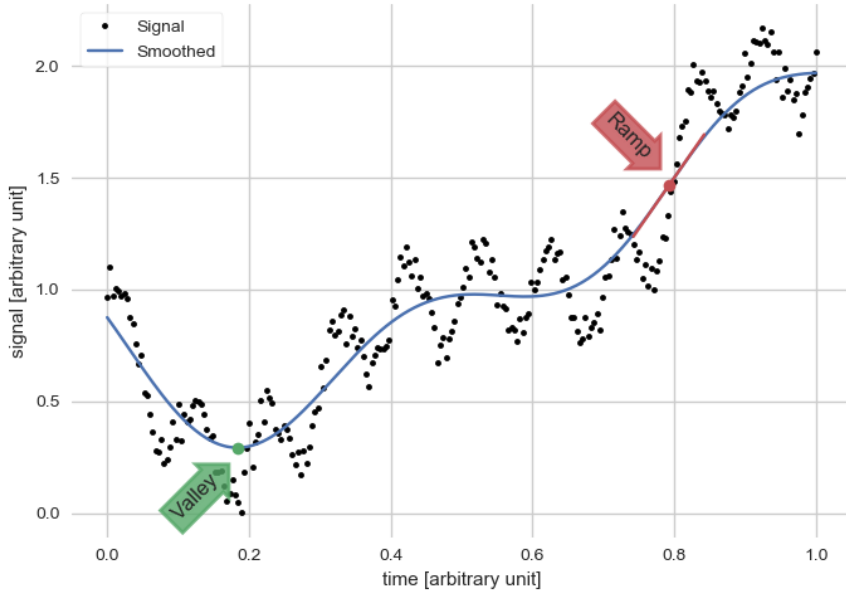


Figure 7: Illustration of two different features based on time derivatives of a signal. An extrema (valley) and a non-saddle inflection point (ramp). The latter we use in this article. An example for the arbitrary unit on the x-axis would be day on the y-axis it could be g m^{-3} . The figure is created with Carbajal and Schneider [47].

ammonium concentration is estimated to be above 1 g m^{-3} . For this prediction to be accurate and robust there should be no other state than an ammonium concentration below or equal to 1 g m^{-3} leading to the **ramp** feature. First, we obtain all the mathematically possible solutions for a **ramp** feature in **ASM1** with **DSA**. See 4.2 for a detailed description of how we obtained these solutions. Next, we further constrain these mathematically possible solutions to the physically plausible ones by restricting them to only positive states. Restrictions were that concentrations cannot be negative and the setting of a wide, yet feasible range for all state variables. See tabs. 5-7 in the supplementary information for ranges reported in the literature, which we consulted to restrict the phase space of the state variables.

4.2 Model signal features

We are concerned with computing signal features based on the signal’s derivatives w.r.t. time (time derivatives). Figure 7 shows two examples of such features: a valley, which is a minimum extreme point and a **ramp**. Extrema are zeros of the signal’s first time derivative, and **ramps** are zeros of the second time derivative in which the first derivative is positive (i.e. non-saddle inflection point). The locus¹ of **ramps** can be described mathematically by (1).

Equation (1) can be used with signals that are differentiable twice. Also, we exclude saddle-points by requiring that the first time derivative is positive. The definition of **ramp** includes the concavity changing points of sigmoid functions like the hyperbolic tangent (as shown in fig. 7) and points at which the signal looks locally like a straight line (i.e. proportional to time), even if its concavity does not change sign.

Therefore, by obtaining an expression of the first and second time derivatives of the signal directly from the model, we can compute the locus of all **ramps** without solving the full differential equations or modelling time series.

4.3 Model structure

We apply our approach to models with the following structure:

$$\dot{\mathbf{x}} := \mathbf{M} \mathbf{r}(\mathbf{x}) + \mathbf{z} \quad (11)$$

$$\mathbf{y} := \mathbf{h}(\mathbf{x}) \quad (12)$$

where we used boldface, e.g. \mathbf{x} , for vectors, and capital typewriter font, e.g. \mathbf{M} for matrices. In equation (11), \mathbf{x} is the vector of states of the model, which herein we take to be a real column matrix, i.e. $\mathbf{x} \in \mathbb{R}^{N \times 1}$ (N is the number of states, e.g. $N = 13$ in the original **ASM1**, Henze et al. [39]). The states are then transformed via the mapping $\mathbf{r}(\mathbf{x})$, into a new vector, which herein we will call *rates* vector and, by slight abuse of notation, denote also \mathbf{r} . This transformation might not preserve

¹set of all points that satisfies the specified conditions

the dimension, so the resulting rates vector might have a different dimension $\mathbf{r}(\mathbf{x}) \in \mathbb{R}^{R \times 1}$ (R is the number of reaction rates, e.g. $R = 8$ in the original [ASM1](#), Henze et al. [39]).

The matrix \mathbf{M} combines the R rates to produce the N time derivatives of the states, $\dot{\mathbf{x}}$. Hence we have $\mathbf{M} \in \mathbb{R}^{N \times R}$. The additional term (\mathbf{z}) in equation (11) represents the *actuation* on the states of the system².

Equation (12) (a.k.a. *measurement model* [48, ch. 4.3]) represents a mapping that generates measurement signals, \mathbf{y} (modelling those that could be produced by a sensor), based on the model states (\mathbf{x}).

Until here we have omitted the dependence of equations (11)-(12) on parameters to keep the notation simple. All the model's components described till now can depend on parameters (free³ and fundamental).

Many models for activated sludge and anaerobic digestion obey this structure, for example, all the [ASM](#) family [26], and several anaerobic digestions models [49] such as [ADM1](#) [27]. As an example, all model components described above correspond to [ASM1](#) and can be found in [B.3](#), including tabs. 4 with all parameters and their description. This means that the mathematical analysis presented here can be applied to any model that follows the form in equations (11)-(12) by inserting the matrix and the rates of a different model than [ASM1](#) into the formula.

As the model evolves in time, equation (11) provides the first time derivative of the signals. The second time derivative of all states is obtained by direct application of the chain rule:

$$\ddot{\mathbf{x}} = \mathbf{M} \dot{\mathbf{r}} + \dot{\mathbf{z}} = \mathbf{M} \mathbf{J}_r \dot{\mathbf{x}} + \dot{\mathbf{z}} \quad (13)$$

where \mathbf{J}_r is the Jacobian of the rates w.r.t. the states, i.e.

$$\mathbf{J}_r := \begin{bmatrix} \nabla_{\mathbf{x}}^{\top} r_1(\mathbf{x}) \\ \vdots \\ \nabla_{\mathbf{x}}^{\top} r_R(\mathbf{x}) \end{bmatrix} = \begin{bmatrix} \partial_{x_1} r_1 & \cdots & \partial_{x_N} r_1 \\ \vdots & \ddots & \vdots \\ \partial_{x_1} r_R & \cdots & \partial_{x_N} r_R \end{bmatrix} \quad (14)$$

The [ramps](#) of the i -th signal in these models is then given by

$$\text{ramps} := \{t : \mathbf{M}_{i,:} \mathbf{r} + z_i > 0 \wedge \mathbf{M}_{i,:} \mathbf{J}_r \dot{\mathbf{x}} + \dot{z}_i = 0\} \quad (15)$$

4.4 Ramp contingency

Here the typical and the physical minimum and maximum values are presented for all eight relevant states that can cause the S_{O_2} [ramp](#) feature.

For all the evaluations in this article, we used Python 3 [50] and SymPy [51] (any other computer algebra system could be used), complemented by a minimal amount of manual analysis that will be further automated in the future [44].

Data availability

All data supporting the findings of this study were generated using code available in the public GitLab repository at <https://gitlab.com/sbrml/dsa-signal-features/>. The repository contains all scripts and instructions necessary to reproduce the datasets and figures presented in this article in the folder `doc/examples`. The code is available under the GNU Public License Version 3, and can be accessed without restriction.

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Authors contributions

MYS: Conceptualisation, Formal Analysis, Methodology, Funding Acquisition, Software, Validation, Visualisation, Writing - Original Draft, Review & Editing ET: Validation, Writing - Original Draft, Review & Editing JPC: Conceptualisation, Formal Analysis, Methodology, Software, Validation, Visualisation, Writing - Original Draft, Review & Editing

²The actuation might not be the inputs defined by a controller, a further *input model* (\mathbf{b}) might be needed: $\mathbf{z} := \mathbf{b}(\mathbf{u})$, where \mathbf{u} are the controllable inputs and \mathbf{z} are the actuations

³parameters that must be estimated, e.g. experimentally

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A Supplementary information

A.1 Time derivatives

Here we detail the calculation for the time derivative of a model with the following structure

$$\dot{\mathbf{x}} = \mathbf{M}\mathbf{r}(\mathbf{x}) \quad (16)$$

to which we introduce a new term in the derivatives

$$\dot{\mathbf{x}} = \mathbf{M}\mathbf{r}(\mathbf{x}) + \mathbf{z}(\mathbf{x}, t) \quad (17)$$

By direct derivation, the second time derivative reads (we do not show the explicit dependence anymore):

$$\ddot{\mathbf{x}} = \mathbf{M}\dot{\mathbf{r}} + \dot{\mathbf{z}} \quad (18)$$

$$\dot{\mathbf{r}} = \partial_{\mathbf{x}}\mathbf{r} \dot{\mathbf{x}} \quad (19)$$

$$\dot{\mathbf{z}} = \partial_{\mathbf{x}}\mathbf{z} \dot{\mathbf{x}} + \partial_t\mathbf{z} \quad (20)$$

replacing

$$\ddot{\mathbf{x}} = \mathbf{M}\partial_{\mathbf{x}}\mathbf{r} \dot{\mathbf{x}} + \partial_{\mathbf{x}}\mathbf{z} \dot{\mathbf{x}} + \partial_t\mathbf{z} \quad (21)$$

Another approach is to think that the introduction of the derivatives modifies the structure of the system in (16):

$$\dot{\mathbf{x}} = \mathbf{M}^*\mathbf{r}^*(\mathbf{x}) \quad (22)$$

with the new elements being concatenations of the old

$$\mathbf{M}^* = \begin{bmatrix} \mathbf{M} & \mathbb{I}_{\dim \mathbf{z}} \end{bmatrix} \quad (23)$$

$$\mathbf{r}^*(\mathbf{x}) = \begin{bmatrix} \mathbf{r}(\mathbf{x}) \\ \mathbf{z}(\mathbf{x}, t) \end{bmatrix} \quad (24)$$

where $\mathbb{I}_{\dim \mathbf{z}}$ is the identity matrix of the size of \mathbf{z} . Expanding:

$$\mathbf{M}^*\mathbf{r}^* = \begin{bmatrix} \mathbf{M} & \mathbb{I}_{\dim \mathbf{z}} \end{bmatrix} \begin{bmatrix} \mathbf{r} \\ \mathbf{z} \end{bmatrix} = \mathbf{M}\mathbf{r} + \mathbf{z} \quad (25)$$

which shows the equivalence to eq. (17). In this case the second derivative is,

$$\ddot{\mathbf{x}} = \mathbf{M}^*\dot{\mathbf{r}}^* = \mathbf{M}^*(\partial_{\mathbf{x}}\mathbf{r}^* \dot{\mathbf{x}} + \partial_t\mathbf{r}^*) \quad (26)$$

which gives a simple general formula applicable also in the absence of actuation.

To verify equality with the previous derivation we expand the terms. Computing the Jacobian

$$\partial_{\mathbf{x}}\mathbf{r}^* = \partial_{\mathbf{x}} \begin{bmatrix} \mathbf{r} \\ \mathbf{z} \end{bmatrix} = \begin{bmatrix} \partial_{\mathbf{x}}\mathbf{r} \\ \partial_{\mathbf{x}}\mathbf{z} \end{bmatrix} \quad (27)$$

then

$$\partial_{\mathbf{x}}\mathbf{r}^* \dot{\mathbf{x}} = \begin{bmatrix} \partial_{\mathbf{x}}\mathbf{r} \\ \partial_{\mathbf{x}}\mathbf{z} \end{bmatrix} \dot{\mathbf{x}} = \begin{bmatrix} \partial_{\mathbf{x}}\mathbf{r} \dot{\mathbf{x}} \\ \partial_{\mathbf{x}}\mathbf{z} \dot{\mathbf{x}} \end{bmatrix} \quad (28)$$

and

$$\mathbf{M}^*\partial_{\mathbf{x}}\mathbf{r}^* \dot{\mathbf{x}} = \begin{bmatrix} \mathbf{M} & \mathbb{I}_{\dim \mathbf{z}} \end{bmatrix} \begin{bmatrix} \partial_{\mathbf{x}}\mathbf{r} \dot{\mathbf{x}} \\ \partial_{\mathbf{x}}\mathbf{z} \dot{\mathbf{x}} \end{bmatrix} = \mathbf{M}\partial_{\mathbf{x}}\mathbf{r} \dot{\mathbf{x}} + \partial_{\mathbf{x}}\mathbf{z} \dot{\mathbf{x}} \quad (29)$$

The temporal term is

$$\mathbf{M}^*\partial_t\mathbf{r}^* = \begin{bmatrix} \mathbf{M} & \mathbb{I}_{\dim \mathbf{z}} \end{bmatrix} \begin{bmatrix} \partial_t\mathbf{r} \\ \partial_t\mathbf{z} \end{bmatrix} = \mathbf{M} \overbrace{\partial_t\mathbf{r}}^{\mathbf{0}} + \partial_t\mathbf{z} \quad (30)$$

Putting all together

$$\ddot{\mathbf{x}} = \mathbf{M}\partial_{\mathbf{x}}\mathbf{r} \dot{\mathbf{x}} + \partial_{\mathbf{x}}\mathbf{z} \dot{\mathbf{x}} + \partial_t\mathbf{z} \quad (31)$$

which is the same as eq. (21).

A.1.1 Single state actuation

If we assume that the actuation does not depend directly on time, and only depends in a single state, the actuation vector has the form:

$$\mathbf{z}(\mathbf{x}, t) = \mathbf{1}_{x_i} z(x_i) \quad (32)$$

where $\mathbf{1}_{x_i}$ is a column vector filled with zeros, except in the location of element x_i . That is, \mathbf{z} affects only the state x_i and it depends only on that state.

We have that

$$\partial_t \mathbf{z} = \mathbf{0} \quad (33)$$

$$\partial_{\mathbf{x}} \mathbf{z} \dot{\mathbf{x}} = \partial_{\mathbf{x}} \mathbf{1}_{x_i} z(x_i) \dot{\mathbf{x}} = \partial_{x_i} z \mathbf{1}_{x_i x_i} \dot{\mathbf{x}} = \partial_{x_i} z \dot{x}_i \mathbf{1}_{x_i} \quad (34)$$

where $\mathbf{1}_{x_i x_i}$ is a matrix with a 1 in the (x_i, x_i) element, and zero otherwise. By replacing into eq. (31) we obtain,

$$\ddot{\mathbf{x}} = M \partial_{\mathbf{x}} \mathbf{r} \dot{\mathbf{x}} + \partial_{x_i} z \dot{x}_i \mathbf{1}_{x_i} \quad (35)$$

and the x_i component is

$$\ddot{x}_i = M_{x_i} \cdot \partial_{\mathbf{x}} \mathbf{r} \dot{\mathbf{x}} + \partial_{x_i} z \dot{x}_i \quad (36)$$

These are the formulas used in the software [44] accompanying this publication.

B Ramp points curve

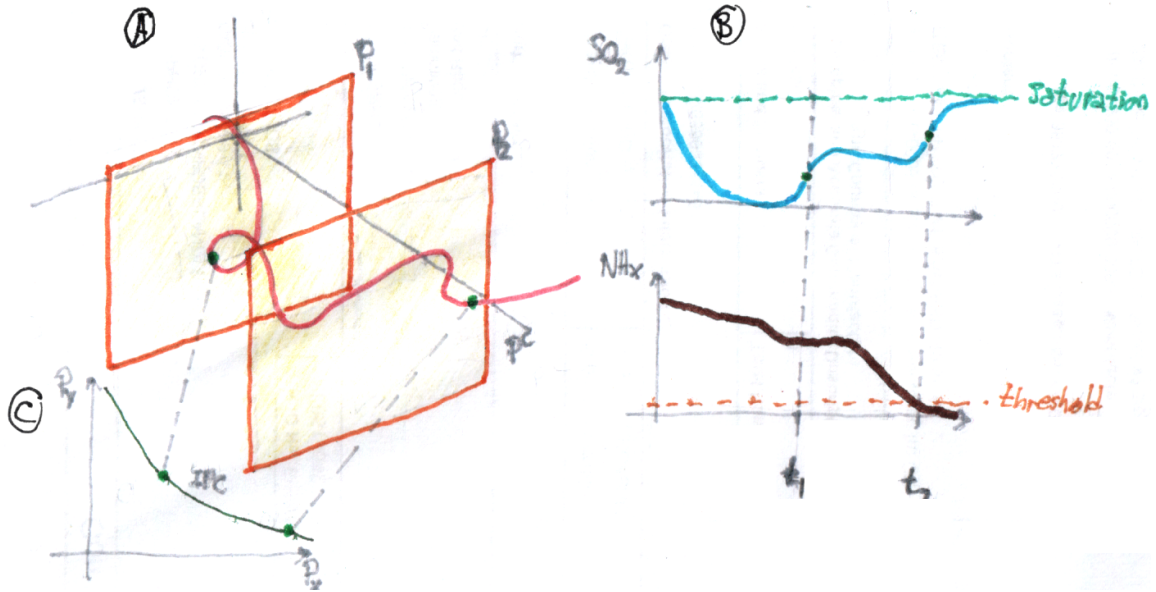


Figure 8: Construction and interpretation of a **ramp** points curve. A: a trajectory of a dynamical system in \mathbb{R}^N . In the drawing, the space is decomposed in two sub-spaces of dimension 2 (the planes are labelled with P in the diagram), and $N - 2$ (the perpendicular direction to the planes, the orthogonal complement P^\perp of P). By fixing the values of all components of a vector in P^\perp a plane is defined as shown. The trajectory crosses these planes as it evolves. B: Components of the trajectory can be plotted as a function of time, as an example the states S_{O_2} and NH_x from **ASM1** are shown. The points in which a ramp is realised are marked here and in the trajectory. The time of the ramps have their corresponding planes in panel A, t_1 with P_1 , etc. C: By merging all the planes and the points corresponding to a **ramp** in them, we obtain the locus of all ramps. These loci might look like curves in the obtained plot. The curve collects points from different planes (different values of the states in P^\perp)

The locus of all ramps defines a hyper-surface of dimension \mathbb{R}^{N-1} . This surface cannot be visualized easily. Therefore we choose a sub-space of dimension 2 that we call P (for plane) and we collect points fulfilling the ramp conditions in it.

In P we have two states of our choice, e.g. S_{O_2} and X_a , and all other states are in the orthogonal complement of P , called P^\perp . In fig. 8 we depict P^\perp as a single direction, but the reader should understand that in fact, this sub-space is of dimension $N - 2$.

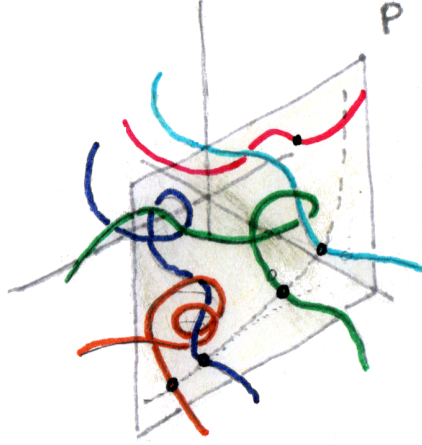


Figure 9: Multiple trajectories crossing the plane P , for fixed values of P^\perp . Some trajectories intersect the plane with a ramp in the time signal of the selected component, e.g. S_{O_2} , while others do not (pink trajectory). Only those with a ramp contribute to the ramp locus (dashed lines on P).

To put a ramp point in P , we must first define a value for all states in P^\perp . These are the planes shown in panel A of fig. 8. For any value we choose for these other states, and for any value of the states in P , we can always find a trajectory of the dynamical system that goes through the point in the full space. Hence, the reader should imagine each of these planes crossed by many trajectories as the ones shown here, each one intersecting the plane at a unique point. We then filter the intersection points by selecting those that fulfil the ramp condition. Then we repeat this process for different values of the states in P^\perp (different planes in the figure). We collect all these points in a single plane. These are the curves shown in fig. 5 in the body of the article.

The points shown in fig. 6 are obtained in a similar fashion. However, for these points, we do not fix any state. All states (including S_{NH_x}) are free, and to sample the hyper-surface defined by the ramp conditions (9) and (10), we solve an optimisation problem, which can be stated as follows: generate distinct points in a level set defined by a function $\phi : \mathbb{R}^n \rightarrow \mathbb{R}$ (in our case $n = 8$):

$$\mathcal{L}_c := \{x \in \mathbb{R}^n : \phi(x) = c\} \quad (37)$$

To warrant good behaviour of the set, we assume that the function has the following properties:

1. is continuous: implies closedness of the sets $\{x \in \mathbb{R}^n : \phi(x) \leq c\}$ (sublevel set)
2. $\nabla\phi(x) \neq 0 \quad \forall x \in \mathcal{L}_c$ (non-degenerate level set, level sets do not intersect)
3. is coercive: $\lim_{\|x\| \rightarrow \infty} \phi(x) = \infty$ (compact sublevel sets). That is: ϕ is coercive $\iff \forall c \in \mathbb{R}, \{x \in \mathbb{R}^n : \phi(x) \leq c\}$ is compact.

We can set the problem of sampling the ramp hyper-surface as a constrained optimization:

$$x = \arg \min_{x \in S \subset \mathbb{R}^n} f(x, \theta) \quad (38)$$

$$\phi(x) = 0 \quad \text{eq. (9)} \quad (39)$$

$$\psi(x) > 0 \quad \text{eq. (10)} \quad (40)$$

where S is chosen such that $\mathcal{L}_c \cap S \neq \emptyset$ (e.g. a hyperbox), and the cost function $f(x) : \mathbb{R}^n \rightarrow \mathbb{R}$ is arbitrary. The quality of the optimization results for our sampling can be verified by looking at the distributions shown in fig. 10. There the values of $\phi(x)$ are shown (second time derivative, which should be zero), together with the values of $\psi(x)$ (first time derivative, must be positive).

The cost function is exploited to achieve other objectives, like the spread of the points over the surface. For example:

$$f(x, \{x_i\}_{1 \leq i \leq I}, r_{\min}) = - \sum_{i=1}^I H(r_{\min} - \|x - x_i\|) \|x - x_i\|^2 \quad (41)$$

where $\{x_i\}_{1 \leq i \leq I}$ is a set of points in \mathcal{L}_c (e.g. the set of previous solutions to the problem). $H(r)$ is the step function, with value 1 when $r > 0$.

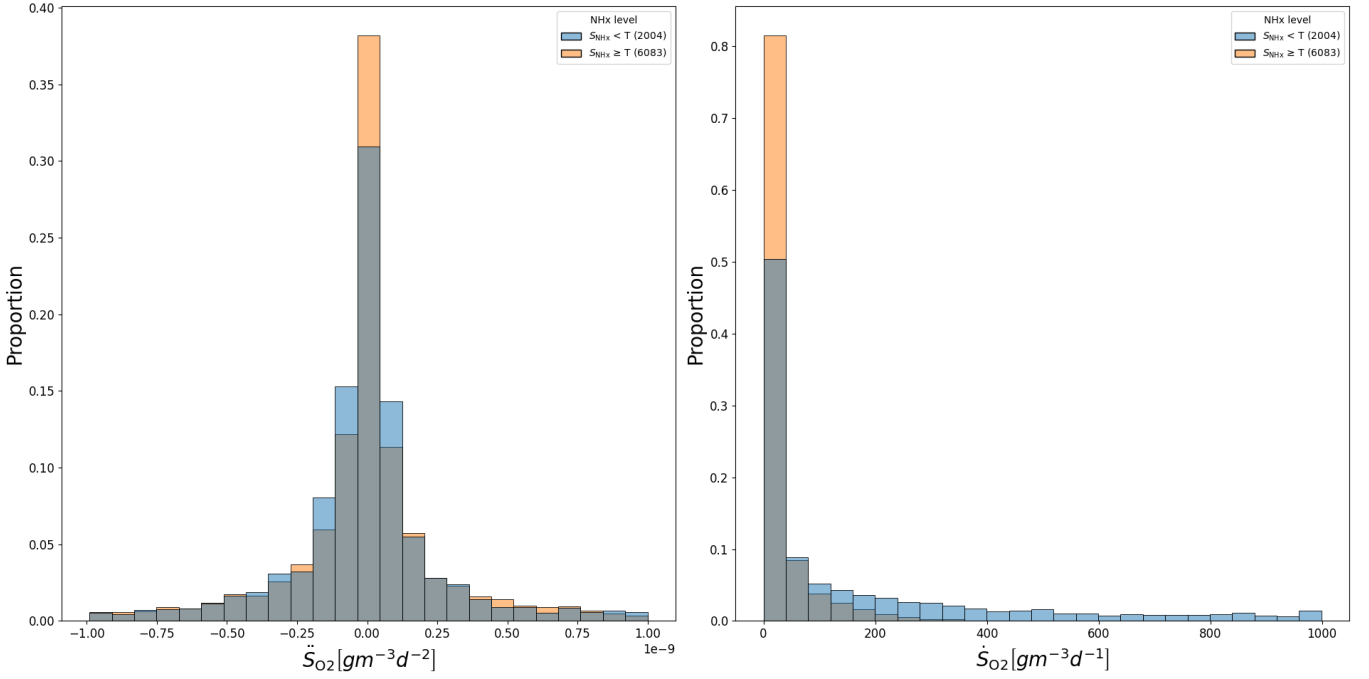


Figure 10: Values of the dissolved oxygen time derivatives of the selected samples from fig. 6

Numerical solutions using Sequential Quadratic Programming (SQP) showed an accumulation of solutions on the boundary of the intersection $\mathcal{L}_c \cap S$, likely due to the Karush–Kuhn–Tucker conditions. Other constrained optimizers are to be tested, but as a workaround, the cost function can be extended to

$$\tilde{f}(x, \{x_i\}_{1 \leq i \leq I}, \partial S) = f(x, \{x_i\}_{1 \leq i \leq I}) + \text{spike}(x, \partial S) \quad (42)$$

where spike is a function that increases rapidly as x approaches the boundary (∂S) of S . This workaround and a very tight hyperbox provided the results reported.

B.1 Interpretability of the inflection point curve

Figure 11 is the result from an interactive plot where points in the (X_h, S_{O_2}) plane can be selected and the time-series are reproduced from these points. The point on the blue line in the top figures are [ramps](#), the points away from the curve or instances where no ramp occurs. From these points then the dissolved oxygen signals are reconstructed. This illustrates the simulation time that we save by computing the inflection point curve directly. Hence, we evaluated [ramps](#) for 100,000 initial conditions in a few minutes on a normal laptop, which covers a large region of the state-space.

B.2 Symbols name

B.3 ASM1 model

$$\mathbf{x} = \begin{bmatrix} S_{nb} \\ S_b \\ X_{nb,in} \\ X_{cb} \\ X_h \\ X_a \\ X_{nb,e} \\ S_{O_2} \\ S_{NO_x} \\ S_{NH_x} \\ S_{bN} \\ X_{cbN} \\ S_{alk} \\ S_{N_2} \end{bmatrix} \quad (43)$$

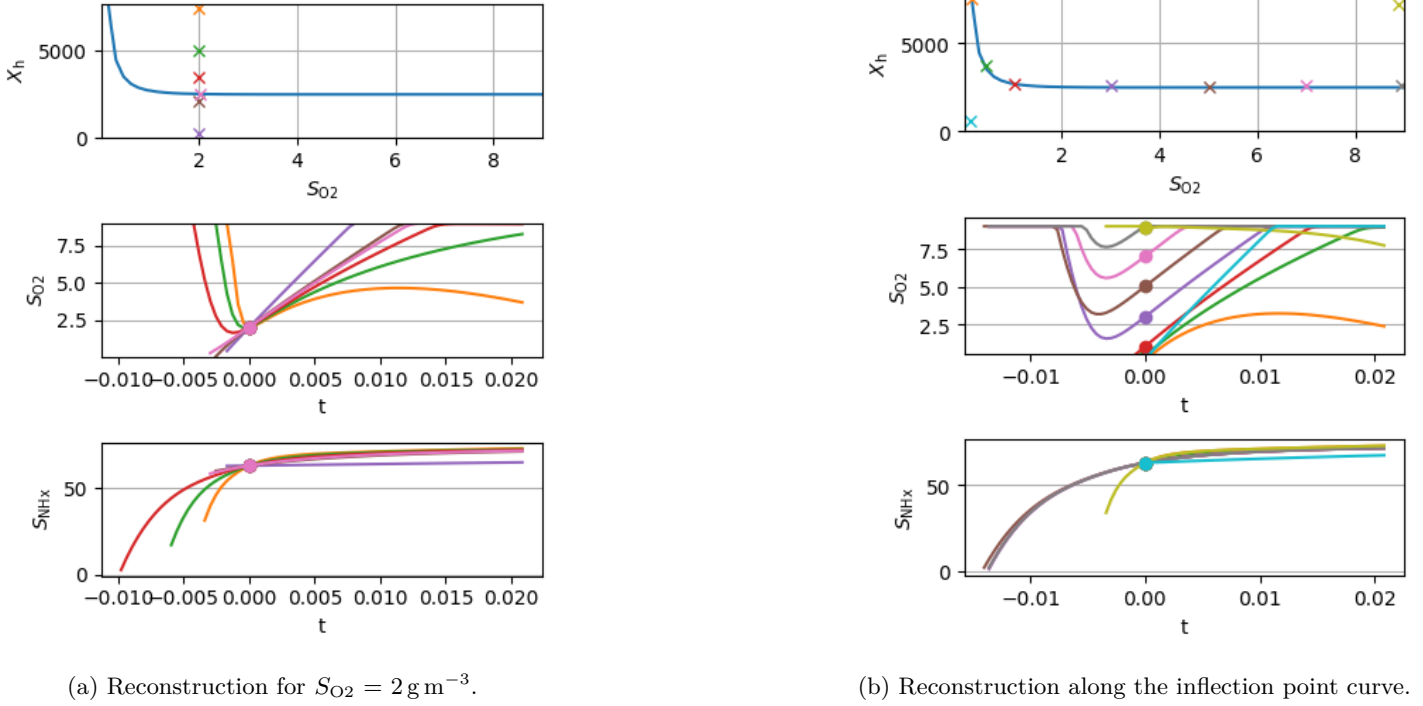


Figure 11: Reconstruction of the S_{O_2} and S_{NH_x} curve from selected points, marked with an x, in the S_{O_2} , X_h plane. In a) only one value is on the inflection point curve, hence has a **ramp**, in b) all but the two in the bottom left and upper right corner are combinations of states with ammonium above the threshold leading to **ramps**.

This work	Other name	Description
S_b	S_B	Soluble biodegradable organics (SS)
S_{nb}	S_U	Soluble nondegradable organics (SI)
S_{O_2}	S_{O_2}	Dissolved oxygen (SO)
X_{cb}	X_{CB}	Particulate and colloidal biodegradable organics (XS)
$X_{nb,in}$	X_{UInf}	Particulate nonbiodegradable organics from the influent (XI)
$X_{nb,e}$	X_{UE}	Particulate nonbiodegradable endogenous products (XP)
S_{NH_x}	S_{NH_x}	Ammonia ($NH_4 + NH_3$) (SNH)
S_{NO_x}	S_{NO_x}	Nitrate and nitrite ($NO_3 + NO_2$) (considered to be NO_3 only for stoichiometry, SNO)
X_{cbN}	X_{CBN}	Particulate and colloidal biodegradable organic N (XND)
S_{bN}	S_{BN}	Soluble biodegradable organic N (SND)
X_h	X_{OHO}	Ordinary heterotrophic organisms (XBH)
X_a	X_{ANO}	Autotrophic nitrifying organisms (NH_4^+ to NO_3^- , XBA)
S_{alk}	S_{Alk}	Alkalinity (HCO_3^- , SALK)
S_{N_2}	S_{N_2}	Dissolved nitrogen (gas, N_2), not in classic ASM1

Table 2: Naming of states in this work in relation to commonly used names, e.g. [26], which is provided in the bracket in the description. The "other name" column is based on [40].

This work	Other name	Description
g_{hO_2}	g_{hO_2}	Aerobic growth of heterotrophs
g_{hAn}	g_{hAn}	Anoxic growth of heterotrophs
g_{aO_2}	g_{aO_2}	Aerobic growth of autotrophs
d_h	d_h	Decay of heterotrophs
d_a	d_a	Decay of autotrophs
a_N	am_N	Ammonification of soluble organic nitrogen
h_o	ho	Hydrolysis of entrapped organics
h_{oN}	ho_N	Hydrolysis of entrapped organic nitrogen

Table 3: Naming of process rates and their description. The "other name" column is following [40].

$$\mathbf{M} = \begin{bmatrix}
0 & 0 & 0 & 0 & 0 & 0 & 0 & 0 \\
m_{S_b, g_{hO_2}} & m_{S_b, g_{hAn}} & 0 & 0 & 0 & 0 & 1 & 0 \\
0 & 0 & 0 & 0 & 0 & 0 & 0 & 0 \\
0 & 0 & 0 & m_{X_{cb}, d_h} & m_{X_{cb}, d_a} & 0 & -1 & 0 \\
1 & 1 & 0 & -1 & 0 & 0 & 0 & 0 \\
0 & 0 & 1 & 0 & -1 & 0 & 0 & 0 \\
0 & 0 & 0 & m_{X_{nb,e}, d_h} & m_{X_{nb,e}, d_a} & 0 & 0 & 0 \\
m_{S_{O_2}, g_{hO_2}} & 0 & m_{S_{O_2}, g_{aO_2}} & 0 & 0 & 0 & 0 & 0 \\
0 & m_{S_{NOx}, g_{hAn}} & m_{S_{NOx}, g_{aO_2}} & 0 & 0 & 0 & 0 & 0 \\
m_{S_{NHx}, g_{hO_2}} & m_{S_{NHx}, g_{hAn}} & m_{S_{NHx}, g_{aO_2}} & 0 & 0 & 1 & 0 & 0 \\
0 & 0 & 0 & 0 & 0 & -1 & 0 & 1 \\
0 & 0 & 0 & m_{X_{cbN}, d_h} & m_{X_{cbN}, d_a} & 0 & 0 & -1 \\
m_{S_{alk}, g_{hO_2}} & m_{S_{alk}, g_{hAn}} & m_{S_{alk}, g_{aO_2}} & 0 & 0 & m_{S_{alk}, a_N} & 0 & 0 \\
0 & m_{S_{N_2}, g_{hAn}} & 0 & 0 & 0 & 0 & 0 & 0
\end{bmatrix} \quad (44)$$

$$m_{S_b, g_{hO_2}} := -\frac{1}{\gamma_h} \quad (45)$$

$$m_{S_b, g_{hAn}} := -\frac{1}{\gamma_h} \quad (46)$$

$$m_{X_{cb}, d_h} := 1 - \zeta_{Xnb,l} \quad (47)$$

$$m_{X_{cb}, d_a} := 1 - \zeta_{Xnb,l} \quad (48)$$

$$m_{X_{nb,e}, d_h} := \zeta_{Xnb,l} \quad (49)$$

$$m_{X_{nb,e}, d_a} := \zeta_{Xnb,l} \quad (50)$$

$$m_{S_{O_2}, g_{hO_2}} := \frac{\gamma_h - 1}{\gamma_h} \quad (51)$$

$$m_{S_{O_2}, g_{aO_2}} := \frac{\gamma_a + \iota_{COD,NO_3}}{\gamma_a} \quad (52)$$

$$m_{S_{NOx}, g_{hAn}} := \frac{\gamma_h - 1}{\gamma_h \iota_{NO_3, N_2}} \quad (53)$$

$$m_{S_{NOx}, g_{aO_2}} := \frac{1}{\gamma_a} \quad (54)$$

$$m_{S_{NHx}, g_{hO_2}} := -\iota_{NXb} \quad (55)$$

$$m_{S_{NHx}, g_{hAn}} := -\iota_{NXb} \quad (56)$$

$$m_{S_{NHx}, g_{aO_2}} := -\iota_{NXb} - \frac{1}{\gamma_a} \quad (57)$$

$$m_{X_{cbN}, d_h} := \iota_{NXb} - \iota_{NXnb} \zeta_{Xnb,l} \quad (58)$$

$$m_{X_{cbN}, d_a} := \iota_{NXb} - \iota_{NXnb} \zeta_{Xnb,l} \quad (59)$$

$$m_{S_{alk}, g_{hO_2}} := -\iota_{NXb} \iota_{cSNHx} \quad (60)$$

$$m_{S_{alk}, g_{hAn}} := -\iota_{NXb} \iota_{cSNHx} + \frac{\iota_{cSNOx} (\gamma_h - 1)}{\gamma_h \iota_{NO_3, N_2}} \quad (61)$$

$$m_{S_{alk}, g_{aO_2}} := \iota_{cSNHx} \left(-\iota_{NXb} - \frac{1}{\gamma_a} \right) + \frac{\iota_{cSNOx}}{\gamma_a} \quad (62)$$

$$m_{S_{alk}, a_N} := \iota_{cSNHx} \quad (63)$$

$$m_{S_{N_2}, g_{hAn}} := \frac{1 - \gamma_h}{\gamma_h \iota_{NO_3, N_2}} \quad (64)$$

$$\quad (65)$$

$$\mathbf{r} = \begin{bmatrix}
g_{hO_2}(S_{NHx}, S_{O_2}, S_b, X_h) \\
g_{hAn}(S_{NHx}, S_{NOx}, S_{O_2}, S_b, X_h) \\
g_{aO_2}(S_{NHx}, S_{O_2}, X_a) \\
d_h(X_h) \\
d_a(X_a) \\
a_N(S_{bN}, X_h) \\
h_o(S_{NOx}, S_{O_2}, X_{cb}, X_h) \\
h_{oN}(S_{NOx}, S_{O_2}, X_{cbN}, X_{cb}, X_h)
\end{bmatrix} \quad (66)$$

$$g_{\text{hO}_2}(S_{\text{NH}_x}, S_{\text{O}_2}, S_b, X_h) := S_{\text{NH}_x} S_b X_h \mu_{\text{max},h} \frac{S_{\text{O}_2}}{(S_{\text{NH}_x} + \kappa_{\text{NH}_x,h})(S_{\text{O}_2} + \kappa_{\text{O}_2,h})(S_b + \kappa_b)} \quad (67)$$

$$g_{\text{hAn}}(S_{\text{NH}_x}, S_{\text{NO}_x}, S_{\text{O}_2}, S_b, X_h) := S_{\text{NH}_x} S_b X_h \mu_{\text{max},h} \frac{S_{\text{NO}_x} \eta_{\text{an},h} \kappa_{\text{O}_2,h}}{(S_{\text{NH}_x} + \kappa_{\text{NH}_x,h})(S_{\text{O}_2} + \kappa_{\text{O}_2,h})(S_b + \kappa_b)(S_{\text{NO}_x} + \kappa_{\text{NO}_x,h})} \quad (68)$$

$$g_{\text{aO}_2}(S_{\text{NH}_x}, S_{\text{O}_2}, X_a) := \frac{S_{\text{NH}_x} S_{\text{O}_2} X_a \mu_{\text{max},a}}{(S_{\text{NH}_x} + \kappa_{\text{NH}_x,a})(S_{\text{O}_2} + \kappa_{\text{O}_2,a})} \quad (69)$$

$$d_h(X_h) := X_h \beta_h \quad (70)$$

$$d_a(X_a) := X_a \beta_a \quad (71)$$

$$a_N(S_{\text{bN}}, X_h) := S_{\text{bN}} X_h \lambda_{\text{am}} \quad (72)$$

$$h_o(S_{\text{NO}_x}, S_{\text{O}_2}, X_{\text{cb}}, X_h) := X_{\text{cb}} \lambda_{\text{hyd},b} \frac{X_h}{X_{\text{cb}} + X_h \kappa_{X_{\text{hyd}}}} \left(\frac{S_{\text{NO}_x} \eta_{\text{hyd},\text{an}} \kappa_{\text{O}_2,h}}{(S_{\text{NO}_x} + \kappa_{\text{NO}_x,h})(S_{\text{O}_2} + \kappa_{\text{O}_2,h})} + \frac{S_{\text{O}_2}}{S_{\text{O}_2} + \kappa_{\text{O}_2,h}} \right) \quad (73)$$

$$h_{\text{oN}}(S_{\text{NO}_x}, S_{\text{O}_2}, X_{\text{cbN}}, X_{\text{cb}}, X_h) := X_{\text{cbN}} \lambda_{\text{hyd},b} \frac{X_h}{X_{\text{cb}} + X_h \kappa_{X_{\text{hyd}}}} \left(\frac{S_{\text{NO}_x} \eta_{\text{hyd},\text{an}} \kappa_{\text{O}_2,h}}{(S_{\text{NO}_x} + \kappa_{\text{NO}_x,h})(S_{\text{O}_2} + \kappa_{\text{O}_2,h})} + \frac{S_{\text{O}_2}}{S_{\text{O}_2} + \kappa_{\text{O}_2,h}} \right) \quad (74)$$

$$(75)$$

$$\dot{S}_b := g_{\text{hAn}} m_{S_b} g_{\text{hAn}} + g_{\text{hO}_2} m_{S_b} g_{\text{hO}_2} + h_o \quad (76)$$

$$\dot{X}_{\text{cb}} := d_a m_{X_{\text{cb}}} d_a + d_h m_{X_{\text{cb}}} d_h - h_o \quad (77)$$

$$\dot{X}_h := -d_h + g_{\text{hAn}} + g_{\text{hO}_2} \quad (78)$$

$$\dot{X}_a := -d_a + g_{\text{aO}_2} \quad (79)$$

$$\dot{S}_{\text{O}_2} := g_{\text{aO}_2} m_{S_{\text{O}_2}} g_{\text{aO}_2} + g_{\text{hO}_2} m_{S_{\text{O}_2}} g_{\text{hO}_2} \quad (80)$$

$$\dot{S}_{\text{NO}_x} := g_{\text{aO}_2} m_{S_{\text{NO}_x}} g_{\text{aO}_2} + g_{\text{hAn}} m_{S_{\text{NO}_x}} g_{\text{hAn}} \quad (81)$$

$$\dot{S}_{\text{NH}_x} := a_N + g_{\text{aO}_2} m_{S_{\text{NH}_x}} g_{\text{aO}_2} + g_{\text{hAn}} m_{S_{\text{NH}_x}} g_{\text{hAn}} + g_{\text{hO}_2} m_{S_{\text{NH}_x}} g_{\text{hO}_2} \quad (82)$$

$$\dot{S}_{\text{bN}} := -a_N + h_{\text{oN}} \quad (83)$$

$$\dot{X}_{\text{cbN}} := d_a m_{X_{\text{cbN}}} d_a + d_h m_{X_{\text{cbN}}} d_h - h_{\text{oN}} \quad (84)$$

$$(85)$$

$$\dot{X}_{\text{nb},e} := d_a m_{X_{\text{nb},e}} d_a + d_h m_{X_{\text{nb},e}} d_h \quad (86)$$

$$\dot{S}_{\text{alk}} := a_N m_{S_{\text{alk}}} a_N + g_{\text{aO}_2} m_{S_{\text{alk}}} g_{\text{aO}_2} + g_{\text{hAn}} m_{S_{\text{alk}}} g_{\text{hAn}} + g_{\text{hO}_2} m_{S_{\text{alk}}} g_{\text{hO}_2} \quad (87)$$

$$\dot{S}_{\text{N}_2} := g_{\text{hAn}} m_{S_{\text{N}_2}} g_{\text{hAn}} \quad (88)$$

$$(89)$$

$$\dot{S}_{\text{nb}} := 0 \quad (90)$$

$$\dot{X}_{\text{nb},\text{in}} := 0 \quad (91)$$

$$(92)$$

This work	Other name	Value	Units	Description
M_N	M_N	14.0	g mol^{-1}	atomic molar mass of nitrogen
COD_C	COD_C	32.0	g mol^{-1}	Theoretical COD of molar carbon
COD_{Fe}	COD_{Fe}	24.0	g mol^{-1}	Theoretical COD of molar iron
COD_H	COD_H	8.0	g mol^{-1}	Theoretical COD of molar hydrogen
COD_N	COD_N	-24.0	g mol^{-1}	Theoretical COD of molar nitrogen
COD_O	COD_O	-16.0	g mol^{-1}	Theoretical COD of molar oxygen
COD_P	COD_P	40.0	g mol^{-1}	Theoretical COD of molar phosphorus
COD_S	COD_S	48.0	g mol^{-1}	Theoretical COD of molar sulphur
COD_+	COD_{pos}	-8.0	g mol^{-1}	Theoretical COD of positive charge
COD_-	COD_{neg}	8.0	g mol^{-1}	Theoretical COD of negative charge
β_a	b_{ANO}	0.15	d^{-1}	Decay rate for XANO (b_A)
β_h	b_{OHO}	0.62	d^{-1}	Decay rate for XOHO (b_H)
$\eta_{\text{an,h}}$	n_{mOHOAx}	0.8	-	Reduction factor for anoxic growth of XOHO (η_g)
$\eta_{\text{hyd,an}}$	n_{qhydAx}	0.4	-	Correction factor for hydrolysis under anoxic conditions (η_h)
γ_a	Y_{ANO}	0.24	g g^{-1}	Yield of XANO growth per SNO3 (Y_A)
γ_h	Y_{OHO}	0.67	g g^{-1}	Yield for XOHO growth (Y_H)
$i_{\text{COD,N2}}$	i_{CODN2}	COD_N/M_N	g g^{-1}	Conversion factor for N2 in COD (i_{CODN2})
$i_{\text{COD,NO3}}$	i_{CODNO3}	$(\text{COD}_N + 3\text{COD}_O + \text{COD}_-)/M_N$	g g^{-1}	Conversion factor for NO3 in COD (i_{CODNO3})
$i_{\text{NO3,N2}}$	i_{NO3N2}	$(-3\text{COD}_O - \text{COD}_-)/M_N$	g g^{-1}	Conversion factor for NO3 reduction to N2 (i_{NO3N2})
i_{NXb}	i_{NXBio}	0.086	g g^{-1}	N content of biomass, meaning XOHO, XPAO, and XANO (i_{XB})
i_{NXnb}	i_{NXUE}	0.06	g g^{-1}	N content of products from biomass (i_{XE})
i_{cSNHx}	$i_{\text{ChargeSNHx}}$	$1/M_N$	mol g^{-1}	Conversion factor for NHx in charge ($i_{\text{ChargeSNHx}}$)
i_{cSNOx}	$i_{\text{ChargeSNOx}}$	$-1/M_N$	mol g^{-1}	Conversion factor for NO3 in charge ($i_{\text{ChargeSNOx}}$)
κ_b	K_{SBOHO}	20.0	g m^{-3}	Half-saturation coefficient for SB (K_S)
$\kappa_{\text{NHx,a}}$	K_{NHxANO}	1.0	g m^{-3}	Half-saturation coefficient for SNHx for XANO (K_{NH})
$\kappa_{\text{NHx,h}}$	K_{NHxOHO}	0.05	g m^{-3}	Half-saturation coefficient for NH4 (not in classic ASM1)
$\kappa_{\text{NOx,h}}$	K_{NOxOHO}	0.5	g m^{-3}	Half-saturation coefficient for SNOx XOHO (K_{NO})
$\kappa_{\text{O2,a}}$	K_{O2ANO}	0.4	g m^{-3}	Half-saturation coefficient for SO2 for XANO (K_{OA})
$\kappa_{\text{O2,h}}$	K_{O2OHO}	0.2	g m^{-3}	Half-saturation coefficient for SO2 XOHO (K_{OH})
κ_{Xhyd}	K_{XCBhyd}	0.03	g g^{-1}	Saturation coefficient for XB/XOHO (K_X)
λ_{am}	q_{am}	0.08	$\text{m}^3 \text{d}^{-1} \text{g}^{-1}$	Rate constant for ammonification (k_a)
$\lambda_{\text{hyd,b}}$	q_{XCBSBhyd}	3.0	g g^{-1}	Maximum specific hydrolysis rate of particulate and soluble biodegradable organics (k_h)
$\mu_{\text{max,h}}$	m_{OHOMax}	6.0	d^{-1}	Maximum growth rate of XOHO (μ_H)
$\mu_{\text{max,a}}$	m_{ANOMax}	0.8	d^{-1}	Maximum growth rate of XANO (μ_A)
$\zeta_{\text{Xnb,l}}$	f_{XUBiolys}	0.08	g g^{-1}	Fraction of XU generated in biomass decay (f_P)

Table 4: Parameters

State variable	min	reference	max	reference
S_b	0.9	reactor 5 [52]	69	[52]
S_{O_2}	2	[16]	2.2	[16]
X_{cb}	49	reactor 5 [52]	82	reactor 1 [52]
S_{NHx}	1.7	reactor 5 [52]	8	reactor 1 [52]
S_{NOx}	6	reactor 1 [52]	10	reactor 5 [52]
X_{cbN}	0.01	reactor 1 [52]	3.5	reactor 5 [52]
S_{bN}	0.7	reactor 5 [52]	2	reactor 1 [52]
X_h	2304	reactor 5 [52]	2816	reactor 1 [52]
X_a	135	reactor 5 [52]	165	reactor 1 [52]

Table 5: Scenario 1 for range of concentrations in the reactor that the wastewater treatment process is designed for. Most values are taken from the BSM1 [52]. The dissolved oxygen S_{O_2} concentration is an exception as the model is not for a sequencing batch reactor and we used the values that we observed in an sequencing batch reactor. With this selection method X_h and X_a have the same value which is not suitable for our approach. Hence we took for the maximum +10% and for the minimum -10% from the value in the BSM1 which are 2560 respectively 150.

State variable	min	reference	max	reference
S_b	0	-	2560	[53]
S_{O_2}	0	-	8	saturation concentration
X_{cb}	0.2	effluent [52]	91	mean of reactor 1 and underflow [52]
S_{NHx}	0	-	400	assumption based on [54]
S_{NOx}	0	-	400	assumption based on [54]
X_{cbN}	0.01	effluent [52]	6.5	mean of reactor 1 and underflow [52]
S_{bN}	0.7	reference	320	[55]
X_h	10	effluent [52]	3780	mean of reactor 1 and underflow [52]
X_a	0.6	effluent [52]	225	mean of reactor 1 and underflow [52]

Table 6: Scenario 2 for range of concentrations in the reactor which could happen in for example on-site wastewater treatment facilities. The assumption based on Dockhorn et al. [54] is made that all nitrogen produces per day is present as NHx respectively NOx and that 60 liters of water are consumed per day.

State variable	min	reference	max	reference
S_b	0	-	8200	observed maximum[56]
S_{O_2}	0	-	8	saturation concentration
X_{cb}	0	-	100	underflow [52]
S_{NHx}	0	-	300000	saturation concentration
S_{NOx}	0	-	9000	fertiliser production [57]
X_{cbN}	0	-	7	underflow [52]
S_{bN}	0	-	320	[55]
X_h	0	-	5000	underflow [52]
X_a	0	-	300	underflow [52]

Table 7: Scenario 3 for range of concentrations in the reactor which are physically possible, though many probably unlikely to happen.

C Causal diagram

Confounding situations are when one sees an association (correlation) but there is no causal relationship. A mediator M ($X \rightarrow M \rightarrow F$) might be present or the situation is spurious ($X \leftarrow F \rightarrow Y$), which means that there is no causal relation that leads to the correlation between the two elements X and Y. Another situation in causal diagrams is a collider ($X \rightarrow F \leftarrow Y$), which is also called a V-structure or immorality and is the structure that we observed between the different states of the ASM1 and the ramp feature. The V-structure means that not only the variable of interest X leads to feature F ($S_{NHx} <$ threshold), but also several other state variables such as Y.

C.1 Ammonium depletion soft-sensor based on the dissolved oxygen ramp feature

In a previous study [16], the soft sensor for a maintained dissolved oxygen sensor reached a 93% accuracy with the [ramp](#) feature for a balanced dataset where half of the data had an ammonium concentration below or equal to a threshold T and half the data was above. The arbitrary threshold was chosen as double the amount of the detection limit of the ammonium measurements in the laboratory (i.e. 1 g m^{-3}). For the unmaintained sensor, it was 80%. A high prediction accuracy could be achieved despite our study exposing alternative causes for the ramp.

In the previous study, we chose the most challenging real-world cases to which we were granted access. Additionally, we used synthetic, modelled data to make the feature fail to validate the feature and still obtained a wide range with high prediction accuracy [6].

Nevertheless, despite the rigour that we used, the results of the [DSA](#) in this article clearly advise caution when barely deciding on fit in data-matching if a prediction is good enough or not. Especially in the face of climate change or on-site applications where conditions outside the training data set are likely to occur. This supports the statement by [Zhong et al.](#) who show the power of data-driven tools, but also caution that they should not be overly trusted either. [DSA](#) hence should indeed be applied much more frequently to decide if a feature is really robust or needs to be further refined or combined with other features, as the [ramp](#) feature will have to be.