

# Multiscale modeling for food drying: A Downscaling approach

Zachary G. Welsh<sup>a</sup>, Vindya Thathsaranee Weligama Thuppahige<sup>a,b</sup> and M. A. Karim<sup>\*a</sup>

<sup>a</sup> Mechanical, Medical and Process Engineering, Queensland University of Technology,  
Brisbane, Australia

<sup>b</sup> Department of Food Science and Technology, Faculty of Agriculture, University of  
Ruhuna, Mapalana, Kamburupitiya, 81100, Sri Lanka

**Corresponding author:** Azharul Karim, Queensland University of Technology  
2 George Street, QLD 4000 Australia, [azharul.karim@qut.edu.au](mailto:azharul.karim@qut.edu.au)

## ABSTRACT

Modeling the cellular level transport of plant-based food materials during drying is very complex. The transportation of intracellular water (ICW) significantly influences the drying kinetics; however, it is rarely considered in computational models. One approach which can investigate the impact of ICW on drying is multiscale modeling. However, most multiscale models fail to consider ICW or predetermine its trend and transport. This work aims to develop a multiscale downscaling model which can actively downscale a materials condition (dependent variables) to investigate the transport of ICW. Two different drying temperatures (47°C and 64°C) and the ICW transport at three different points of interest within the domain are investigated. The results were compared with experimental data and a control model. The downscaling model predicts drying at 47°C very well achieving mean relative errors (MREs) of 2.4% and 0.53% for the average moisture content and average temperature respectively. At 64°C, the model also predicts drying well achieving MREs of 5.32% and 0.78% for the average

23 moisture content and average temperature respectively. The ICW at each point of interest was  
24 calculated and its trend through the cell wall revealed interesting findings.

25 **Keywords:** multiscale modeling, cell rupturing, food drying, downscaling

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## 27 **1. INTRODUCTION**

28 Plant-based food materials (PBFMs) have heterogeneous cellular structures comprising  
29 of three main cellular environments with each environment containing water of different  
30 characteristics: free water (FW) in intercellular spaces, intracellular water (ICW) in  
31 intracellular spaces and strongly bound water within the cell walls (Khan et al., 2018), Figure  
32 1. Drying PBFMs involves simultaneous heat and multiphase mass transfer which dynamically  
33 changes a material in terms of its properties, characteristics and final quality (Weligama  
34 Thuppahige et al., 2024). These changes originate within the heterogeneous cellular structure  
35 of the material and propagate across multiple length scales (cellular/microscale to  
36 tissue/macroscale). These changes are a direct result of the complex internal water migration  
37 during drying, specifically the transport of ICW is a major concern during drying due to its  
38 influence on materials cellular structure. However, the transport of ICW is complex as it  
39 depends on the spatial and temporal drying conditions in addition to the property changes the  
40 material is experiencing (Welsh et al., 2023). ICW can transport two main ways, apoplastic  
41 transport where ICW migrates from intracellular spaces to intercellular spaces after a cell  
42 membrane ruptures or through symplastic transport where ICW migrates from cell to cell via  
43 fine capillaries (Weligama Thuppahige et al., 2024). The use of computational mathematical  
44 models to understand and control the effects of migrating ICW is essential in designing optimal  
45 drying processes. However, ICW and its effects are rarely considered in mathematical models  
46 of food drying.

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**[Figure 1 can be placed here]**

Whitaker (1977) theory forms the foundation for many modern mathematical models. The theory derives a mechanistic macroscale model using the concept of a representative elementary volume (REV) considering the microscale heat, mass and momentum conservation equations of each phase (solid, liquid water, water vapor and air). However, Whitaker theory fails to consider the material's heterogeneity and assumes no ICW exists, rather it assumes all water is located outside the solid matrix. To overcome and compensate for this oversimplistic assumption, condition dependent properties are embedded within the computational models. Condition dependent properties are the properties which are accurate only when modeling the specific scenario/conditions in which the property was originally derived, for example properties which depend on a material's, drying conditions (i.e. air temperature, air speed) and/or the dryer configuration. Additionally, the further the simulated conditions deviate from the original conditions in which the property was derived, the less accurate the property becomes. One approach which can aid in transitioning away from condition-dependent properties while considering ICW is multiscale modeling. Multiscale modeling considers a series of sub-models over multiple length scales, giving it the ability to incorporate large amount of physics for a reasonable computational cost (Welsh et al., 2018). There are two main types of multiscale models, one way coupled models and two-way coupled (concurrent) models (Welsh et al., 2018). A one way coupled multiscale model transfers information in one direction, either through upscaling or downscaling. In such models, properties are often computed considering the structure of the particular material and upscaled to inform a larger scale. Alternatively, dependent variables can be downscaled to analyze the behavior at a smaller scale. Concurrent or two way coupled models, on the other hand, allow bidirectional information exchange. Here, data flows back and forth between scales, with each scale

72 evolving simultaneously and influencing the other. This concurrent exchange includes  
73 upscaling, downscaling, and mutual information exchange throughout the simulation, enabling  
74 both scales to adjust based on the other's evolving state (Welsh et al., 2018). Multiscale  
75 modeling has gained recent attention in the field of food drying modeling with various  
76 multiscale model being developed based on various assumptions. Qiu et al. (2022) developed  
77 a pore scale multiscale model for impingement drying of a porous slab. The work demonstrated  
78 the strength of predicting the drying performance through their multiscale approach to transport  
79 properties rather than the traditional condition dependent approach. Tan et al. (2023) also  
80 developed a pore-scale model for the heat-moisture and stress-stain distributions for high-  
81 moisture porous media. However, these works failed to consider ICW as all water considered  
82 was located in the pores/capillaries rather than the solid material.

83 Over the years there has been a few cellular models and multiscale models which have  
84 considered ICW within the solid material. Fanta et al. (2013) developed a microscale model to  
85 better understand the underlying phenomena of pear dehydrating under isothermal conditions  
86 at 25 °C. The work considered water in each cellular environment, allowing the work to  
87 investigate the different pathways water can flow. The largest gradients in water content were  
88 observed across the cell walls and cell membranes. Rahman et al. (2018) also developed a  
89 micro-level transport model for drying apple parenchyma cells, considering ICW and  
90 conducted an in-depth sensitivity investigation. Prawiranto et al. (2018) constructed a cellular  
91 3D continuum model for the dehydration of apple. The study considered water in various  
92 cellular environments and modelled dehydration with a predetermined set of cellular  
93 changes/conditions (Turgid, free shrinkage, lysis and plasmolysis). However, these works  
94 developed only cellular models rather than multiscale models, utilizing a single microscale  
95 domain to represent the transport occurring and often modelled predetermined  
96 conditions/scenarios for deformation and/or relative humidity. During drying, significant

97 spatial and temporal variations (in terms of moisture and temperature) develop within a product  
98 which cannot be fully understood/investigated with a singular microscale domain. These  
99 variations are especially evidence in the later stages of drying when the inner can be fully  
100 wet/saturated, whereas the outer cells of the material can be fully dried and deformed. Recently,  
101 Welsh et al. (2021a) developed a multiscale model for predicting the diffusivity of apple  
102 through upscaling during convective drying. The work considered ICW coupled with  
103 predetermined temperature dependent domains. The authors later extended their multiscale  
104 approach to inversely calculate the temporal cellular diffusivity of ICW in apples (Welsh et al.,  
105 2021b) and to construct a generalized diffusivity tensor which is applicable to multiple  
106 materials and conditions (Welsh et al., 2023). However, again their work focused on multiscale  
107 models which upscaled properties by utilizing predetermined microscale domains and/or  
108 considered every possible microscale scenario rather than predicting the spatiotemporal  
109 conditions occurring throughout the material. To predict the spatiotemporal conditions the  
110 cellular structure is experiencing a multiscale model which is capable of downscaling is  
111 required.

112 This work aims to develop a multiscale model capable of downscaling the dependent variables  
113 to appropriately predict the spatiotemporal conditions a material's cellular structure  
114 experiences during drying. The work investigates two different drying temperatures (47°C and  
115 64°C) and focuses on the microscales at three different points of interest within the tissue  
116 domain. Additionally, the trend of ICW and its flux through the cell wall is investigated. The  
117 downscaling approach also incorporated cellular shrinkage through the shrinkage velocity  
118 approach paired with X-ray Micro-Computed Tomography (X-ray  $\mu$ CT) data to future improve  
119 the generalizability of the multiscale model. The modeling results are also compared to  
120 experimental data and a single scale control model.

## 121 **2. Model Development**

### 122 *2.1 Drying model*

123 A preliminary multiscale model was previously developed considering two spatial  
124 scales, a microscale and a macroscale (Welsh et al., 2021a), Figure 2. However, the previous  
125 model was only capable of upscaling information, specifically diffusivity, using predetermine  
126 conditions at the microscale. This study extends the preliminary model, allowing the model to  
127 downscale the required variables and evolve the microscale with time to investigate the  
128 different spatiotemporal conditions the cellular structure experiences throughout the entire  
129 material. Additionally, the ICW content required to calculate the diffusivity [Equation (16)], is  
130 calculate from the size of the intracellular space, outlined in 2.2.6, which is unique to this study.  
131 The modeling methods will present the multiscale governing equations, boundary conditions,  
132 properties, structure representation at each scale, computational approach, and temporal  
133 coupling. The modeling domains for both scales can be found in Figure 2. Additionally, the  
134 three points of interest, point 1 (P1), point 2 (P2) and point 3 (P3) are evenly distributed  
135 horizontally across the center of the macroscale [denoted by the red dots Figure 2 (a)].

136 **[Figure 2 can be placed here]**

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### 138 *2.2 Structure Representation at each scale*

139 The macroscale can be considered as homogeneous following the common approach in  
140 literature, however, careful consideration needs to be taken when it comes to the microscale.  
141 Two common cellular structure definitions/classifications exist in the literature, (1)  
142 macroscopically connected and (2) disconnected inclusions, each effecting the model's  
143 assumptions and physics. Over the years, Perré (2019) has considered timber as connected  
144 inclusions for their multiscale models, resulting in an additional "secondary flux" of the

145 divergence of the microscopic flux being considered in the macroscale governing equations  
146 when completing the scaling (Perré, 2007, 2019). Karunasena et al. (2015) considered the food  
147 material as hexagon shaped cells evolving the porosity through removing some cells from the  
148 arrangement for their meshfree models. The resulting structure appears to be disconnected  
149 inclusions, though the work does not scale any results. Prawiranto et al. (2018) investigated the  
150 transport and cellular changes of a single cuboctahedron shaped apple cell during drying. The  
151 cell was located in the center of their 3D domain with voids (or pores) in all eight corners of  
152 the cube, however, only basic volume averaging was considered to interpolate the results to the  
153 macroscale. Fanta et al. (2014) and Rahman et al. (2018) considered an arrangement of multiple  
154 intracellular and intercellular spaces while modeling the transport within the cellular structure  
155 of PBFMs. The resulting structure is considered disconnected inclusions, though these works  
156 also considered cell walls within the domains. Additionally, these works only model the  
157 microscale transport phenomena and no scaling was conducted. In reality, fresh PBFMs  
158 generally contain disconnected inclusions (pores) and during drying the structure evolves and  
159 if cell rupturing occurs the inclusions will become connected. As a result, the model will  
160 consider the structure as connected inclusions, however, as the model is being applied to  
161 convective drying, internal convective flow can be neglected and the “secondary flux”  
162 representing the flow through the cell structure is not considered. Nevertheless, if the multiscale  
163 approach was applied to model other drying techniques (such as microwave drying) the  
164 secondary flux would need to be considered.

165         The size of the microscale is important due to its effect on computational cost and  
166 possible influence on the results. Additionally, the microscale must accurately represent the  
167 dynamic microstructure, specifically the domain must represent the two key characteristics  
168 which dynamically change during drying, namely the porosity and free/bound water.  
169 Prawiranto et al. (2020) consider multiple domain sizes for dehydrating apple concluding

170 0.5mm cubed to be sufficient to accurately represent the structure. Carr et al. (2013) developed  
171 a multiscale model for timber utilizing a microscale size of 50  $\mu\text{m}$  with two sub-domains, a  
172 cell and its cell wall. With this in mind, the microscale has a size of 250  $\mu\text{m}$  with an initial cell  
173 size of 215  $\mu\text{m}$  calculated from the obtained X-ray  $\mu\text{CT}$  images, Figure 2.

### 174 2.2.1 Macroscale conservation equations

175 In an axisymmetric coordinate system, the macroscale transport model was developed  
176 with the following assumptions: (1) internal convective flow and heat generation are negligible,  
177 (2) drying air properties remain constant, (3) heating is axisymmetric, (4) moisture evaporates  
178 solely from the surface, and (5) thermal equilibrium is maintained across all phases. The  
179 macroscale mass and energy conservation equations are derived using Fick's law of diffusion  
180 and Fourier's law of heat transfer. These equations determine the moisture concentration  $C$   
181 ( $\text{mol}/\text{m}^3$ ) and temperature  $T$  (K) as functions of position and time. The respective equations are  
182 as follows (Golestani et al., 2013).

$$183 \quad \frac{\partial C}{\partial t} + \frac{1}{r} \frac{\partial}{\partial r} \left[ -D_{H,eff} r \frac{\partial C}{\partial r} \right] + \frac{\partial}{\partial z} \left[ -D_{H,eff} \frac{\partial C}{\partial z} \right] = 0 \quad , \quad (1)$$

$$184 \quad \rho c_p \frac{\partial T}{\partial t} + \frac{1}{r} \frac{\partial}{\partial r} \left[ -k r \frac{\partial T}{\partial r} \right] + \frac{\partial}{\partial z} \left[ -k \frac{\partial T}{\partial z} \right] = 0 \quad , \quad (2)$$

185 where  $t$  is the time (s),  $\rho$  represents the density of the food tissue ( $\text{kg}/\text{m}^3$ ),  $c_p$  is the specific heat  
186 of the food tissue ( $\text{J}/(\text{kg}\cdot\text{K})$ ),  $k$  denotes the thermal conductivity of the food material ( $\text{W}/(\text{m}\cdot\text{K})$ )  
187 and  $D_{H,eff}$  is the upscaled effective moisture diffusivity ( $\text{m}^2/\text{s}$ ). The effective moisture diffusivity  
188 varies with temperature and  $WC_{ICW}$ , the ICW content (%).

### 189 2.2.2 Boundary conditions

190 The macroscale boundary conditions for the mass flux and heat transfer at the exterior  
191 surface was given by,

192 
$$\mathbf{n} \cdot [-D_{H,eff} \nabla C] = h_m \frac{(p_{v,eq} - p_{v,air})}{RT}, \quad (3)$$

193 
$$\mathbf{n} \cdot [-k \nabla T] = h_T (T - T_{air}) - h_m \frac{(p_{v,eq} - p_{v,air})}{RT} h_{fg} M_w, \quad (4)$$

194 where  $h_m$  is the mass transfer coefficient (m/s),  $h_T$  is the heat transfer coefficient (W/(m<sup>2</sup>·K)),  
 195  $p_{v,eq}$  is the equilibrium vapor pressure,  $p_{v,air}$  is the vapor air pressure of ambient air (Pa),  $\mathbf{n}$  is  
 196 the unit vector normal to the boundary,  $R$  is the universal gas constant (J/(mol·K)),  $T_{air}$  is the  
 197 drying air temperature (K),  $M_w$  molar mass of water (g/mol) and  $h_{fg}$  is the latent heat of  
 198 evaporation (J/kg). The equilibrium vapor pressure for apple can be determined from the  
 199 sorption isotherm. Thus, the equilibrium vapor pressure for apple ( $p_{v,eq,apl}$ ) is (Ratti et al., 1989),

200 
$$p_{v,eq,apl} = p_{v,sat}(T) \exp(-0.182 M_{db}^{-0.696} + 0.232 e^{-43.949 M_{db}} M_{db}^{0.0411} \ln[p_{v,sat}(T)]). \quad (5)$$

201 The saturated vapor pressure is given by (Vega-Mercado et al., 2001),

202 
$$p_{v,sat} = \exp \left[ \frac{-5800.2206}{T} + 1.3915 - 0.0486 T + \right. \\ \left. 0.4176 \times 10^{-4} T^2 - 0.1445 \times 10^{-7} T^3 + 6.546 \ln(T) \right]. \quad (6)$$

203 The last boundary condition for the macroscale is the symmetric condition applied at  
 204  $r=0$ , defined as

205 
$$\mathbf{n} \cdot [-D_{H,eff} \nabla C] = 0, \quad (7)$$

206 
$$\mathbf{n} \cdot [-k \nabla T] = 0. \quad (8)$$

### 207 2.2.3 Microscale mass conservation

208 To develop the multiscale model, a microscale was defined at each point within the  
 209 macroscale to represent the heterogeneous structure of the food. This approach is valid when  
 210 the scale parameter approaches zero, causing the heterogeneities to vanish and resulting in a

211 homogeneous material (Pavliotis & Stuart, 2008; Welsh et al., 2021a). This assumption is  
 212 crucial for deriving a model through homogenization and is often referred to as the separation  
 213 of scales (Auriault et al., 2010; Carr & Turner, 2014). The microscale considered two sub-  
 214 domains, ICW denoted as  $\omega_{ICW}$  and intercellular water [free water (FW)] denote as  $\omega_{FW}$ , Figure  
 215 2 (b). The macroscale mass transport, Equation (1), is coupled to the microscale mass transport  
 216 equation defined as,

$$217 \quad \frac{\partial c}{\partial t} + \frac{\partial}{\partial x} \left( -D \frac{\partial c}{\partial x} \right) + \frac{\partial}{\partial y} \left( -D \frac{\partial c}{\partial y} \right) = 0, \quad (9)$$

218 where  $c$  is the microscale concentration (mol/m<sup>3</sup>) and  $D$  is the cellular diffusivity (m<sup>2</sup>/s)  
 219 equaling  $D_{ICW}$  if  $x,y \in \omega_{ICW}$  and  $D_{FW}$  if  $x,y \in \omega_{FW}$ .

220

#### 221 2.2.4 Microscale boundary conditions

222 The coupling between the macroscale and microscale variables occurs at the boundary  
 223 of the microscale given by,

$$224 \quad c_{micro}(a, y) = c_{micro}(0, y) + a \frac{\partial C_{\psi}}{\partial r}, \quad 0 < y < b \quad (10)$$

$$225 \quad c_{micro}(x, b) = c_{micro}(x, 0) + b \frac{\partial C_{\psi}}{\partial z}, \quad 0 < x < a \quad (11)$$

226 where  $a$  and  $b$  are the size of the microscale domain and  $\psi$  is the point of interest. The fluxes  
 227 at the boundaries are also coupled through,

$$228 \quad \begin{aligned} q_c(0, y) \cdot \mathbf{n} &= q_c(a, y) \cdot \mathbf{n}, & 0 < y < b \\ q_c(x, 0) \cdot \mathbf{n} &= q_c(x, b) \cdot \mathbf{n}. & 0 < x < a \end{aligned} \quad (12)$$

229 The boundary condition for the cell wall is

230 
$$\mathbf{n} \cdot [-D_{H,eff} \nabla C] = P_m (c - C), \quad (13)$$

231 where  $P_m$  is the permeability of the cell wall. The permeability of the cell wall does have some  
 232 uncertainty as the thickness of the cell wall changes during drying (Joardder et al., 2015) and  
 233 as such is investigated in a sensitivity investigation.

234 *2.2.5 Macroscale and microscale diffusivity*

235 To complete the multiscale model the diffusivity for the macroscale and cellular  
 236 subdomains is required. The ICW diffusivity is (Welsh et al., 2021b),

237 
$$D_{ICW} = \exp[-110.32 + 0.55536T_{cell} + 0.095148WC_{ICW} - 0.00090738T_{cell}^2 - 0.0007235WC_{ICW}^2],$$
  
 238 (14)

239 where  $T_{cell}$  is the cell temperature (K) and  $WC_{ICW}$  is the intracellular water content (mass %).  
 240 The diffusivity of FW is (Pace, 1962),

241 
$$D_{FW} = 2.26 \times 10^{-5} \left[ \frac{T}{273.15} \right]^{1.81}. \quad (15)$$

242 The model does not actively upscale the cellular properties to form the macroscale  
 243 diffusivity, rather the previously developed homogenized diffusivity is utilized (Welsh et al.,  
 244 2021a). The function for the macroscale diffusivity is (Welsh et al., 2023),

245 
$$D_{H,eff} = \exp[-106.6 + 0.5548T_{cell} + 0.029502WC_{ICW} - 0.00090648T_{cell}^2 - 0.00041019WC_{ICW}^2].$$
  
 246 (16)

247 *2.2.6 Evolution of intracellular water- rupturing threshold*

248 The ICW diffusivity function [Equation (14)] incorporates all three transport  
 249 mechanisms (Welsh et al., 2023), therefore, cell rupturing must be incorporated within the  
 250 evolution of the ICW. Welsh et al. (2023) introduced a rupturing threshold, exploiting the

251 macroscale equilibrium vapor pressure to determine if and when cell rupturing occurs within  
252 the material. Once the average macroscale equilibrium vapor pressure of the domain reaches  
253 the threshold, cells will begin to rupture. However, once the threshold was reached, Welsh et  
254 al. (2023) work utilized time dependent functions for ICW which require sophisticated  
255 microscale experimental techniques (such as nuclear magnetic resonance). This significantly  
256 limits the approaches applicability to other studies as these techniques are costly and not  
257 commonly available. As the current multiscale downscaling model includes an evolving  
258 microscale, the evolution of ICW can now be determined directly from the microscale.  
259 Therefore, when the cell is at its full size (215 $\mu$ m), the material is at its maximum ICW water  
260 content, 87.8% for apple (Khan et al., 2016). Once the cell rupturing threshold is reached the  
261 ICW is determined considering the change in cell size. The threshold of apple is considered as  
262 7080 Pa for apple (Welsh et al., 2023).

### 263 2.2.7 *Intracellular deformation*

264 A shrinkage velocity approach was utilized to approximate the shrinkage of the  
265 intracellular space given by (Tuly et al., 2023),

$$266 \quad \frac{\partial X}{\partial t} \cdot \mathbf{n} = v_n , \quad (17)$$

267 where  $v_n$  is the shrinkage velocity.

### 268 2.2.8 *Other input parameters*

269 The remaining input parameters are presented in Table 1.

270 **[Table 1 can be placed here]**

271

## 272 2.3 *Simulation Methodology*



297 2.4 *Experimental Investigations*

298 Two separate experimental investigations were conducted, the macroscale drying  
299 experiments and X-ray  $\mu$ CT. The macroscale drying experiments were conducted to produce  
300 validation data to validate the output from the multiscale model whereas the X-ray  $\mu$ CT was  
301 conducted to investigate the evolution of the material's microstructure.

302 2.4.1 *Macroscale drying experiments*

303 Granny Smith apples at commercial maturity were purchased from a local supermarket  
304 and prepared following the standard sample preparation procedure (Asabe & Home). The  
305 samples were washed and then cut into cylindrical shapes using a stainless-steel Cork Borer  
306 (JH-1225). Each sample consisted of the mesocarp (pulp) portion, initially measuring 30 mm  
307 in length and 18 mm in diameter. Convective drying was performed at two selected  
308 temperatures within the typical air temperature range for drying (40–80 °C): 47 °C and 64 °C,  
309 representing different levels of cellular deformation and ICW transport mechanisms. The dryer  
310 maintained an air velocity of 1 m/s and the airflow was perpendicular to the sample. Model  
311 validation was conducted using two data sets: average moisture content and average surface  
312 temperature of the samples. To measure moisture content, each sample was weighed  
313 individually at regular intervals. This occurred every 10 minutes during the first hour, then  
314 every 30 minutes until the drying process was completed. The samples were weighed using a  
315 digital balance with a 50 g capacity and  $\pm 0.001$  g accuracy, with measurements completed  
316 within 10 seconds. A FLIR-E6390 thermal imaging camera was used to record surface  
317 temperature. This experimental procedure was repeated three times. The detailed drying  
318 experimental procedure and some of the results can be found in the authors' previous  
319 publication (Welsh et al., 2023).

320

321 2.4.2 *X-ray  $\mu$ CT Experiment*

322 In order to extract the cellular deformation of dried apple cells, both the fresh and  
323 convective dried (both at 45 °C and 60 °C) cylindrical-shaped Granny Smith apple samples  
324 were scanned using an X-ray  $\mu$ CT scanner ( $\mu$ CT50, Scanco Medical, Switzerland). Apple  
325 samples were packed in foam and horizontally placed in scanning tubes with a diameter of  
326 34mm. The native resolution, electric potential, current and the power used for these scans  
327 were as 10.3  $\mu\text{m}^3$ , 55 kVp, 145  $\mu\text{A}$ , and 8 W, respectively with an average sample scanning  
328 time of 1000 ms and an Aluminum filter (0.1 mm). The scans were carried out in triplicates.  
329 Once the scans were completed, the image stacks were exported as DIACOM files into Mimics  
330 (Version 24.0, Materialise NV, Belgium) for segmentation using semi-automated thresholding.  
331 The consistency of the segmentations was ensured by maintaining a constant threshold value.  
332 It should be noted due to the study requirements and equipment limitations the X-ray  $\mu$ CT  
333 experimentation was conducted on cylindrical samples with a height of 15 mm and a diameter  
334 of 8mm 15 mm and a diameter of 8mm drying at an average temperature of 45°C and 60°C.

335

### 336 **3. Results**

337 The results are presented into five main sections, the cellular deformation, the  
338 macroscale drying kinetics, the macroscale temperature evolution, the ICW results and a  
339 sensitivity investigation. Three points of interest within the macroscale domain, (P1, P2 and  
340 P3) were selected along the center of the domain to analyses the model and approach, visually  
341 shown in Figure 2. These points were selected as examples to demonstrate how the microscales  
342 vary spatially depending on its location in the macroscale. The multiscale downscaling model  
343 was compared to the experimental data and the single scale macroscale model as a control  
344 (Welsh et al., 2023). All properties and heat and mass transfer coefficients were the same for  
345 the control and downscaling model.

346 3.1 Cellular deformation

347 X-ray  $\mu$ CT was utilized to extract the cellular deformation information of granny smith  
348 apple dried at 45°C and 60°C. Evaluating 30 intracellular spaces resulted in averages sizes of  
349  $215.4 \pm 30 \mu\text{m}$  to fresh apples cells,  $128.15 \pm 10 \mu\text{m}$  for dried apple cells with convective air  
350 at 45°C to 0.8 kg/kg dry bases and  $119.10 \pm 10 \mu\text{m}$  for dried apple cells with convective air at  
351 60°C to 1.23 kg/kg dry bases, Figure 5. This resulted in, using Equation (17), temperature  
352 dependent shrinkage velocities of  $-25 \times 10^{-9} \text{ m/s}$  and  $-30 \times 10^{-9} \text{ m/s}$  for drying at 47°C and apple  
353 dried 64°C respectively. It should be noted, the shrinkage velocity also had to consider the  
354 multiscale temporal coupling.

355 **[Figure 5 can be placed here]**

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357

358 3.2 Macroscale - Drying Kinetics

359 The downscaling model accurately predicted the moisture transport at 47°C achieving a mean  
360 relative error (MRE) of 2.4% when compared to the experimental results (Welsh et al., 2023),  
361 Figure 6. The control model (Welsh et al., 2023) achieved a similarly low MRE of 1.52%. The  
362 multiscale downscaling model slightly overpredicts the experimental data consistently till the  
363 later stages of drying. In the later stage, the model slightly underpredicts the transport matching  
364 the trend of the control model. The downscaling model and control model resulted in almost  
365 identical diffusivities between  $1.94 \times 10^{-10}$  and  $2.16 \times 10^{-10} \text{ m}^2/\text{s}$  for drying at 47°C, Figure 7. At  
366 64°C the downscaling model predicted the moisture transport accurately achieving a MRE of  
367 5.32% when compared to the experimental results, Figure 6. The control macroscale model  
368 achieved a similarly low MRE of 4.80%, predicting the experimental data slightly better. The  
369 multiscale downscaling model slightly overpredicted the experimental data between 30-180

370 mins for drying at 64°C and then slightly underpredicted the experimental data in the later  
371 stages. The lack of macroscale deformation is likely contributing to the difference in the later  
372 stages of drying. When drying at 64°C, the rupture threshold is reached and the downscaling  
373 model estimated the ICW content from the change in cell size. This resulted in a diffusivity  
374 range of  $4.78 \times 10^{-10} - 2.58 \times 10^{-10}$  m<sup>2</sup>/s which is similar to the control model, Figure 7. The  
375 resulting diffusivities fell in the typically range of  $10^{-11}$  and  $10^{-9}$  m<sup>2</sup>/s for food material  
376 (Panagiotou et al., 2004; Saravacos & Maroulis, 2001).

377 **[Figure 6 can be placed here]**

378 **[Figure 7 can be placed here]**

379

### 380 3.3 *Macroscale – Temperature evolution*

381 The downscaling model accurately predicted the average sample temperature for drying  
382 at both 47°C and 64°C, with MREs of 0.53% and 0.78% respectively, Figure 8. Similarly, the  
383 control model achieved MREs of 0.46% and 0.89% respectively when compared to the  
384 experimental results, Figure 8. There is minimal difference between the downscaling model  
385 and the control model at both drying temperatures. This is because no properties within the  
386 heat transfer are in terms of ICW, rather just macroscale moisture. However, both the multiscale  
387 downscaling model and the control model significantly overpredicted the experimental data in  
388 the early stages of drying. It should be noted, the experimental data is of a sample under  
389 convective drying with perpendicular airflow. Due to the perpendicular airflow, a large ‘dead  
390 zone’ existed behind the trailing edge of the sample during drying and thus, the heat transfer  
391 coefficient heavily effected in this region of the sample (Khan et al., 2020). As a result, the  
392 temporally and spatially constant heat transfer coefficient contributed heavily to the initial  
393 overprediction.

394

**[Figure 8 can be placed here]**

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396 *3.4 Intracellular water transport*

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The spatial moisture profile at macroscale and the three locations of interest for drying at 47°C and 64°C can be seen in Figure 9 and Figure 10 respectively, in which the dashed circle denotes the original cell size. It should be noted, the moisture profile shown in Figure 9 and Figure 10 are at the exchange steps. As expected, each point of interest has its own temporal moisture profile corresponding to the macroscale conditions occurring at that particular point. This is unlike the existing microscale models in literature which utilize a single domain to characterize the sample transport (Rahman et al., 2018). As the moisture profile develops within the sample, the intracellular space at the outside point (P3), dries first. As expected, in the later stages, the intracellular space at P3 is entirely dry whereas the intracellular space at P1 only starts to dry. This trend observed in both drying temperatures. Additionally, due to the temporal coupling and the size of the exchange time step utilized, there can be a delay in the moisture transfer between the end of the microscale and the initialization of the microscale. The microscale is only simulated for 5 seconds after an exchange step and the data exchange only occurs every 600 seconds (10 mins). This creates a time skip of 595 seconds. Evidence of this delay can be seen in the microscale at P3 at 100 minutes in Figure 9.

412

**[Figure 9 can be placed here]**

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**[Figure 10 can be placed here]**

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The average moisture content of the intracellular spaces at each point of interest and the flux through the cell walls can be found in Figure 11 (a) and (b) respectively. Throughout drying, the flux through the cell wall at P3 (point closest to the exterior wall) is significantly

418 larger than the flux at P2 and P1, especially in the middle portion of drying. This trend is  
419 consistent for both drying temperatures. Additionally, the flux through the cell walls at each  
420 point exhibit peaks at different stages of drying and as expected drying at higher temperatures  
421 resulted in additional flux through the cell wall. P3 near the outside wall peaks first at 110 mins  
422 for drying at 64°C with a magnitude of  $-4.33 \times 10^{-7}$  mol/s. For drying at 47°C, P3's peak occurs  
423 later at 170 mins. The different shrinkage velocity for the two drying temperatures resulted in  
424 slightly different final cell sizes of 119  $\mu\text{m}$  for drying at 47°C and 109  $\mu\text{m}$  for 64°C following  
425 the trend uncovered in the X-ray  $\mu\text{CT}$  investigation. It should be noted, the shrinkage velocity  
426 approach results in ideal shrinkage.

427 **[Figure 11 can be placed here]**

428

429

### 430 3.5 *Sensitivity Investigation*

431 A sensitivity analysis was conducted on a parameter with uncertainty, specifically on  
432 cell wall permeability, which was considered constant. Literature has demonstrate when drying  
433 at 70°C the average cell wall thickness of granny smith apples changes from 9.312 $\mu\text{m}$  when  
434 fresh to 4.685 $\mu\text{m}$  when dried (Joardder et al., 2015). This change mainly occurs when the FW  
435 content reduces surrounding an intracellular space and ICW begins to transport through the cell  
436 wall. Permeability is the diffusivity of a cell wall over its width, hence as diffusivity and the  
437 wall thickness changes uncertainty exists within permeability. To evaluate the sensitivity of the  
438 downscaling model, the cell permeability was investigated by varying the utilized value by  
439  $\pm 10\%$  while drying at 47°C. This parameter was investigated in relation to the average  
440 macroscale moisture content (kg/kg dry bases), the microscale concentrations (mol) and the  
441 cell wall flux (mol/s).

442 The downscaling model's sensitivity to cell wall permeability can be seen in Figure 12.  
443 The macroscale moisture content and the microscale ICW content were not sensitive to the  
444 change in cell wall permeability with their magnitudes and trend not changing. However, as  
445 permeability is key in the cellular boundary condition, the changes in permeability do influence  
446 the magnitude of the cell wall flux. Plus 10% results in an increase to the magnitude of the flux  
447 whereas minus 10% decrease the flux through the cell wall. The difference is significant and  
448 clearly evidenced for P3 but much smaller for P1 and P2.

449 **[Figure 12 can be placed here]**

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451

#### 452 **4. Discussion**

453 The downscaling model provides new insight into the transport of ICW while  
454 accurately predicting drying at 47°C and 64°C. When drying at lower temperatures (47°C) the  
455 consideration of a constant ICW with the upscaled diffusivity [Equation (16)] creates a fairly  
456 constant diffusivity which was sufficient to accurately predict drying. This finding is consistent  
457 with pre-existing literature (Defraeye & Verboven, 2017; Welsh et al., 2023). When drying at  
458 medium temperatures (64°C), the multiscale downscaling model accurately predicted drying,  
459 suggesting that ICW estimation based on cell size changes is suitable to utilize the upscaled  
460 diffusivity property with the rupture threshold. This technique enhances the generalizability of  
461 the modeling approach and differs from previously published work (Welsh et al., 2023).  
462 Notably, the shrinkage velocity approach, based on X-ray  $\mu$ CT data, was employed to estimate  
463 cell size changes. The model could be further refined by incorporating a physics-based  
464 approach to cellular deformation, such as van der Sman et al. (2024)'s recent work on pore  
465 development in viscoelastic foods during drying.

466           The microscales at the three points of interest provided interesting insight, Figure 9 to  
467 Figure 10. Each produced spatially dependent microscales which is different to the previously  
468 published microscale models where a single microscale is used to predict the material's  
469 evolution (Rahman et al., 2018). Specifically, the flux through the cell wall is very unique for  
470 each intracellular space at each point of interest. The microscale near the outside edge (P3)  
471 experienced significantly more flux throughout the entire duration of drying. The magnitude  
472 of the flux also depended on the drying air temperature and is sensitive to the magnitude of the  
473 cell wall permeability. Literature has demonstrated when drying at medium temperature (above  
474 50°C), progressive and ultimate cell rupturing would have occurred (Khan et al., 2018). It is  
475 theorized, due to the convective heating profile, when the surface reaches 50°C the intracellular  
476 space near the surface lose their rigidity and tend to become droopy, hence nearing their  
477 rupturing limit (Khan et al., 2018). The intracellular spaces lose turgor pressure and tend to fail  
478 due to the imposed thermal stress of continuous penetration of heat energy. Though past  
479 research on cell rupturing has been conducted using nuclear magnetic resonance which is only  
480 able to produce results of the entire sample (domain averages) and does not give insight into  
481 the spatial propagation of the rupturing mechanism. Further investigation is needed in the  
482 future, but the cell wall flux may provide valuable insight in predicting when and/or where cell  
483 rupturing spatially propagates within PBFMs.

484           One key component of any multiscale model is its temporal coupling. Previous works  
485 within the field (Rahman et al., 2018) have only utilized multiscale modeling to upscale  
486 properties. Such an approach produces a property, often a homogenized property, through  
487 computing the microscale in a steady state condition for specific discrete points of interest (i.e.  
488 when a cell has a moisture content of 3 kg/kg dry bases). However, as the current study is a  
489 multiscale downscaling model, the microscale evolves with time and hence a temporal coupling  
490 is required. Specifically, the HMM approach was utilized. This approach saves on

491 computational cost but does not simulate the microscale for the entire simulation (as described  
492 in 2.3.1 Macro-micro temporal coupling). Instead, the approach just computes the microscale  
493 for a small period of time after the exchange point. Hence, there is a portion of time (9.16 mins)  
494 which is skipped in between the exchange steps. This saves on computational cost however it  
495 can create a delayed effect on the transport, which will not always be desired. Depending on  
496 the purpose of the model and the available computational resources, this skipped time can be  
497 either reduced or increased. Alternatively, temporal couplings like the continuous HMM  
498 simulate the microscale across the entire time period and can be employed based on the model's  
499 purpose.

500

## 501 **5. Conclusion**

502 The multiscale model accurately described the drying process at both low and medium  
503 temperatures (47°C and 64°C). For drying at 47°C the model achieved MREs of 2.4% and  
504 0.53% for average macroscale moisture content and temperature respectively and for drying at  
505 64°C the model achieved MREs of 5.32% and 0.78% for average macroscale moisture content  
506 and temperature, respectively. Furthermore, the integration of downscaling coupling in the  
507 model provided valuable insights into the microscale evolution during drying. When drying at  
508 medium temperatures, the rupturing threshold was reached and the ICW of the sample was  
509 predicted considering the changes in the cells size. To demonstrate the capacity of the approach,  
510 microscales at three points of interest were investigated. Each microscale evolved differently  
511 depending on the applied drying conditions and their spatial location within the domain. The  
512 ICW trend through the cell wall provided important insight into what the cell wall experiences  
513 at different locations and was also unique for each microscale location and drying temperature.  
514 Additionally, the convective drying temperature significantly influenced the magnitude of the

515 flux the cell wall experienced due to the moisture migration. Further investigating is required  
516 but such a multiscale approach could be utilized to investigate how cell rupturing propagates  
517 within a domain. The model could also be refined to calculate cellular deformation through a  
518 more mathematical approach with deformation on both spatial scales coupled with some  
519 consideration of cell rupturing. The multiscale downscaling approach, coupled with Welsh et  
520 al. (2023) upscaling technique will facilitate the construct of an in-depth concurrent multiscale  
521 model being developed in the future.

## 522 **Acknowledgements**

523 This research work is supported by Australian Research council (DP220103668). The  
524 authors would like to acknowledge Dr. M.U.H Joardder for his suggestion in relation to the  
525 cellular transport. Moreover, the authors also wish to express their gratitude to the KG-Q  
526 Research Facility, QUT for allowing access to the X-ray  $\mu$ CT equipment.

527

## 528 6. References

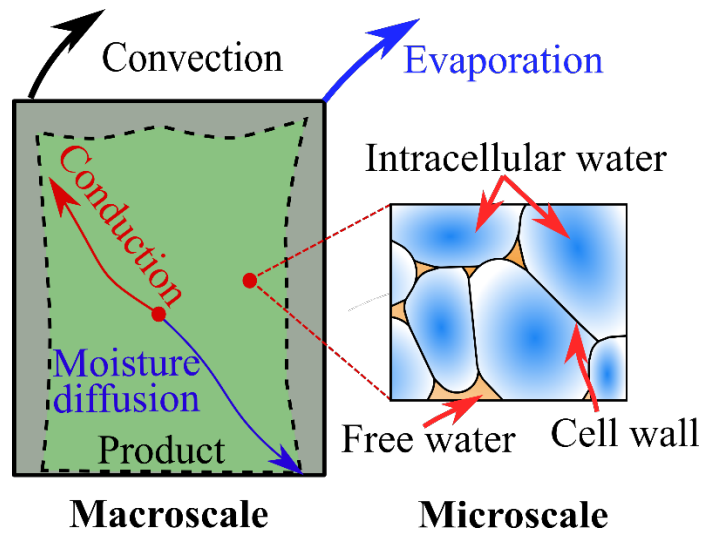
- 529 Asabe, J., & Home, A. American society of agricultural and biological engineers.
- 530 Auriault, J.-L., Boutin, C., & Geindreau, C. (2010). *Homogenization of coupled phenomena in*  
531 *heterogenous media* (Vol. 149). John Wiley & Sons.
- 532 Białobrzewski, I. (2006). Simulation of changes in the density of an apple slab during drying.  
533 *International Communications in Heat and Mass Transfer*, 33(7), 880-888.  
534 <https://doi.org/https://doi.org/10.1016/j.icheatmasstransfer.2006.02.017>
- 535 Carr, E. J., & Turner, I. W. (2014). Two-scale computational modelling of water flow in  
536 unsaturated soils containing irregular-shaped inclusions. *International Journal for*  
537 *Numerical Methods in Engineering*, 98, 157-173.  
538 <https://doi.org/https://doi.org/10.1002/nme.4625>
- 539 Carr, E. J., Turner, I. W., & Perré, P. (2013). A dual-scale modeling approach for drying  
540 hygroscopic porous media. *Multiscale Modeling & Simulation*, 11, 362-384.  
541 <https://doi.org/10.1137/120873005>
- 542 Cengel, Y. (2003). *Heat, Transfer Mass: A practical approach*. Mc-Graw Hill Education,  
543 Columbus, GA, USA.
- 544 Cengel, Y. A., & Boles, M. A. (2002). *Thermodynamics: an engineering approach. Sea, 1000,*  
545 8862.
- 546 Defraeye, T., & Verboven, P. (2017). Convective drying of fruit: Role and impact of moisture  
547 transport properties in modelling. *Journal of Food Engineering*, 193, 95-107.  
548 <https://doi.org/http://dx.doi.org/10.1016/j.jfoodeng.2016.08.013>
- 549 Fanta, S. W., Abera, M. K., Aregawi, W. A., Ho, Q. T., Verboven, P., Carmeliet, J., & Nicolai,  
550 B. M. (2014). Microscale modeling of coupled water transport and mechanical  
551 deformation of fruit tissue during dehydration. *Journal of Food Engineering*, 124, 86-  
552 96. <https://doi.org/10.1016/j.jfoodeng.2013.10.007>
- 553 Fanta, S. W., Abera, M. K., Ho, Q. T., Verboven, P., Carmeliet, J., & Nicolai, B. M. (2013).  
554 Microscale modeling of water transport in fruit tissue. *Journal of Food Engineering*,  
555 118, 229-237. <https://doi.org/https://doi.org/10.1016/j.jfoodeng.2013.04.003>
- 556 Ferrando, M., & Spiess, W. E. L. (2001). Cellular response of plant tissue during the osmotic  
557 treatment with sucrose, maltose, and trehalose solutions. *Journal of Food Engineering*,  
558 49(2), 115-127. [https://doi.org/https://doi.org/10.1016/S0260-8774\(00\)00218-1](https://doi.org/https://doi.org/10.1016/S0260-8774(00)00218-1)

- 559 Golestani, R., Raisi, A., & Aroujalian, A. (2013). Mathematical modeling on air drying of  
560 apples considering shrinkage and variable diffusion coefficient. *Drying Technology*,  
561 *31*, 40-51. <https://doi.org/https://doi.org/10.1080/07373937.2012.714826>
- 562 Joardder, M. U., Brown, R. J., Kumar, C., & Karim, M. (2015). Effect of cell wall properties  
563 on porosity and shrinkage of dried apple. *International Journal of Food Properties*,  
564 *18*(10), 2327-2337. <https://doi.org/https://doi.org/10.1080/10942912.2014.980945>
- 565 Karunasena, H. C. P., Gu, Y. T., Brown, R. J., & Senadeera, W. (2015). Numerical  
566 investigation of plant tissue porosity and its influence on cellular level shrinkage during  
567 drying. *Biosystems Engineering*, *132*(Supplement C), 71-87.  
568 <https://doi.org/https://doi.org/10.1016/j.biosystemseng.2015.02.002>
- 569 Khan, M. I. H., Farrell, T., Nagy, S. A., & Karim, M. A. (2018). Fundamental Understanding  
570 of Cellular Water Transport Process in Bio-Food Material during Drying. *Scientific*  
571 *Reports*, *8*(1), 15191. <https://doi.org/https://doi.org/10.1038/s41598-018-33159-7>
- 572 Khan, M. I. H., Wellard, R. M., Nagy, S. A., Joardder, M. U. H., & Karim, M. A. (2016).  
573 Investigation of bound and free water in plant-based food material using NMR T2  
574 relaxometry. *Innovative Food Science & Emerging Technologies*, *38*, 252-261.  
575 <https://doi.org/https://doi.org/10.1016/j.ifset.2016.10.015>
- 576 Khan, M. Imran H., Welsh, Z., Gu, Y., Karim, M. A., & Bhandari, B. (2020). Modelling of  
577 simultaneous heat and mass transfer considering the spatial distribution of air velocity  
578 during intermittent microwave convective drying. *International Journal of Heat and*  
579 *Mass Transfer*, *153*, 119668.  
580 <https://doi.org/https://doi.org/10.1016/j.ijheatmasstransfer.2020.119668>
- 581 Mattea, M., Urbicain, M. J., & Rotstein, E. (1986). Prediction of Thermal Conductivity of  
582 Vegetable Foods by the Effective Medium Theory. *Journal of Food Science*, *51*(1),  
583 113-115. <https://doi.org/https://doi.org/10.1111/j.1365-2621.1986.tb10848.x>
- 584 Mavroudis, N. E., Gekas, V., & Sjöholm, I. (1998). Osmotic dehydration of apples—effects of  
585 agitation and raw material characteristics. *Journal of Food Engineering*, *35*(2), 191-  
586 209. [https://doi.org/https://doi.org/10.1016/S0260-8774\(98\)00015-6](https://doi.org/https://doi.org/10.1016/S0260-8774(98)00015-6)
- 587 Moshelion, M., Moran, N., & Chaumont, F. (2004). Dynamic changes in the osmotic water  
588 permeability of protoplast plasma membrane. *Plant Physiol*, *135*(4), 2301-2317.  
589 <https://doi.org/https://doi.org/10.1104/pp.104.043000>
- 590 Pace, E. L. (1962). Scientific foundations of vacuum technique (Dushman, Saul). *Journal of*  
591 *Chemical Education*, *39*(8), A606. <https://doi.org/https://doi.org/10.1021/ed039pA606>

- 592 Panagiotou, N., Krokida, M., Maroulis, Z., & Saravacos, G. (2004). Moisture diffusivity:  
593 literature data compilation for foodstuffs. *International Journal of Food Properties*,  
594 7(2), 273-299. <https://doi.org/https://doi.org/10.1081/JFP-120030038>
- 595 Pavliotis, G. A., & Stuart, A. (2008). *Multiscale methods: averaging and homogenization*.  
596 Springer Science & Business Media.
- 597 Perré, P. (2007). Multiscale aspects of heat and mass transfer during drying. *Transport in*  
598 *Porous Media*, 66, 59-76. <https://doi.org/https://doi.org/10.1007/s11242-006-9022-2>
- 599 Perré, P. (2019). Coupled heat and mass transfer in biosourced porous media without local  
600 equilibrium: A macroscopic formulation tailored to computational simulation.  
601 *International Journal of Heat and Mass Transfer*, 140, 717-730.  
602 <https://doi.org/https://doi.org/10.1016/j.ijheatmasstransfer.2019.06.043>
- 603 Prawiranto, K., Carmeliet, J., & Defraeye, T. (2020). Identifying in silico how microstructural  
604 changes in cellular fruit affect the drying kinetics [10.1039/D0SM00749H]. *Soft*  
605 *Matter*, 16(43), 9929-9945. <https://doi.org/10.1039/D0SM00749H>
- 606 Prawiranto, K., Defraeye, T., Derome, D., Verboven, P., Nicolai, B., & Carmeliet, J. (2018).  
607 New insights into the apple fruit dehydration process at the cellular scale by 3D  
608 continuum modeling. *Journal of Food Engineering*, 239, 52-63.  
609 <https://doi.org/https://doi.org/10.1016/j.jfoodeng.2018.06.023>
- 610 Qiu, S., Xu, S., Rao, B., Mujumdar, A. S., & Xu, P. (2022). A multi-scale model for  
611 impingement drying of porous slab. *Journal of Food Engineering*, 335, 111194.  
612 <https://doi.org/https://doi.org/10.1016/j.jfoodeng.2022.111194>
- 613 Rahman, M. M., Kumar, C., Joardder, M. U. H., & Karim, M. A. (2018). A micro-level  
614 transport model for plant-based food materials during drying. *Chemical Engineering*  
615 *Science*, 187, 1-15. <https://doi.org/10.1016/j.ces.2018.04.060>
- 616 Ratti, C., Crapiste, G., & Rotstein, E. (1989). A new water sorption equilibrium expression for  
617 solid foods based on thermodynamic considerations. *Journal of Food Science*, 54(3),  
618 738-742. <https://doi.org/https://doi.org/10.1111/j.1365-2621.1989.tb04693.x>
- 619 Saravacos, G. D., & Maroulis, Z. B. (2001). *Transport properties of foods*. CRC Press.
- 620 Tan, L., Yuan, Y., Zhao, Z., Xu, Y., & Yuan, Y. (2023). Insights in mechanism of drying  
621 shrinkage by pore-scale modeling of heat-moisture and stress-strain distribution for  
622 high-moisture porous media. *International Journal of Thermal Sciences*, 188, 108226.  
623 <https://doi.org/https://doi.org/10.1016/j.ijthermalsci.2023.108226>

- 624 Tuly, S. S., Joardder, M. U. H., Welsh, Z. G., & Karim, A. (2023). Mathematical Modelling of  
625 Heat and Mass Transfer during Jackfruit Drying Considering Shrinkage. *Energies*,  
626 *16*(11), 4461. <https://www.mdpi.com/1996-1073/16/11/4461>
- 627 van der Sman, R., Curatolo, M., & Teresi, L. (2024). Pore development in viscoelastic foods  
628 during drying [10.1039/D4SM00201F]. *Soft Matter*, *20*(26), 5183-5194.  
629 <https://doi.org/10.1039/D4SM00201F>
- 630 Vega-Mercado, H., Marcela Góngora-Nieto, M., & Barbosa-Cánovas, G. V. (2001). Advances  
631 in dehydration of foods. *Journal of Food Engineering*, *49*(4), 271-289.  
632 [https://doi.org/https://doi.org/10.1016/S0260-8774\(00\)00224-7](https://doi.org/https://doi.org/10.1016/S0260-8774(00)00224-7)
- 633 Weligama Thuppahige, V. T., Welsh, Z. G., Joardder, M., & Karim, A. (2024). Recent  
634 advances in determining the cellular-level property evolutions of plant-based food  
635 materials during drying. *Trends in Food Science & Technology*, *143*, 104291.  
636 <https://doi.org/https://doi.org/10.1016/j.tifs.2023.104291>
- 637 Welsh, Z., Simpson, M. J., Khan, M. I. H., & Karim, M. (2018). Multiscale Modeling for Food  
638 Drying: State of the Art. *Comprehensive Reviews in Food Science and Food Safety*,  
639 *17*(5), 1293-1308. <https://doi.org/https://doi.org/10.1111/1541-4337.12380>
- 640 Welsh, Z. G., Khan, M. I. H., & Karim, M. (2021a). Multiscale modeling for food drying: A  
641 homogenized diffusion approach. *Journal of Food Engineering*, *292*, 110252.  
642 <https://doi.org/https://doi.org/10.1016/j.jfoodeng.2020.110252>
- 643 Welsh, Z. G., Simpson, M. J., Khan, M. I. H., & Karim, A. (2021b). A multiscale approach to  
644 estimate the cellular diffusivity during food drying. *Biosystems Engineering*, *212*, 273-  
645 289. <https://doi.org/https://doi.org/10.1016/j.biosystemseng.2021.10.017>
- 646 Welsh, Z. G., Simpson, M. J., Khan, M. I. H., & Karim, M. A. (2023). Generalized moisture  
647 diffusivity for food drying through multiscale modeling. *Journal of Food Engineering*,  
648 *340*, 111309. <https://doi.org/https://doi.org/10.1016/j.jfoodeng.2022.111309>
- 649 Whitaker, S. (1977). Simultaneous Heat, Mass, and Momentum Transfer in Porous Media: A  
650 Theory of Drying. *Advances in Heat Transfer*, *13*, 119-203.  
651 [https://doi.org/http://dx.doi.org/10.1016/S0065-2717\(08\)70223-5](https://doi.org/http://dx.doi.org/10.1016/S0065-2717(08)70223-5)
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654 **Figures:**



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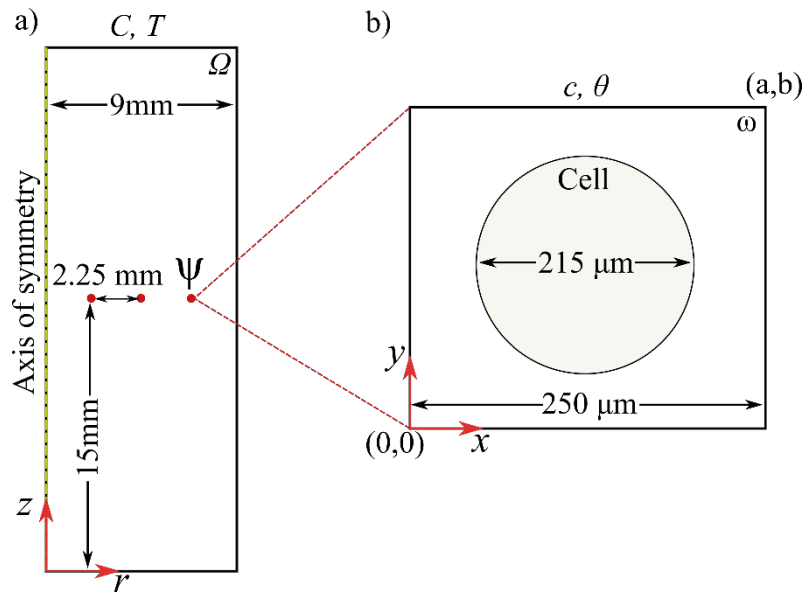
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Figure 1. Cellular structure of plant-based food material

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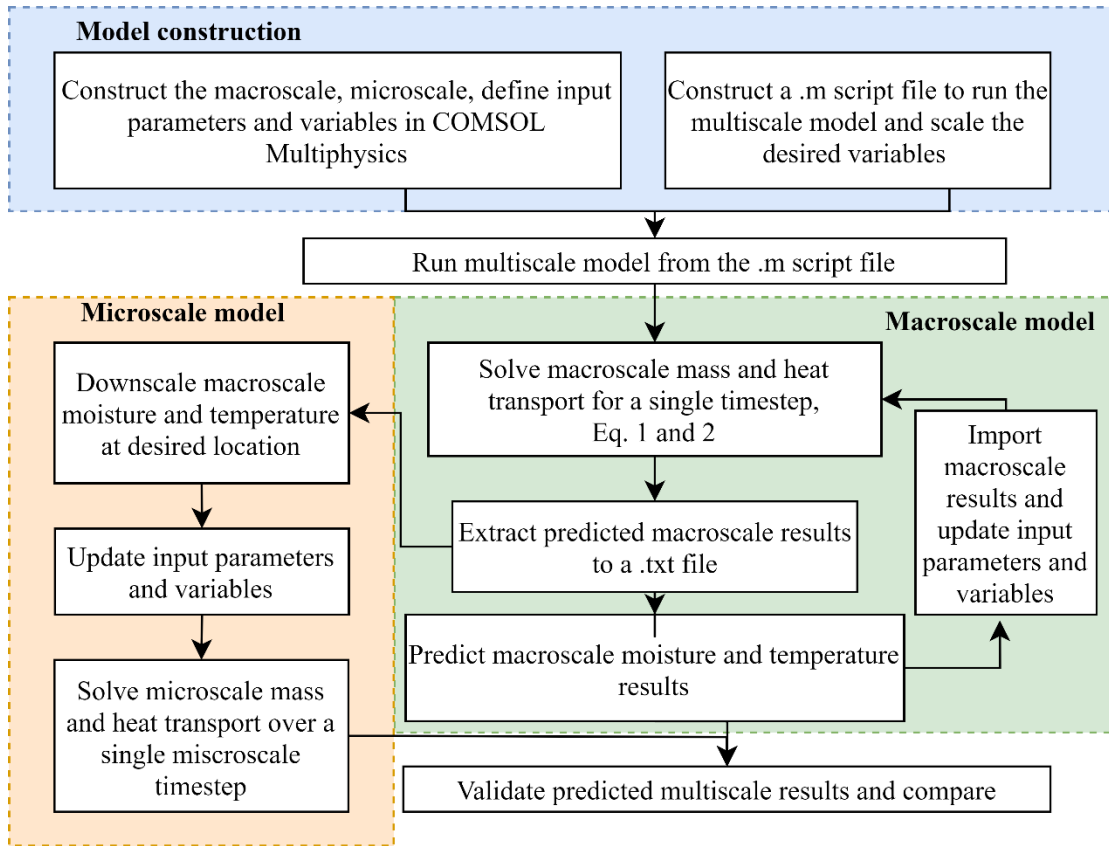
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Figure 2. Domains, a) axisymmetric macroscale domain with the three locations of interest marked by red dots, and b) microscale domain.

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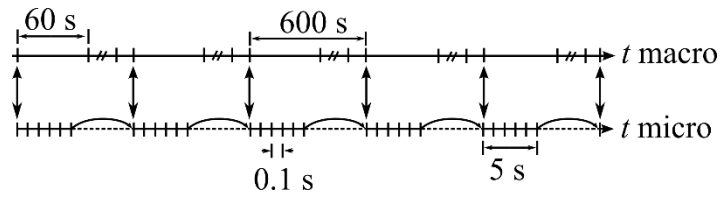
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Figure 3. Simulation Methodology

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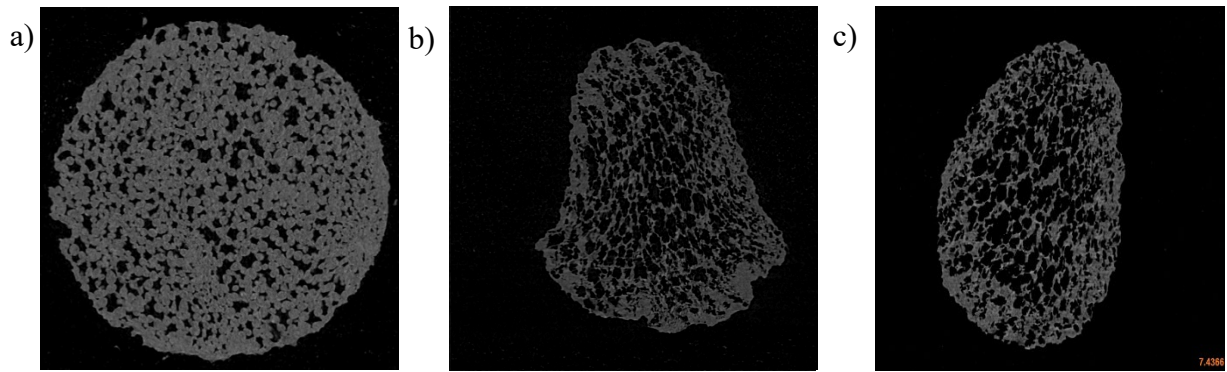
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Figure 4. Multiscale temporal coupling.

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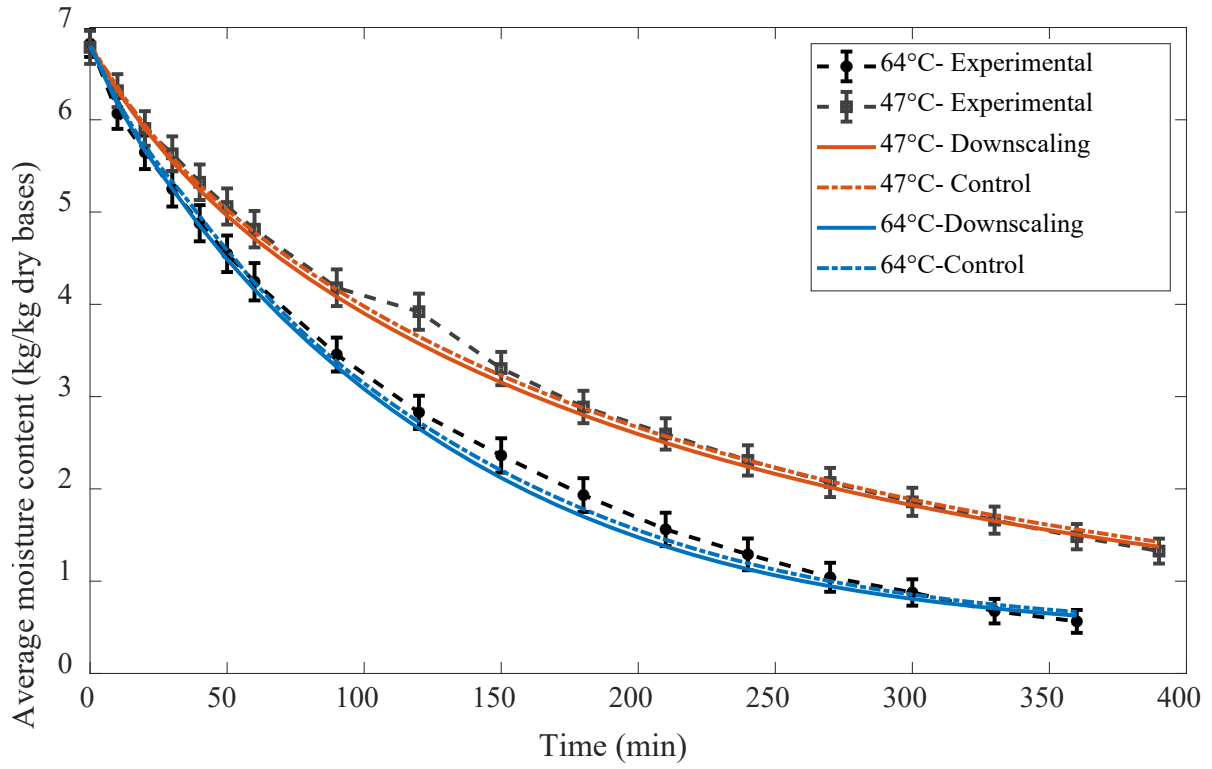
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Figure 5. X-ray  $\mu$ CT images of, (a) fresh, (b) dried at 45°C at 0.8 kg/kg dry bases, (c) dried at 60°C at 1.23 kg/kg dry bases.



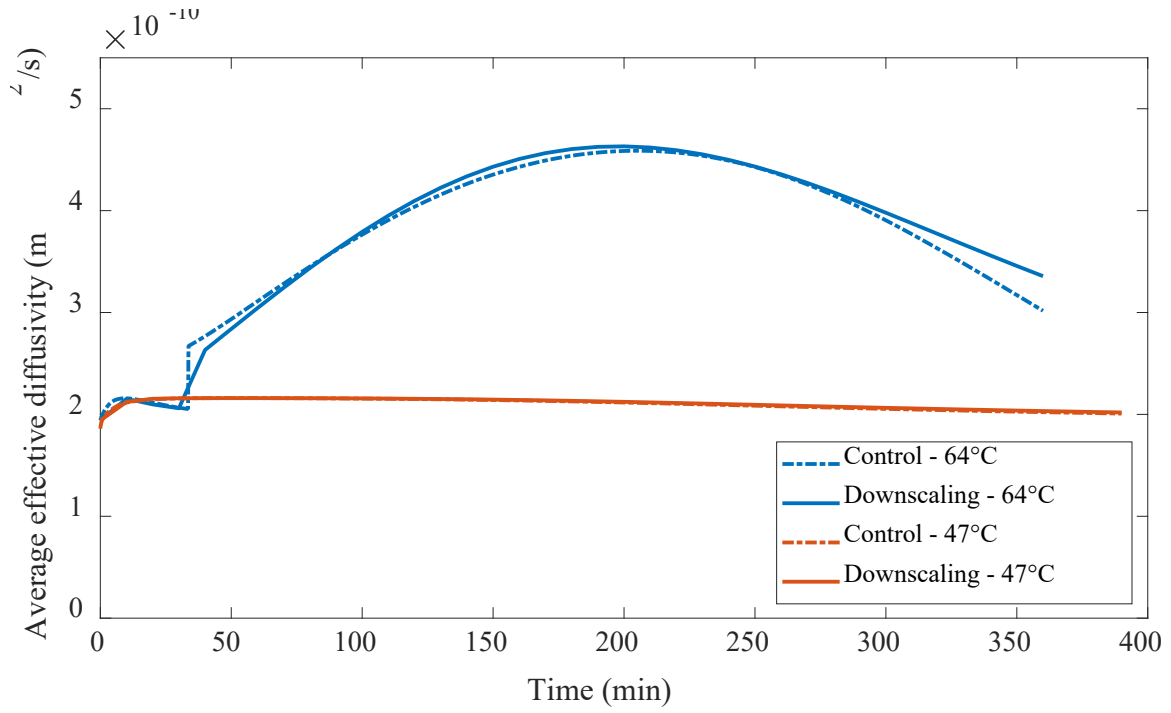
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Figure 6. Average moisture content (kg/kg dry bases) at low and medium temperature drying.

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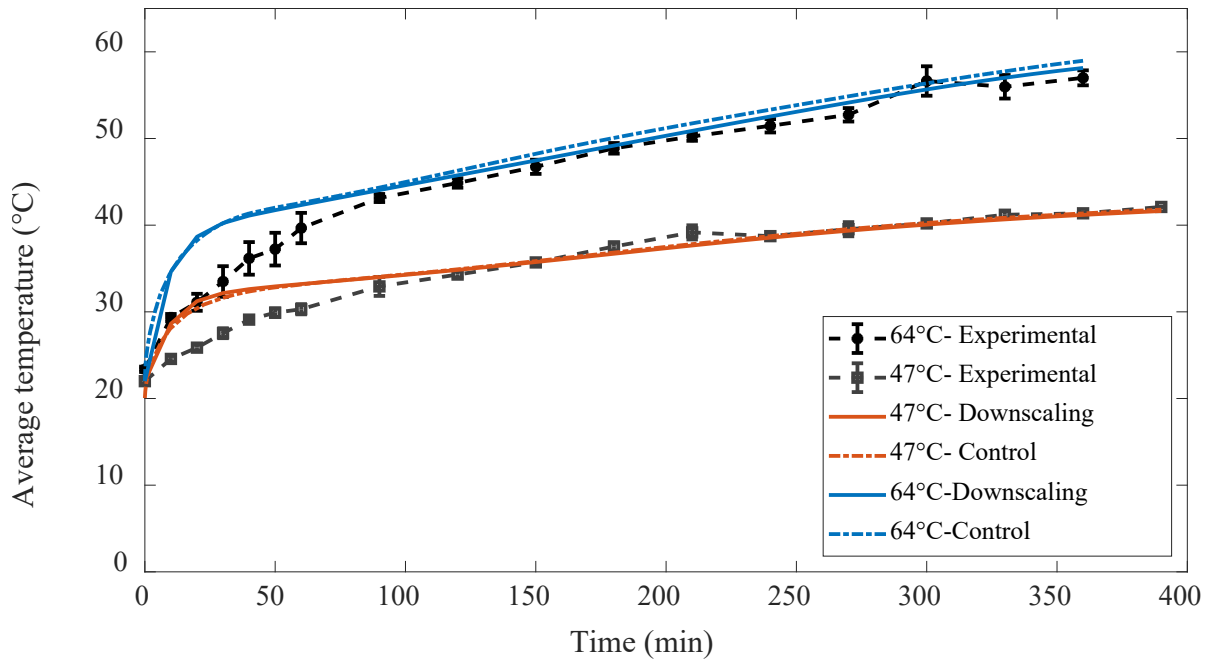


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Figure 7. Average macroscale diffusivity (m<sup>2</sup>/s) for both drying temperatures.

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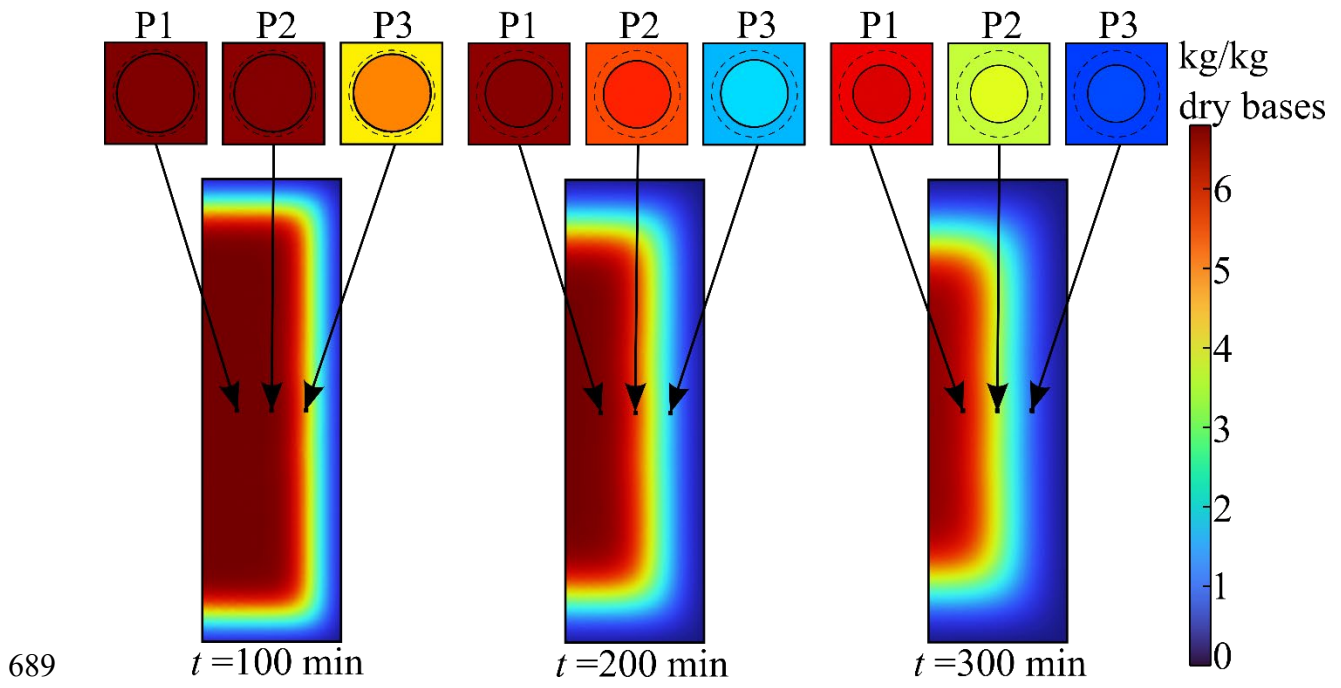


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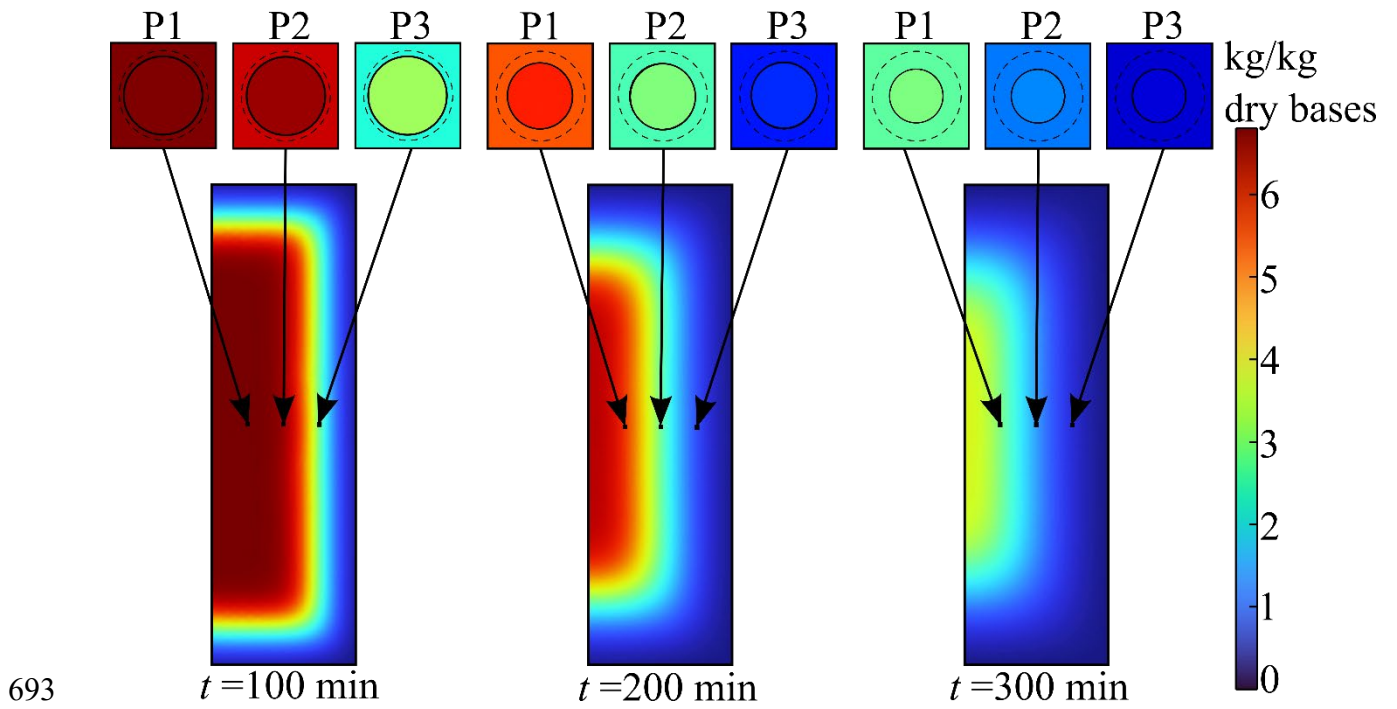
Figure 8. Average surface temperature (°C) for both drying temperatures.

688



690 Figure 9. Multiscale moisture distribution (kg/kg dry bases) for drying at 47°C. Original cell  
691 geometry visually represented by the dashed circle.

692



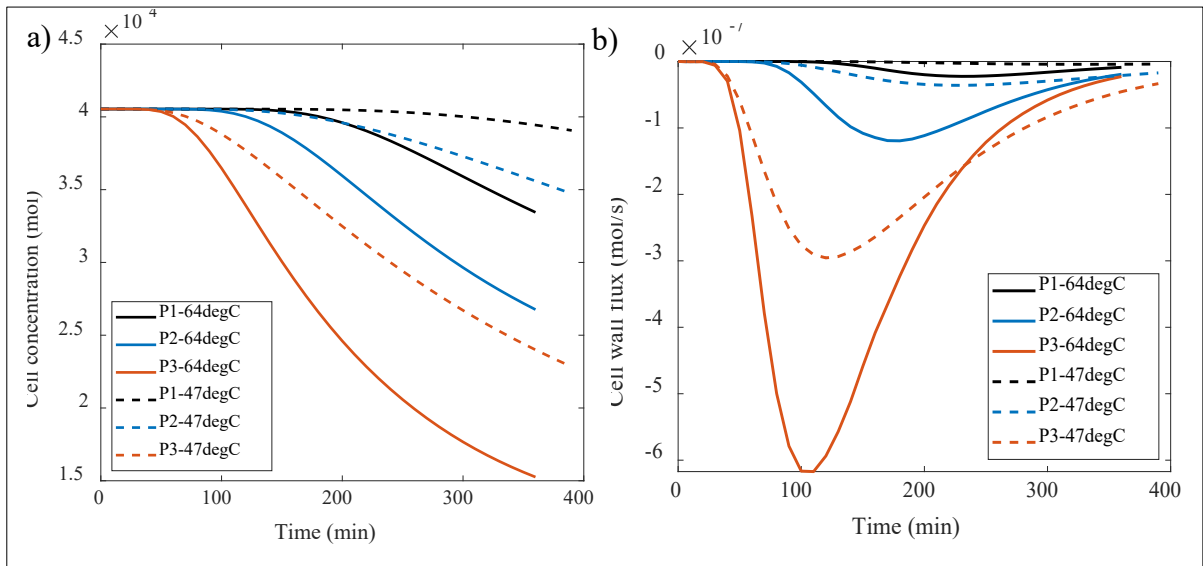
693

694 Figure 10. Multiscale moisture distribution (kg/kg dry bases) for drying at 64°C. Original cell

695

geometry visually represented by the dashed circle.

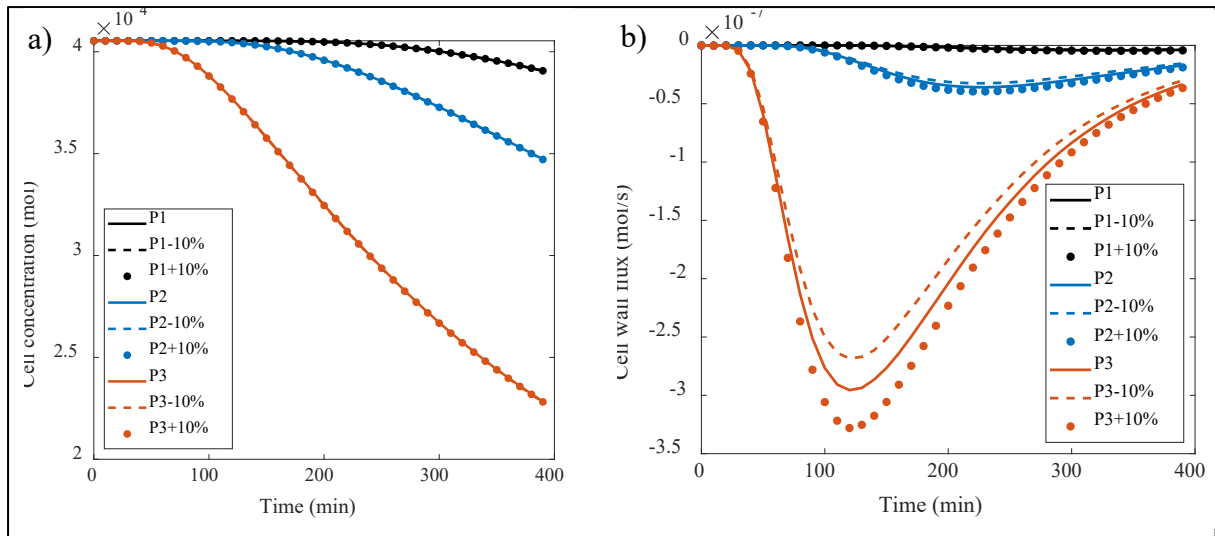
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697

698 Figure 11. Intracellular water evolution for both drying temperatures, (a) concentration within  
699 each intracellular space at the points of interest (mol), (b) Flux through the cell wall (mol/s) at  
700 the points of interest.

701



702

703 Figure 12. Sensitivity investigation for the cell wall permeability  $\pm 10\%$ , a) moisture  
704 concentration (mol), at each point of interest b) cell wall flux (mol/s) at each point of interest.

705

706 **Tables:**

707 Table 1. Input parameters

Parameter	Value (unit)	Reference
Density of apple, $\rho$	837 (kg m <sup>-3</sup> )	Mavroudis et al. (1998)
Density of water, $\rho_w$	995 (kg m <sup>-3</sup> )	Cengel (2003)
Specific heat of apple, $c_p$	1000(1.4+3.22 $M_{wb}$ ) (J kg <sup>-1</sup> K <sup>-1</sup> )	Białobrzewski (2006)
Thermal conductivity of apple, $k$	0.49-0.443exp(-0.206 $M_{wb}$ ) (W/(m·K))	Mattea et al. (1986)
Initial moisture content, $M_0$	6.8 (kg kg <sup>-1</sup> dry bases)	This study
Initial sample temperature, $T_0$	296 (K)	This study
Equilibrium moisture content, $M_e$	0.2 (kg kg <sup>-1</sup> dry bases)	Białobrzewski (2006)
Initial wet basis moisture content, $M_{0,wb}$	0.86 (kg kg <sup>-1</sup> wet bases)	This study
Latent heat of evaporation, $h_{fg}$	2358600 (J kg <sup>-1</sup> )	Cengel (2003)
Universal gas constant, $R$	8.314 (J mol <sup>-1</sup> K <sup>-1</sup> )	Cengel and Boles (2002)
Molar mass of water, $M_w$	0.018016 (kg mol <sup>-1</sup> )	Cengel and Boles (2002)
Partial vapor pressure, $p_{v\ air}$	1250 (Pa)	(Welsh et al., 2023)
Heat transfer coefficient, $h_T$	13.2 (W m <sup>-2</sup> K <sup>-1</sup> )	(Welsh et al., 2023)
Mass transfer coefficient, $h_m$	0.00381 (m s <sup>-1</sup> )	(Welsh et al., 2023)
Cell wall permeability	2 x 10 <sup>-5</sup> (m s <sup>-1</sup> )	(Ferrando & Spiess, 2001; Moshelion et al., 2004)

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